Chemical Investigation of Annona squamosa (Stem bark)

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ABSTRACT

4, 9-Dihydroxy -3, 8-dimethoxy- benzo [4, 5] furo [3, 2-C] chromen -6- one, 6, 7'-dihydroxy -3methoxydihydroflavonol, 5,7- dihydroxy -4'-methoxy isoflavone, 7-hydroxy -4'-methoxy isoflavone, 7,3'dihydroxy -4'-methoxy isoflavone, 4', 5, 7-trihydroxy isoflavone, 2'- hydoxy genistein have been isolated for the first time from the stem bark of *Annona squamosa* and identified by spectroscopic data.

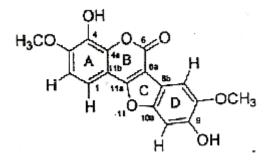
Key words: Annona squamosa, Spectroscopic data.

INTRODUCTION

Annona squamosa (family Annonaceae) is native of Africa. It is also cultivated in India¹. The decoction of the leaves of this plant is given for cholera². In continuation to earlier work³ we investigated the stem bark of *Annona squamosa* and isolated seven phenolic compounds for the first time from this source. Earlier workers isolated a wide variety of compounds viz. Amino acids⁴, terpenes⁵, sesquiterpenes⁶, diterpenes⁷, fats and oils⁸, steroids9, vitamins10, bezyltetrahydroisoquinaline¹¹, proaporphines¹², aporphines¹³, oxoaporphines¹⁴, and a large number of acetogenins¹⁵. The compounds are provisionally designated as AO-1 (I) to AO-7 (VII) and were characterised on the basis of their detailed spectroscopic analysis.

Isolation and Characterization of Compounds

The stem bark of *Annona squamosa* were purchased from United Chemicals & Allied work, Clive Row-10, Kolkatta, India. The stem bark were milled by conventional method. Defatted milled stem bark of *Annona squamosa* (5 kg) were extracted with methanol (15 l) for 16 hours. The methanolic extract (63.8 g) was then fractionated into four parts according to the increasing polarity of solvents viz, n-hexane chloroforms, ethylacetate and n-butanol. Chloroform soluble portion was chromatographed over SiO_2 gel column and eluted with solvents of increasing polarity. The silica gel column chromatography of ethyl acetate soluble portion (see experimental) yielded seven compounds compund I to VII by elution with hexane-ethyl acetate 95 : 5 upto ethyl acetate by gradual increasing of the polarity of solvents.

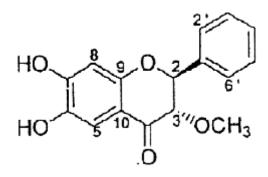


AO-1 (I)

White crystals of AO-1 (I) MF $C_{17}H_{12}O_7$ (M+ 328), m.p. 310°C responded positive to $FeCI_3$ test. Its IR absorption bands showed the presence of

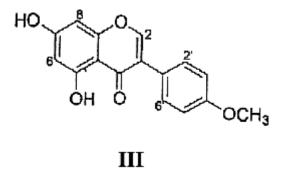
unsaturated lactone (1716 & 1253 cm⁻¹), hydroxyl group (3350 cm⁻¹) and aromatic functionality (1608 and 1427 cm⁻¹). The 1H NMR spectrum of AO-1 (I) showed the presence of one AB system [δ 7.54 (1H, d, J=8.6 Hz, H-1)] and [δ 7.00 (1H, d, J=8.6 Hz, H-2)] and one AX system [δ 7.42, 1H, s and 7.20, 1H, s]. In addition to four proton signals the spectrum also showed two singlets (δ 4.02 and 3.99, 3H, each) for two methoxyl group and two exchangeable signal at δ 9.96 and 9.07 (1H, each) for two hydroxyl group.

The ¹³C NMR, nOe studies in 1H NMR and UV spectral studies bathochromic shift with (AICl₃ and H_3BO_4) has indicated the structure of AO-1 (I) as 4, 9-dihydroxy - 3, 8 - dimethoxy - benzo [4, 5] furo [3, 2-C] chromen - 6 - one16.



