Isolation of Marine Organisms and their Antifungal Studies

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doi: http://dx.doi.org/10.13005/bbra/1429

(Received: 15 August 2014; accepted: 10 October 2014)

Marine organisms are a good source of new secondary metabolites that possess many biological activities including antibacterial, antifungal, anticancer, insecticidal and enzyme inhibition. Actinomycetes are known to produce bioactive substances, especially antibiotics that are effective against phytopathogenic fungi. Biocontrol with beneficial bacteria is one promising alternative to fungicides. Hydrolases such as chitinase contribute to degradation of fungal cell walls . Chitin is the second most abundant polysaccharide in nature and a major component of fungal walls, insect exoskeletons and crustacean shells. Chitinase secreted by a BCA is likely to be effective against pathogenic fungi, the cell walls of which are mainly Chitin. Several species have been isolated and screened from the soil in the past decades. Many of these secondary metabolites possess biological activities and have the potential to be developed as therapeutic agents. Bio-active compounds from marine organisms possess distinct chemical structures that are used in the synthesis of new drugs that could be used against pathogens. In this paper isolation of marine organism samples are collected at different marine environments and habitats and their antifungal studies are described.

Key words: Secondary metabolites, Bio-active compounds, Antifungal activity.

Certain marine organisms are best known for their ability to produce antibiotics and are gram positive bacteria which comprise a group of branching unicellular microorganisms. They produce branching mycelium which may be of two kinds viz., substrate mycelium and aerial mycelium. Among several marine organisms, many actinomycetes and the Streptomycetes are the dominant. The non-Streptomycetes, are called rare actinomycetes, comprising approximately 100 genera. Members of the actinomycetes, which live in marine environment, are poorly understood and

only few reports are available pertaining to actinomycetes from mangroves¹⁻². Terrestrial actinomycetes are soil organisms which have characteristics common to bacterial and fungi and yet possess sufficient distinctive features to delimit them into a distinct category. . On agar plates, they can easily be distinguished from true bacteria. Unlike slimy distinct colonies of true bacteria which grow quickly. Actinomycetes colonies appear slowly, show powdery consistency and stick firmly to agar surface. A closer look at a colony under the compound microscope reveals slender unicellular branched mycelium (diameter of the hyphaerarely exceeding one micron) forming asexual spores for propagation. The above marine organisms are proved to be producing bio active substances. Literature studies clearly indicate antibiotics are effective against phytopathogenic fungi³⁻⁴.

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MATERIALSAND METHOD

Identification Methods

Numerical Taxonomic Approach

Numerical taxonomy involves examining many strains for a large number of characters prior to assigning the test organism to a cluster based on shared features. The numerically defined taxa are polythetic; so, no single property is either indispensable or sufficient to entitle an organism for membership of a group. Once classification has been achieved, cluster-specific or predictive characters can be selected for identification⁵

Numerical taxonomy was first applied to Streptomyces by⁶.The numerical taxonomic study of the genus Streptomyces by Williams et al. (1983).⁷⁻⁸ Involves determination of 139 unit characters for 394 type cultures of Streptomyces; and the former co-efficient is being used to define the clusters. His study includes 23 major, 20 minor and 25 single member clusters. Thus, numerical taxonomy provides us with an invaluable framework for streptomyces taxonomy, including identification of species⁹.

Secondary metabolites

The marine habitat is particularly important for discovery of novel bioactive products. Although major attention has been dedicated to the study of shallow and deep water sediments, microbial associates of marine molluscs have proven to be a rich source of biologically active substances such as antimicrobial and antitumor compounds¹⁰⁻¹¹. Shellfish molluscs are known to be natural accumulators of microorganisms due to their filter-feeding habit. Their internal tissues are enriched by nutrient compounds and adhesive substances which provide a unique set of conditions for microbial association, quite different from those in the surrounding seawater or sediment¹². Value of shellfish microbial associates include the metabolic removal of waste products, protecting their hosts by chemically mediated defense mechanisms from dangers like predation or pathogen, and production of bioactive metabolites of pharmaceutical application¹³.

