## Laboratory Diagnosis and Potential Application of Nucleic Acid Biosensor Approach for Early Detection of Dengue Virus Infections

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Dengue fever is caused by the dengue virus, the genus of Flaviviridae virus family. Until now, there is no specific medication to kill the dengue virus and patients just solely depend on the treatment of the dengue infection symptoms. Thus, a highly sensitive and rapid diagnostic tool for early diagnosis of dengue virus is very desirable, especially in resource limited-condition. We briefly review pro and cons of existing diagnostic methods for the detection of dengue virus (virus isolation, PCR, NS1Ag, Serology). We also highlight the recent advances of the biosensor technology in the dengue diagnostic dengue as a promising point-of-care diagnostic in the future. The DNA based biosensor technology combined miniaturized sample preparation offers a good opportunity for the commercialized point of care testing for dengue diagnosis in the future.

> Keyword: Dengue infections, Point of care, Rapid diagnostic test, Laboratory diagnosis, biosensor technology.

About 50% of world's population or 2.5 billion people may have the potential risk of dengue infection with the number of 50-100 million cases of dengue reported annually including approximately 500,000 of dengue hemorrhagic fever cases and 22,000-24,000 deaths<sup>1, 2</sup>. For the past 50 years, there are four different strains of dengue virus referred as dengue serotypes type 1, 2, 3 and 4, which is transmitted to the human through the bite of female mosquitoes, Aedes *aegypti*. In October 2013, a new dengue serotype fifth has been announced as the result of blood's

patients screening from the hospital of Sarawak state, Malaysia<sup>3, 4</sup>. Dengue virus infection caused by any of these serotypes can be asymptomatic infection (mild) or displayed in the range of severity disease from undifferentiated febrile fever and classical dengue fever to the more severe fatal dengue hemorrhagic fever (DHF) and dengue shock syndrome (DHS)<sup>5, 6</sup>. Until now, there are no fully established specific vaccine or antiviral for dengue infection<sup>7, 8</sup> and most of the treatments only rely on the symptoms and a good maintenance fluid replacement to patient's body<sup>9</sup>. Therefore, the

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effective ways to prevent the dengue infection is to control the spread of mosquito vector by reducing and destroying the mosquito vector population<sup>10</sup>. **Diagnostic methods of dengue virus** 

An early dengue diagnosis becomes important for the confirmation of dengue virus infection, clinical treatment, dengue outbreaks surveillance and monitoring efficiency of dengue vaccine trials. The clinical diagnosis of dengue infection commonly depends on reported syndromes and physical examination, which are not effective. This is because the dengue symptoms are not specific, and it is difficult to differentiate it with other infections such as malaria, West Nile, leptospirosis, measles, influenza, yellow fever and St Louis encephalitis at the early stage of infection<sup>11, 12</sup>. For this reason, several types of laboratory diagnostic tests have been done to confirm the dengue virus infection such as virus isolation, genome detection, virus antigen detection and specific antibody (serology) from patient's serum<sup>13-15</sup>. In general, dengue diagnosis is classified into two approaches of dengue virus detection; early detection and indirect detection

#### Virus isolation

For the past century, virus isolation technique remains as a "gold standard" for diagnosing dengue virus infection<sup>16, 17</sup>. The dengue virus may be recovered from infected samples such as whole blood, serum, plasma and tissues (acute samples, 4-5 days) before inoculating it into either 1 to 3-day baby mice, mosquito or cell cultures (mosquito's cell lines, C6/36 or mammalian cell lines, LLC-MK2, BHK-21 cells). This is followed by dengue virus confirmation tests using immunofluorescence (IF) technique with a specific monoclonal antibody or FTIC-conjugated anti-flavivirus antibody under UV microscope<sup>18,</sup> <sup>19</sup>. Mosquito inoculation has been reported as the most sensitive for dengue virus isolation technique in comparison with cell cultures and baby mice inoculation<sup>20,21</sup>. However, the disadvantages of this isolation technique are; it required high technical skill for dengue inoculation, include tedious steps and need high maintenance cost.