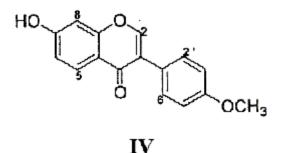
AO-2 (II)

Greenish crystals, m.p. 210-211°C. IR absorption bands showed hydroxyl group (3333 cm-1) aromaic ring (1515 and 1741 cm-1) and a carbonyl group (1653 cm-1) in its molecule, UV absorption bands at λ max 216, 235 276 and 341 nm and bathochromic shift of band II with MeOH + NaOAc suggested the presence of 7-OH group, addition of boric acid also showed bathochoromic shift of band II, thus confirming o-dihydroxyl group. 1H NMR spectrum has suggsted ring B as unsubstituted [\delta7.55 (2H, m, H-2' & H-6') and (\delta7.45, 3H, m, H-3', H-4' and H-5')] while two singlets at δ 7.19 and 6.43, 1H each was assigned to H-5 and H-8. The only methoxy singlet at δ 3.80 was assigned to methoxyl at C-3 position. Other two nonaromatic hydrogens at δ 5.14 and 4.53, dd where assigned to H-2 and H-3 respectively. Thus compound was identified as 6, 7- dihydroxy -3- methoxydihydro flavonol.



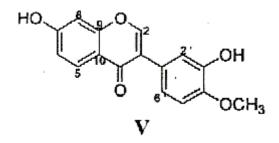
AO-3 (III)

 $C_{16}H_{12}O_5$ (MH⁺ at 285), white needles from methanol, m.p. 215-216°C. The UV data and the UV with shift reagents suggested it to be 5, 7- dihydroxy flavonoid. Its isoflavone nature was deduced by characteristic signal for H-2 at δ 8.19 (1H, s) and C-2 at δ 15.5 ppm. The ¹H NMR data AX system (δ 6.28 and 6.42, ¹H d, each J= 1.9 Hz) and A₂B₂ system (δ 7.5 and 6.98 2H each d, J= 8.8 Hz) along with two exchangeable hydrogens and a methoxy singlet (δ 3.91) has allowed us to assign the structure of AO-3 (III) as 5, 7- dihydroxy -4'- methoxy isoflavone commonly known as biochanin¹⁷.



AO-4 (IV)

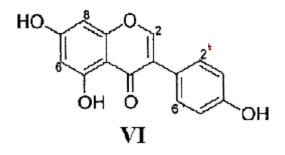
 $C_{16}H_{12}O_4$ (M⁺268) white crystal, m.p. 258°C showed in its IR spectrum absorption bands at 3424 cm–1 (hydroxyl group) and 1624 cm–1 (conjugated carbonyl group). The isoflavone nature of AO-4 (IV) was established by 1H NMR signal at δ 8.17 (1H, s). The 1H NMR spectrum also showed one ABX system in A ring [δ 8.07 (1H, d, J=9.2 Hz) δ 6.99 (1H, dd, J = 9.2 and 2.1 Hz and δ 6.90 (1H, d, J=2.1 Hz)] and an A₂B₂ system in B ring [δ 7.56 (2H, d, J=8.7 Hz) and δ 6.97 (2H, d, J=8.7 Hz)]. The position of hydroxyl and methoxyl groups were established by study of shift reagents in UV spectrum. On the basis of these data and its comparison with reported literature data18, 19 AO-4 (IV) was identified 7-hydroxy -4'- methoxy isoflavone commonly known as formonentin.



AO-5 (V)

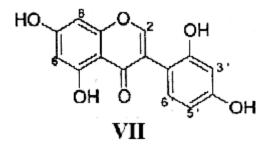
 $C_{16}H_{12}O_5$ (MH+ 285), colourless needles from methanol, m.p. 241-245°C. IR spectrum showed absorption bands for hydroxyl (3420 cm–1) carbonyl (1624 cm⁻¹) and aromatic (1580 cm⁻¹) functionalities. The UV spectrum in methanol showed λ max 224, 247 and 288 nm while in NaOAc band II showed a bathochromic shift which confirmed the presence of hydroxyl group at C-7 position in flavonoid.

¹H NMR spectrum of AO-5 (V) confirms the isoflavone nature (δ 8.83, 1H s, δ c 153.2 s). The 1H NMR spectrum also showed two ABX system in the molecule one in ring A [δ 8.03 (1H, d, J = 8.7 Hz) δ 7.02-6.97 (1H, m) and δ 6.92 (1H, d, J=2 Hz)] for H-5, H-6 and H-8 respectively and the other ABX system of B ring at δ 7.09 1H and 7.02, 6.92, 2H, m). The spectrum also showed the presence of a methoxy singlet at δ 3.84. The compound was established as 7, 3' - dihydroxy -4'- methoxy signal at δ 3.84 enhanced the signal for H-5'. This compound AO-5 (V) was thus identified as calicosin by comparison of the spectral data reported in literature²⁰.



