Effect of some inhibitors on hyaluronidase production by Streptococcus mitis MTCC *2695 and Streptococcus equi SED 9 species

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ABSTRACT

The inhibitory effects of some metal ions, some synthesized compounds and their derivatives with 2-aryl acenaphthimidazoles and their N-Mannich bases on optimum production of hyaluronidase was evaluated employing *Streptococcus mitis* MTCC *2695 and *Streptococcus equi* SED 9 by submerged fermentation. The maximum enzymatic activity was obtained with an initial pH (6 and 5.5), incubation temperature 37°, incubation time for 48 h and inoculum level (6 and 10 %) with inoculum age 24 h by *Streptococcus mitis* and *Streptococcus equi* SED-9 respectively. The results indicated that metals like Fe³⁺, Zn²⁺ Mn²⁺, Ag⁺, Ni²⁺, Sr²⁺ and Cu²⁺ acted as strong inhibitors of the enzyme. N-Mannich bases of 2-aryl acenaphthimidazoles exhibited promising inhibitory activity.

Key words: Hyaluronidase production, Streptococcus sp.

INTRODUCTION

Hyaluronidase (Hyase) enzymes are of three main classes: (i) hyaluronate 4-glycanohydrolase (hyaluronoglucosaminidase, EC 3.2.1.35), e.g. testicular hyaluronidases¹, (ii) hyaluronate 3- glycanohydrolase (EC 3.2.1.36), e.g. leech hyaluronidases² and (iii) hyaluronate lyase (EC 4.2.99.1), e.g. bacterial hyaluronidases³. The enzyme increases absorption and dispersion of injected drugs, fluids, resorption of radiopaque agents, dyes and toxins⁴. Hyases are widely used in many fields like orthopaedics⁵, ophthalmology⁶, surgery, dermatology, dentistry, fertilization and oncology⁷.

Two microbial strains *Streptococcus mitis* MTCC *2695 (procured from IMTECH, Chandigarh) and a local pathological isolate *Streptococcus equi* SED 9 (isolated from gingival crevices of diseased periodontium) producing an extracellular hyaluronidase enzyme within 48 h were used in this study. After rejuvenation of both cultures, they were preserved and transferred onto in nutrient agar slants. The growth content of each slant was suspended in 5 mL of sterile water and the optical density (OD) of the pooled suspension was measured at 675 nm resulting 0.580 OD that constitutes the inoculum of S. mitis and S. equi respectively. A 5 % level of each inoculum was transferred into 250 mL Erlenmeyer flask containing 50 mL of modified nutrient broth with composition (g/L) peptic digest of animal tissue, 5; sodium chloride, 5; beef extract, 1.5; yeast extract, 1.5; casein enzyme hydrolysate (type-1), 4; KH₂PO₄, 3; MgSO₄, 3; hyaluronic acid (HA), 0.001 % with pH 7. After inoculation, the flasks were incubated at 37° on a rotary shaker at 150 rpm for 48 h. At the end of fermentation 5 ml broth was aseptically withdrawn and centrifuged at 10000 x g for 30 min at 4°.

The clear supernatant obtained was subjected to enzyme assay. Hyase activity was measured spectrophotometrically by turbidity reduction assay8 using HA sodium salt (Sigma Aldrich, USA) as a substrate. The enzymatic assay is based on Dorfmans method⁹ in which the enzymatic reduction in turbidity, resulting when 1 ml of HA at 70 µg/ml was incubated with 1 ml of enzyme sample solution and incubated at 37° for 10 min and reduction in turbidity was read by measuring the absorbance at 600 nm. One unit of enzyme activity was defined as the amount of enzyme that reduced the absorbance by 0.1 at 600 nm (A $_{\!_{600}}$) in 30 min at 37°, pH 7.0 under assay conditions similar to that caused by one unit of an international standard. Some of the metal ions¹⁰ and few synthetic compounds like Imidazoles, benzimidazoles, Nacetylated, S-alkylated, N-alkylated benzimidazole derivatives, 2-mercaptobenzimidazole derivatives, 2- oxazoles, 1,3-diacetylbenzimidazoles-2-thione and its derivatives¹¹ were reported to be potent inhibitors¹² of hyaluronidase. An attempt has been made to evaluate the inhibitory effect of some synthesized compounds with 2arvl acenaphthimidazoles and their N-Mannich bases on hyase.

To investigate the influence of initial pH (4.0-9.0), temperature ranging from 20° to 55°, incubation period for 96 h, cell growth, inoculum age and inoculum level at 0.1 to 12 % and the samples were withdrawn at regular interval of 12 h, assayed for biomass (OD A_{675nm}) and enzymatic activity.

Different metallic ions (Sr²⁺, Mo²⁺, Co²⁺, Cu²⁺, Hg²⁺, Cd²⁺, Fe³⁺, Pb²⁺, Ni²⁺, Mn²⁺, Zn²⁺, Ag⁺, W²⁺, Ca²⁺ (10 mM)) in salt forms and chelators(N, N, N', N'-Tetraacetyl ethylene diamine and Ethylene diamine tetraacetate (10 mM)) were added to the production medium aseptically and assayed for enzyme activity. The inhibitory effect of some 2- aryl acenaphthimidazoles and their N-mannich bases compounds¹³ (**1a-e** and **2a-e**) were tested at 0.001 M against the enzyme hyaluronidase by colorimetric (Morgan-Elson) assay¹⁴.