#### Antigen detection

In recent years, the detection of nonstructural protein 1 (NS1) has gained attention in the development of dengue diagnostic tests<sup>17,22</sup>. NS1, a highly-conserved glycoprotein (445KDa)

which was produced during dengue virus replication and circulated in bloodstream or serum at high concentration during acute infection<sup>23-25</sup>. According to Libraty et al. [26], high level of NS1 in plasma has been found in DHF patients in comparison with DF patients. However, the level of circulating NS1 in serum, plasma or whole blood will decrease with antibodies production of IgM and IgG after 5 days' infection making NS1 detection applicable at early acute dengue infection. Some of commercial diagnostic tests based on NS1 detection employing immune-chromatographic (ICT) and Enzyme-linked immunoassay ELISA methods have been developed for dengue diagnosis (Table 1). The disadvantages of this tests are unable to differentiate among dengue serotypes due to NS1 glycoprotein that is highly conserved for all dengue serotypes<sup>27</sup>. As anti-NS1 antibodies that are present more frequently in secondary infection, the sensitivity of NS1 assays for secondary infection is lower than in primary infection as reported by previous literature<sup>24, 28-31</sup>. As shown in Table 1, most of the commercial NS1 tests exhibit a poor sensitivity of NS1 detection ranging from 37-68%. **Genomic detection** 

Nucleic acid detection using reversetranscriptase polymerase chain reaction (RT-PCR) protocol by amplifying the specific region of interest gene has become a powerful tool for early diagnosis of dengue virus due to the ability in providing a better sensitivity, more specificity and rapid result (less than 24 hours)<sup>40-42</sup>. Furthermore, the RT-PCR technique shows higher sensitivity and is cheaper than dengue detection gold standard, virus isolation technique <sup>43-45</sup>. By using specific primers in polymerase chain reaction, the dengue virus serotypes can be determined and differentiated allowing this technique to be highly suitable for the detection of new dengue serotypes in endemic areas.

RT-PCR technique involves three steps; RNA extraction, cDNA gene amplification, detection and characterization of amplified cDNA gene. The RT-PCR procedure starts from the extraction of RNA dengue virus from human samples (serum, plasma, whole blood, urine or saliva) followed by DNA transcription process where RNA extract is converted into complementary DNA (cDNA) using reverse transcription enzyme before cDNA amplification process is performed<sup>46</sup>. According to Chen *et al*<sup>47</sup>, the analysis of amplified cDNA gene by PCR-based technique can be performed in three-technique: a) agarose gel electrophoresis b) southern blot methods c) ELISA technique. Several modified PCR-based technique has been established for the diagnosis of dengue infection such as real-time RT-PCR<sup>48-50</sup>, nucleic acid sequence-based amplification (NASBA)<sup>51</sup> and most recently, the loop-mediated isothermal amplification (LAMP)<sup>52</sup>. **Serology tests** 

After 5 days of illness, serological methods are the best choice for dengue diagnosis, which relies on dengue antibodies detection of IgM and IgG. It is important to note that these antibodies, IgM and IgG are not produced at early acute of dengue infection and can be found in serum, plasma, saliva and blood. Different responses of antibodies level are observed and measured where the antibodies of IgM and IgG rise rapidly in primary and secondary infection. Several serological methods have been established for dengue virus detection including the hemagglutination inhibition test (HI), Neutralization test (NT), MAC ELISA and IgG ELISA. Among these tests, IgM/IgG assays using ELISA and Immune-chromatographic (ICT) technique are commonly used for dengue diagnosis due to simple operation without the use of the sophisticated machine, rapid results, inexpensive and reasonable sensitivity<sup>53, 54</sup>. However, high cross-reactivity with other flaviviruses and inability to determine the dengue serotypes become the major limitations in this dengue diagnosis as it can lead to false positive/negative results such as leptospirosis and rheumatoid arthritis<sup>27, 55</sup>. A few commercials of ELISA IgM and IgG assays are available for dengue diagnostic (Table 2) where IgM commercial assays exhibit the range of sensitivity of 20.5% to 77.8% and specificity of 86% to 100%. For commercial IgG assays, the sensitivity is in the range of 66% to 94.4% while specificity in the range of 86% to 100%. Summary of the pro and cons for all dengue diagnostic tests are listed in Table 3.