AO-6 (VI)

 $C_{15}H_{10}O_5$ (MH⁺ at m/z 271), yellow needles m.p. 301-302°C. IR showed absorption bands for hydroxyl (3430 cm–1) and carbonyl (1650 cm⁻¹) functionalities. UV absorption maximum in methanol was found at 262 and 337 nm. Use of shift reagents has suggested the presence of phenolic group at 5 and 7 position. The presence of isoflavonic 2-H was settled by 1H NMR signal (δ 8.31, 1H, s) and A₂B₂ system in B ring (δ 7.22 and 6.81, 2H, d, each, J=8.6 Hz) and AX system in A ring (δ 7.13 and 6.21, 1H, d each J=1.8 Hz) Thus settled the structure of AO-6 (VI) as 4¹, 5, 7 trihydroxy isoflavone a commonly known isoflavone by comparison with reported data^{21,22}.



AO-7 (VII)

 $C_{15}H_{10}O_6$ (MH+ at M/z 287), m.p. 270-273°C. The IR absorption bands at umax 3350 and 1655 cm–1 and UV absorption maxima (λ max 258 and 315 nm) taken in consideration with molecular formula has suggested it to 2', 4', 5, 7- tetrahydroxy isoflavone. This is found to be in full agreement with 1H NMR data which showed AX system in A ring and ABX system in B ring along with characteristic 2-H. The compound was readily recognised as 2'hydroxy genistein, by comparison with reported spectroscopic data in literature²³.

EXPERIMENTAL

The melting point were measured on a Yazawa hot stage microstage apparatus and are uncorrected. Optical rotations were measured on JASCO DIP-360 Polarimeter (cell length 5 cm). UV absorption spectra were recorded on JASCO UV/ visible spectrophotometer (model no. 7800) while IR on JASCO FT-IR 5300 spectrometer. 1H and 13C NMR data of compounds I, II, III, IV, V, VI, VII recorded in CDCl₃ at 300 MHz and 75 MHz respectively.

Compound I

White light crystals, m.p. 310°C, EI-MS : m/z 328 [M]+, IR (KBr) υ max cm $^{-1}$ 3350, 2928, 1716, 1601, 1520, 1427, 1253 and 745, UV λ_{max} nm (MeOH) 208, 244, 345; (MeOH + NaOAc) 210, 245, 347; (MeOH + NaOAc/ H_3BO_3) 208, 244, 347, (MeOH + NaOMe) 209, 245, 378, (MeOH + AICl_3) 209, 247, 345, (MeOH + AICl_3/ HCl) 208, 244, 344

¹H NMR : δ 9.96 (1H, s, 9-OH); 9.07 (1H, s, 4-OH); 7.54 (1H, d, J=8.6 Hz, C-1H); 7.42 (1H, s, 4-OH) 7.20 (1H, s, C-10-H); 7.00 (1H, br, d, J=8.6 Hz, C-2-H); 4.02 (3H, s, 3-OCH3); 3.99 (3H, s, 8-OH-₃)

¹⁵C NMR: δ 161.0 (C-6); 159.0 (C-3); 157.0 (C-10a); 154.6 (C-11a); 149.8 (C-4); 147.2 (C-8); 147.1 (C-9); 135.4 (C-4a); 116.6 (C-1); 114.4 (C-6b); 114.1 (C-2); 105.5 (C-6a); 102.6 (C-11b); 102.2 (C-7); 99.4 (C-10); 61.0 (3-OCH₃); 56.4 (8-OCH₃)

Compound II

Greenish crystals, m.p 210-211°C, $[\alpha]^{22}D$ + 13.3 (methanol + CHCl₃, c, 0.06), FAB-MS m/z 287, IR (KBr) *v*max 3333, 1653, 11515, 1471, 830 and 766 cm⁻¹, UV λ maxnm (MeOH) 341, 276, 235, 216; (MeOH + NaOAc) 346, 255, 218; (MeOH + AlCl₃) 377, 238, 218, (MeOH + AlCl₃ + HCl) 339, 236, 219