Actinomycetes, including the common soil genus Streptomyces, are proven source of structurally diverse natural products, possessing broad ranges of biological activities¹⁴. Examples include antibiotic (erythromycin and tetracycline), anticancer (mitomycin and daunomycin), immunosuppressant (rapamycin and FK506), and veterinary (thisostrepton and monensin) agents. While terrestrial strains have been actively isolated and screened for decades in academia and industry, Streptomycetes isolated from the marine environment have been largely ignored until recently. Early reports suggested that marine actinomycetes were derived from terrestrial sources and that they existed as metabolically inactive spores¹⁵.

Isolation of marine actinomycetes

Sample of each sediment was first mixed, suspended in sterile distilled water as described below. 5g of soil sample was mixed within 45 ml of sterile distilled water blank. This suspension was serially diluted up to 10⁻². One ml of the diluted sample was taken from 10⁻²dilution and the samples were pour plated on Starch Casein Agar (SCA) medium and they were incubated at 37°C for 7 days.

Composition of Starch Casein Agar (g/L) [Kuster and Williams, (1964)]

Starch	10
Casein	0.3
CaCOf	0.02
Fef SO,, .	0.001
KNOf	2
K,HPO,	2
MgSO,,	0.005
Nacl	2
Distilled water	1000
Agar	20
pH	7.0 ± 0

Quadrant Streak Technique

In order to obtain well- isolated discrete colonies, the quadrant streak should be used. As the original sample is diluted by streaking it over successive quadrant, the number of organism decreases. Usually by the third or fourth quadrant only a few organisms are transferred on the inoculating loop and these produce a few isolated colonies.

The quadrant streak technique is described below,

- 1. Flame the inoculating loop until it is red hot and then allows it to cool. Remove a small amount of actinomycetes growth with the sterile inoculating loop.
- 2. Immediately streak the inoculation loop very gently over a quarter of the plate using a

back and forth motion

- 3. Flame the loop again and allow it cool. Going back to the edge of area 1 that you just streaked, extend the streaks into the second quarter of the plate (area 2).
- 4. Flame the loop again and allow it cool. Going back to the area that you just streaked (area 2), extend the streaks into the third quarter of the plate (area 3)
- 5. Flame the loop again and allow it cool. Going back to the area that you just streaked (area 3), extend the streaks into the third quarter of the plate (area 4).

Fungal cultures

Fungal phytopathogen *Rhizoctoniasolani* which causes sheath blight in rice. They were grown on Potato Dextrose Agar (PDA) medium supplemented with chloramphenicol (50¹/4g/ml) antibiotic to avoid the bacterial contamination. They were preserved in PDA slants at 4ÚC.

Composition of Potato Dextrose A	.gar (g/L)
Potato	200
Dextrose	20
Agar	20
pH	6.5 ± 0.2