#### Biosensor as new approach in clinical diagnostic

The biosensor application in clinical diagnostic may overcome the limitation of the existing dengue diagnostic-based laboratory method, which allows the continuous monitoring, rapid response, cost-effective method, high sensitivity and specificity and easy operation with minimum sample preparation. The biosensor can be defined as an analytical device incorporated with sensing material and molecular recognition elements such as an enzyme, protein antibodies, nucleic acid, hormone, chemical compounds integrated within transducers<sup>62, 63</sup>. The basic principle of this biosensor detection relies on the interaction between molecule recognition in sensor devices with its specific target and the change of this biochemical is translated into quantifiable signal responses via the transducer, whether in the form of electrochemical<sup>64, 65</sup>, electrical<sup>66, 67</sup>, optical<sup>68, 69</sup> or piezoelectric<sup>70</sup>.

# Nucleic acid-based biosensor in dengue virus detection

With the demand for rapid, simple and point of care (POC) tests in clinical diagnostic, there have been growing interests in exploring biosensor application for dengue virus detection<sup>71-79</sup>. Most of them applied nucleic acid aptamers (genosensor) as the molecule recognition element in their fabricated DNA biosensor for dengue virus detection by the hybridization process [80]. Nucleic acid aptamers are a short sequence of synthetic oligonucleotide (single-stranded DNA or RNA) approximately 25-100 bases with the specific sequence that can recognize and bind with high affinity to its nucleic acid target<sup>80-82</sup>. In general, the nucleic acid hybridization detection via biosensor approach is based on a duplex of nucleic acid formation (DNA-DNA, DNA-RNA) as the results of two single-stranded nucleic acids are combined between nucleic acid aptamers with specific sequence (known as DNA probe) with its specific complementary sequences (known as DNA target)<sup>83, 84</sup>. Using a suitable hybridization indicator, this duplex DNA hybridization can be detected and converted into a measurable signal including optical, electrochemical, electrical and piezoelectric.

In our previous work , we successfully constructed a simple fabrication of DNA dengue electrochemical biosensor where DNA probes are immobilized (23 bases) on the surface of indium tin oxide (ITO) and screen-printed gold electrode (SPGE), respectively before allowing hybridization with its complementary DNA dengue sequences<sup>80, 81, 84</sup>. The DNA-DNA hybridization detection was

monitored using the electrochemical method based on methylene blue oxidation. The RNA dengue virus detection based on DNA/RNA hybridization using optical biosensor approach has been shown in the work of Kwakye et al<sup>85</sup>. In their studies, two different DNA probes were used; the DNA probes functionalized liposome and reporter probe tagged with magnetic beads were hybridized with amplified RNA dengue target forming a hybridized DNA/RNA complex-liposome that can be visualized under fluorescent microscopy. The interesting fact about their work is the level of RNA virus concentration can be estimated and it also managed to achieve 10 times lower of the limit of detection (LOD) of RNA dengue virus than of conventional laboratory methods. Nascimento et al<sup>86</sup> had utilized the gold nanoparticles-polyaniline hybrid composite layer as DNA matrix in their fabricated DNA sensor to improve probe loading which in turn increased the DNA target loading and thus, resulted in the high sensitivity and able to discriminate the dengue serotypes 1,2 and 3 at the picomolar concentration of DNA dengue virus detection. The incorporation of RT-PCR methods in DNA biosensor for rapid detection of dengue virus has been reported by Rai et al75. In their work, a nanop orous alumina membrane biosensor was constructed where the DNA probes are immobilized inside the alumina channel and upon hybridization with amplified RT-PCR products, the reduction of the electrochemical signal occur indicating successful detection of dengue virus. In this strategy, the RT-PCR technique is used to amplify the level of concentration of genomic nucleic acid