The results of incubation period and temperature on the fermentation cycle by *S. mitis* and *S. equi* at pH 7.0 was noticed at 48 h at 37°C, exhibiting highest enzyme activity (151 and 143 U/mL) and biomass (0.87 and 0.68). The biomass and optimum enzyme production was detected at pH 6.0 (1.61, 181 U/mL) and 5.5 (1.44, 165 U/mL) from *S. mitis* and *S. equi* respectively. The highest enzyme activity (179 and 168 U/mL) and cell mass (1.64 and 1.47) was found in case of *S. mitis* and *S. equi* respectively

Among metallic ions used Mo²⁺ (135 and 119 %) and Ca²⁺ (117 and 88 %) stimulated enzyme production by *S. mitis* and *S. equi* SED 9 respectively where as other metallic salts decreased the enzyme activity by both the strains. The metals like Fe³⁺, Zn²⁺ Mn²⁺, Ag⁺, Ni²⁺, Sr²⁺ and Cu²⁺ acted as strong inhibitors of the enzyme hyase produced by both the strains. Metals like Co²⁺(94 and 33 %), Hg²⁺(56 and 22 %), Cd²⁺(46 and 11 %), Pb²⁺(48 and 18 %) and W²⁺ (69 and 75 %) decreased the percentage activity of hyase by *S. mitis* and *S. equi* SED 9 respectively.

From the synthesized compounds of both the series N-Mannich bases of 2-aryl acenaphthimidazoles (**2a-e**) exhibited promising inhibitory activity in comparison to derivatives of 2aryl acenaphthimidazoles (**1a-e**). The compounds 2b exhibited the highest percentage of activity (71 and 89 %) followed by 1a (49 and 81 %), 2c (64 and 79 %), 1b (63 and 77 %) and the lowest activity was found to be 1e (13 %) and 2e (17 %) against hyase produced by *S. mitis* and *S. equi* SED 9 respectively.

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REFERENCES

- Cramer, J. A., Bailey, L. C., Bailey, C. A. and Miller, R. T. Kinetic and mechanistic studies with bovine testicular hyaluronidase, *Biochim Biophys Acta*, **1200**, 315–321 (1994).
- Hotez, P., Cappello, M., Hawdon, J., Beckers, C. and Sakanari, J. Hyaluronidases of the gastrointestinal invasive nematodes *Ancylostoma canium* and Anisakis simplex: Possible functions in the pathogenesis of human zoonoses, *J Infect Dis*, **170**, 918–926 (1994).
- Berry, A. M., Lock, R. A., Thomas, S. M., Rajan, D. P., Hansman, D. and Paton, J. C. Cloning and nucleotide sequence of the *Streptococcus pneumoniae* hyaluronidase gene and purification of the enzyme from recombinant *Escherichia coli, Infect Immunol*, 62, 1101–1108 (1994).
- Radhakrishnan, V. V., Mathai, A., Shanmugham, J. and Mathews, G. J. The role of hyaluronidase in experimental cryptococcal infections, *Surgical Neurol*, 17(4), 239-244 (1982).
- Dowthwaite, G. P., Flannery, C. R., Flannelly, J., Lewthwaite, J. C. and Archer, C. W. A mechanism underlying the movement requirement for synovial joint cavitation, *Matrix Biology*, **22**(4), 311-322 (2003).
- 6. Meyer, K. and Palmer, J. W. The polysaccharides of the vitreous humour, *J*

Biol Chem, 107, 629-634 (1934).

- Menzel, E. J. and Farr, C. Hyaluronidase and its substrate hyaluronan: biochemistry, biological activities and therapeutic uses, *Cancer Lett*, **131**(1), 3-11 (1998).
- Tam, Y. C. and Chan, E. C. S. Modification enhancing reproducibility and sensitivity in the turbidity assay of hyaluronidase, *J Microbiol Methods*, 1, 255-66 (1983).
- Dorfman, A. Methods in Enzymology, Vol. I, Academic Press, New York, 166-73 (1955).
- Saxena, B. D., Khajuria, R. K. and Suri, O. P. Synthesis and spectral studies of 2mercaptobenzimidazole derivatives, *J Heterocycl Chem*, **19**, 681-683 (1982).
- Dorfman, A., Ott, M. L. and Whitney, R. The hyaluronidase inhibitor of human blood, *J Biol Chem*, **174**, 621-629 (1948).
- Meyer, K., Rapport, M. M., Hyaluronidases. Adv Enzymol Relat Subj Biochem, 13, 199-236 (1952).
- Mishra, S. K., Sahoo, S., Panda, P. K., Mishra, S. R., Parida, R. K., Ellaiah, P. and Panda, C. S. Synthesis and antimicrobial activity of some 2- aryl Acenaphthimidazoles and their N-Mannich bases, *JTR Chem*, **13**(2), 49-54 (2006).
- Hynes, W. L. and Ferretti, J. J. Assays for hyaluronidase activity, *Methods Enzymol*, 235, 606-616 (1994).

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