S.No	Compound	Source	activity
1	Erythromycin	Saccharopolyspora erythrae	Antibacterial
2	Kanamycin	Streptomyces kanamyceticus	Antibacterial
3	Rapamycin	Streptomyces hygroscopicus	Antifungal
4	Himastatin	Streptomyces hygroscopicus	Antitumor
5	Amphotericin	Streptomyces nodosus	Antifungal
6	Tylosin	Streptomyces fradiae	Antibacterial
7	Urdamycin A	Streptomyces fradiae	Antitumor
8	Fosfomycin	Streptomyces fradiae	Antibacterial
9	Streptomycin	Streptomyces griseus	Antibacterial
10	Valinomycin	Streptomyces griseus	Mitochondrial toxin
11	Chloramphenicol	Streptomyces venezuelae	Antibacterial
12	Rifamycin	Amycolatopsis mediterranei U32	Antibacterial
13	Amythiamicins	Amycolatopsis sp.	Antibacterial
14	Clorobiocin	Streptomyces coelicolor	Inhibitor of bacterial gyrase
15	Meilingmycin	Streptomyces nanchangensis	Antiparasitic
16	Nanchangmycin	Streptomyces nanchangensis	Insecticidal
17	Nikkomycins	Streptomyces ansochromogenus	Antifungal
18	TubelactomycinA	Nocardia sp.	Antibacterial
19	Spiramycin	Streptomyces ambofaciens	Antibacterial
20	Nogalamycin	Streptomyces nogalater	Antibacterial
21	Pristinamycin	Streptomyces pristinaespiralis	Antibacterial
22	Oligomycin	Streptomyces avermililis	Cell growth inhibitor
23	Actinomycin	Streptomyces chrysomallus	Antitumor
24	Dunamycin	Streptomyces sp.	Antitumor
25	Resormycin	Streptomyces platensis	Herbicidal, antifungal
26	Ileumycin	Streptomyces lavendulae	Antifungal
27	Mitomycin	Streptomyces lavendulae	Antitumor
28	Lomofungin	Streptomyces lomodensis	Antifungal, antibacterial
29	Axenomycins	Streptomyces lisandri nov.sp.	Antiprotozoal, Antifungal
30	Tetracycline	Streptomyces aureofaciens	Antibacterial
31	Saptomycins	Streptomyces sp.HP 530	Antitumor, Antimicrobial
32	Fattiviracin A1	Streptomyces microflavus	Antiviral
33	FK 506	Streptomyces tsukubaensis	Antiviral
34	Retamycin	Streptomyces olindensis	Antitumor
35	Apramycin	Streptomyces tenebrabrius	Antibacterial

Table 1. Secondary	metabolites	produced	bv	actinomycetes:
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RESULTS AND DISCUSSION

Table 2. Primary screening against plant pathogen

Isolation of actinomycetes

A total of 16 actinomycetes isolates (Fig. 1) were obtained from marine sediments collected from Kanathur East coast area, Chennai, India. The pure cultures of actinomycetes were maintained on starch casein agar (SCA) slants at -20°C. Primary screening of culture filtrates of marine actinomycetes against plant pathogens:

S. No.	Isolate code	Antifungal activity	Zone of Inhibition (mm)
1	P-304	+	3
2	P-309	+	2
3	P-311	+	2
4	P-312	+	2

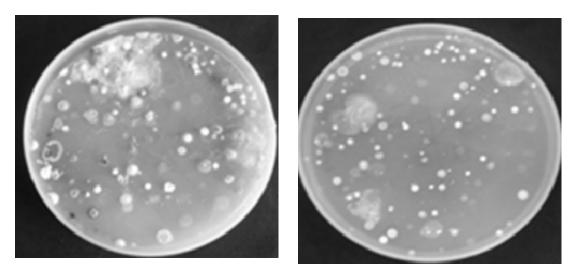


Plate showing actinomycetes colonies in sample A dilution 10⁻¹

Plate showing actinomycetes colonies in sample A dilution 10^{-3}

Fig. 1. Isolation of acitnomycetes

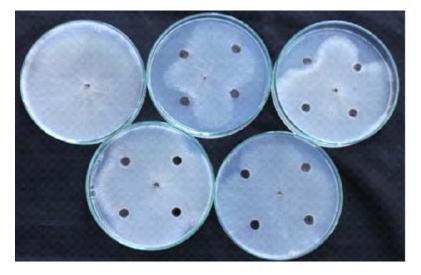


Fig. 2. Screening the culture filtrates of marine actinomycetes for antifungal activity

Among 16 isolates, four isolates showed antifungal activity against *Rhizoctoniasolani*. The four isolates, p- 304, p- 309, p- 311,p- 312 showed antifungal activity with a inhibition of 3 mm followed by 2 mm respectively. (Fig 2, Table 2).

CONCLUSION

Among the 16 isolates of marine actinomytes that were collected from the marine sediments, only four potential strains were isolated. These four potential strains showed high antifungal activity.