1H NMR : δ 7.55 (m, C-H -2') 7.55 (m, C-H-6'); 7.48-7.41 (m, C-H-3'); 7.48-7.41 (m, C-H-4'); 7.48-7.41 (m, C-H-5'); 7.19 (1H, s, C-H-5); 6.43 (1H, s, C-H-8) 5.68 (1H, br d, 1 × OH); 5.14 (1H, d, J=11.6 Hz, C-H-2); 4.53 (1H, dd, J=3.7, 11.6 Hz, C-H-3); 3.80 (3H, s, 3-OCH₂)

 $\label{eq:constraint} {}^{13}\text{C NMR}: \delta 192.5 (C-4); 157.5 (C-9); 155.3 (C-7); 144.3 (C-1'); 137.9 (C-6) 128.8 (C-2'); 128.5 (C-3'); 128.5 (C-5'); 128.5 (C-6'); 128.3 (C-4'); 110.0 (C-10); 107.6 (C-8); 103.6 (C-5); 84.0 (C-3); 72.9 (C-2); 56.2 (3-OCH_2) \\ \end{array}$

Compound III

White needles, m.p 215-216°C, FAB-MS m/z 285 [M+ H], 569 [2M+ H], IR (KBr) ν max 3500, 1724, 1687, 1606, 1594, 1455, 1383, 1246, 1181, 1043 cm⁻¹, UV λ maxnm MeOH : 261, 330 sh; MeOH + NaOAc : 272, 327; MeOH + AlCl₃ + HCl) 273, 310 sh, 373

¹H NMR : δ 12.85 (1H, s, 5-OH); 8.19 (1H, s, C-H-2); 7.54 (2H, d, J=8.8 Hz, C-H-2') 7.54 (2H, d, J=8.8 Hz, C-H-8'); 6.98 (2H, d, J=8.7 Hz C-H-3'); 6.98 (2H, d, J=8.7 Hz, C-H-5'); 6.42 (1H, d, J=2.1, C-H-8); 3.91 (3H, s, 4-OCH₂)

 $\label{eq:scalar} {}^{13}\text{C}\ \text{NMR}: \delta180.5\ (\text{C-4});\ 165.1\ (\text{C-7});\ 163.9\\ (\text{C-5});\ 160.7\ (\text{C-4'});\ 159.1\ (\text{C-9});\ 154.5\ (\text{C-2});\ 131.1\\ (\text{C-6'});\ 124.1\ (\text{C-3});\ 123.7\ (\text{C-1'});\ 114.4\ (\text{C-3'});\ 114.4\\ (\text{C-5'});\ 106.1\ (\text{C-10});\ 99.1\ (\text{C-6});\ 94.4\ (\text{C-8});\ 56.3\\ (\text{4-OCH}_3) \\ \end{array}$

Compound IV

White crystals, m.p. 240°C, EI-MS m/z 284 [M]+, IR (KBr) vmax 3500, 1724, 1687, 1606, 1594, 1455, 1383, 1246, 1181, 1043 cm⁻¹, UV λ maxnm MeOH : 261, 330 sh; MeOH + NaOAc : 272, 327; MeOH + AICl₂ + HCl : 273, 310 sh, 373

¹H NMR : δ 9.72 (1H, s, 7-OH); 8.17 (1H, s, H-2, C-H-2); 8.07 (1H, d, J=8.7 Hz, C-H-5); 7.56 (2H, d, J=8.7 Hz, C-H-2') 7.56 (2H, d, J=8.7 Hz, C-H-6'); 6.99 (1H, dd, J=9.2 Hz, 2.1 Hz, C-H-6); 6.97 (2 H, d, J=8.7 Hz, C-H-3'); 6.97 (2H, d, J=8.7 Hz, C-H-5') 3.83 (3H, s, 4'-OCH₃)

 $\label{eq:constraint} {}^{13}\text{C}\,\text{NMR}: \delta\,162.7\,(\text{C-7});\,159.1\,(\text{C-4'});\,157.6\,\\ (\text{C-9});\,152.6\,(\text{C-2});\,147.8\,(\text{C-4});\,130.3\,(\text{C-6'});\,130.0\,\\ (\text{C-2'});\,127.2\,(\text{C-5});\,124.4\,(\text{C-1'});\,123.5\,(\text{C-3});\,116.8\,\\ (\text{C-10});\,115.1\,(\text{C-6});\,113.5\,(\text{C-3'});\,113.5\,(\text{C-5'});\,102.1\,\\ (\text{C-8});\,56\,(4'-\text{OCH}_2) \\ \end{array}$