REFERENCES

- 1. Siva kumar, K., Actinomycetes of an Indian Mangrove (pichavaram) environment: An Inventry. Ph.D. thesis, Annamalai university, India, 2001; 91.
- 2. Rathna kala, R. and chandrika, V., Effect of different media for isolation, growth and maintenance of actinomycetes from mangrove sediments Indian *J. Mae Sci.*, 1993; **22**: 297-299.
- Crawford DL, Lynch, JM, Whipps JM, Ousley MA., Isolation and characterization of actinomycycete antagonistis of a fungal root pathogen. *Applied and environmental microbiology* 1993; **59**: 3899- 3905.Crawford, D.L., US patent 55275226,1995.
- Bressan W., Biological control of maize seed pathogenic fungi by use of actionomycetes. *Biocontrol* 2003; 48:233-240.
- Hapwood, D.A., Bill, M.J., Charter K.F., Kieser, T., Bruton, C.J., Kieser, H.M., Lydiate, D.J., Smith, C.P., Ward, J.M. and Schrempf, H. Genetic Manipulation of Streptomycetes: A Laboratory manul, John Innes Foundiation, Norwich, United Kingdom, 1985; 71-80.
- 6. Sivakumar K, Sahu MK & Kathiresan K., isolation and characterization of streptomuycetes, producing antibiotics, from a mangrove environment. *Asian J Microbial Biotech Envi Sc* 2005a; **7**:87-94
- 7. Xu L.H., Jin, X., Mao, P.M., Lu, Z.f., Cui, X.L.

and Jiang, C.L., Three new species of the genus Actinobispora of the family Pseudonocardiaceae, Actinobispora alaniniphila sp. Nov., Actinobispora aurantiaca sp. Nov.and Actinopora xinjiangensis sp. 1999.

- Lee RJ, Younger AD., developing microbiological risk assessment for shellfish purification. Int biodeterior biodegrade 2002; 50: 177-183. Doi: 10.1016/s0964-8305(02)00084-7
- Rao, M.B., Tanlsale, M. A., Ghatge, S.M., Deshpande, V., "molecular and biotechnological Aspects of Microbial proteases,"microbiological and molecular biology Reviews 1998; 62: 597-635.
- Austin B, Zhang XH., vibrio harveyin a significant pathogen of marine vertebrates and invertebrates. *Lett Appl microbial* 2006; 43:119-124
- Beg, Q.K., Kapoor, m., Mahajan, L., Hoondal, G. S.," Microbial xylanases and their industrial Applications: A Review ," *applied microbial biotechnology*. 2001; 56: 326-338.
- Zhou j, Sun C, Wanq N, Gao R, Bai S, Zhenq H, You X and Li R., Preliminary report on the biological effects of space flight on the producing strain of a new immunosuppressant kanglemycin C. J Ind Microbio 2006; 33:707-712
- Zheng Z, Zeng W, Huang Y, Yang Z,Li J, Cai H,Su W., Detection of antitumor and antimicrobial activities in marine organism associated actinomycetes isolated from the Taiwan strait, China. FEMS Microbiol Lett 2000; 188: 87-89. Doi:10.1111/j.1574-6968. 2000.tb09173.x
- 14. Omura S., Ikeda H., Ishikawa J.,Hanamoto A,. Takahashi C., Shonosw M., Takahashi Y., Horikawa H.,Nakazawa H., osonoe T., Kikuchi H., H., Shiba T., Sakaki Y. and Hattori M., Genome sequence of an industrial microorganism streptomyces avermetilis: deducting the ability of producing secondary metabolites. Proc.Natl. Acad. Sci. USA 2001; **98**: 12215-12220
- Goodfellow M. and Haynes J.A., Actinomycetes in marine sediments. In: Yakoleff V. (ed.), Biological, Biochemical, and Biomedical Aspects of Actinomycetes. Academic Press, Orlando, 1984; 453-472.