Compound V

Colourless crystals, m.p. 245-247°C, FAB-MS m/z 285 [M+H]+, IR (KBr) λ max 3420, 1624, 1580, 1510, 1470, 1381, 1023, 853 cm⁻¹, UV λ_{max} nm MeOH : 288, 247, 224; MeOH + NaOAc : 327, 255, 221; NaOAc + boric acid : 288, 247, 225

¹H NMR : δ 9.10 (1H, br. hump, 4' OH) 8.33 1H, s, C-H-2); 8.03 (1H, d, J=8.7 Hz, C-H-5);7.09 (1H, C-H-2'); 7.02-6.97 (C-H-6); 7.02-6.97 (C-H-5'); 7.02-6.97 (C-H-6') 6.92 (1H, d, J=2.0 Hz, C-H-8); 3.84 (3 H, s, 7- OCH₂)

 ^{13}C NMR : δ 177.8 (C-4) δ 162.6 (C-4'); 157.5 (C-3'); 153.2 (C-2); 147.6 (C-7); 146.1 (C-10); 127.4 (C-5); 124.7 (C-9); 123.5 (C-1') 119.8 (C-2'); 116.7 (C-4); 116.5 (C-5') 115.3 (C-6); 112.0 (C-6'); 55.7 (7-OCH_3)

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Compound VI

Bright yellow needles, m.p. 301-302°C, FAB-MS m/z 271 [M+H]⁺, IR (KBr) *ν*max 3430, 2920, 1650, 1617, 1571, 1510, 1465, 1240, 1188, 1170 cm⁻¹, UV λmaxnm MeOH : 337, 262

1H NMR : δ 12.94 (1H, s, 5-OH) δ 9.56 (1H, br. hump, 7- OH); 8.31 (1H, s, C-H-2); 7.72 (2H, d, J=8.4 Hz, C-H-2'); 7.72 (2H, d, J=8.4 Hz, C-H-2') 7.72 (2H, d, J=8.4 Hz, C-H-6') 7.13 (1H, d, J=1.8 Hz, C-H-6); 6.81 (2H, d, J=8.4 Hz, C-H-3'); 6.81 (2H, d, J=8.4 Hz, C-H-5'); 6.21 (1H, d, J = 1.8, C-H-8)

¹³C NMR : δ 181.3 (C-4); 164.7 (C-7); 163.7 (C-5); 158.8 (C-9); 158.2 (C-4'); 154.0 (C-2); 130.9 (C-2'); 130.9 (C-6');123.8 (C-3); 122.9 (C-1'); 115.8 (C-3'); 115.8 (C-5'); 106.0 (C-10); 99.7 (C-6); 94.3 (C-8)

Compound VII

White needles, m.p. 270-273°C, FAB-MS m/z 287 [M+H]⁺, IR (KBr) υmax 3350, 1655, 1575, 1500, 1464, 1234, 1178, 1104 cm–1, UV λmax nm MeOH : 315 (sh), 258, MeOH + AlCl₃ : 315 (sh), 268; MeOH + AlCl₃ + HCl : 315 (sh), 268

¹H NMR : δ 12.97 (1H, s, 5-OH); ²9.29 (2H, br. d, 2 × OH) 8.13 (1H, s, C-H-2); 6.95 (1H, d, J=8.4 Hz, C-H-6'); 6.36 (1H, d, J=1.5, C-H-6); 6.34 (1H, d, J=2.1, C-H-3'); 6.25 (1H, d, J=8.4, 2.1 Hz, C-H-5'); 6.20 (1H, d, J=1.5 Hz, C-H-8)

 $\label{eq:constraint} \begin{array}{c} {}^{13}\text{C}\ \text{NMR}: \delta\ 180.5\ (\text{C-4});\ 164.2\ (\text{C-5});\ 161.2\\ (\text{C-7});\ 158.6\ (\text{C-4}');\ 157.7\ (\text{C-9});\ 156.5\ (\text{C-2}');\ 155.3\\ (\text{C-2});\ 132.2\ (\text{C-6}');\ 120.5\ (\text{C-3});\ 108.7\ (\text{C-1}');\ 106.3\\ (\text{C-5}');\ 104.5\ (\text{C-10})\ ;\ 102.7\ (\text{C-3}');\ 98.9\ (\text{C-6})\ 93.7\\ (\text{C-8}) \end{array}$

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