Table 1. The commercial of dengue diagnostic tests based on the NS1 detection

Commercial NS1 diagnostic test	Sensitivity	Specificity	References
ELISA technique			
Platelia Dengue NS Ag-ELISA	68%	96%	[32]
Platelia Dengue NS Ag-ELISA	37%	100%	[33]
Platelia Dengue NS Ag-ELISA	45%	92%	[24]
PanBio Dengue Early ELISA kit	65%	98%	[23]
PanBio Dengue Early ELISA kit	45%	92%	[24]
PanBio Dengue Early ELISA kit	69%	96%	[34]
PanBio Dengue Early ELISA kit	66%	100%	[35]
Dengue NS1 Elisa	65%	98%	[36]
Immune-chromatographic (ICT) technic	que		
PanBio NS1 antigen strip	46%	98%	[37]
Dengue NS1 Ag Strip kit	49%	99%	[38]
Dengue NS1 Ag Strip kit	51%	97%	[39]

Table 2. The commercial of IgM and IgG assays for dengue diagnosis

Commercial of IgM/IgG assays	Sensitivity	Specificity	References
IgM assays			
SD Bioline IgM (ICT)	61%	90%	[56]
Zephyr IgM (ICT)	20.5%	86%	[56]
SD Duo IgM (ICT)	53.5%	100%	[36]
Omega capture ELISA	62.3%	97.8%	[56]
Venture IgM ELISA	68.7%	100%	[57]
PanBio IgM ELISA	77.8%	90.6%	[41]
IgG assays			
PanBio IgG capture ELISA	81.2%	63.5%	[58]
Panbio Dengue Duo cassette (ICT)	66.4%	94.4%	[59]
Panbio Dengue Duo (ICT)	87.5%	66.6%	[60]
SD Bioline IgG	90.06%	92.48%	[61]

dengue virus before it can be detected by their fabricated DNA biosensor. This combination of PCR-DNA biosensor can replace the conventional PCR products analysis, which relies on agarose gel electrophoresis and southern/northern blotting. The use of agarose gel electrophoresis analysis exhibit low sensitivity and does not provide specific sequence information of amplified nucleic acid<sup>87</sup>. Besides that, this conventional PCR product analysis is usually time-consuming, consist of tedious steps requires specialized equipment and hazardous elements such as ethidium bromide and ultraviolet<sup>88, 89</sup>. Another study of DNA biosensor coupling with RNA target amplification based PCR technique was reported by Baeumner et al79. Based on their work, the RNA extract was first amplified using isothermal nucleic acid sequencebased amplification (NASBA) for DNA/RNA hybridization using liposome amplification signal. The first reporter probe tagged with liposome was mixed and hybridized with the amplified RNA before being introduced to the surface of a polyethersulfone membrane trip containing immobilized DNA probe. With this DNA/RNA hybridization detection, the amount of RNA dengue virus was correlated with the amount of

liposome that can be measured using a portable reflectometer. They have found that their fabricated DNA biosensor could detect the RNA dengue virus concentration at low concentration of 10 PFU/mL.

Recently, some studies have employed dengue monoclonal antibodies based biosensor (also known as immunosensors) which are based on antibodies-antigen interaction for the detection of dengue biomarker proteins such as NS190, 91 and IgM92. However, the application of DNA biosensor is more advantageous and can be an alternative compared to immunosensors based on few reasons. In contrast to antibodies, nucleic acid aptamers (DNA probe) are more easily synthesized in a large-scale production which can offer high sensitivity and specificity, high stability (pH and temperature), long shelf life, and can be regenerated after the heat-denatured process of hybridized DNA<sup>82, 89, 93</sup>. Besides that, the production of nucleic acid aptamers is a cost-effective and less time-consuming as compared to antibodies which rely on expensive animal hosts to produce antibody up to 6 months94,95. As aptamers are easily modified and conjugated with labelled/reporter molecules compared to antibodies, making DNA biosensor is more versatile than immunosensors.

Diagnostic tests	Advantages	Disadvantages
Virus isolation	Specific	Time consuming (6-10 days)
	Reliable results	Lower sensitivity
		Expensive
		Laborious steps
		Require appropriate facilities and expertise skills
		Require acute sample (0-5 days)
RT-PCR technique	High sensitivity	Expensive
	High specificity	Require specialized instrumentation
	Results less than 24 hours	Require careful handling to avoid cross-contamination
	Reliable results	during the RT-PCR procedure
		Involve hazardous chemicals
		Require acute sample (0-5 days)
NS1 detection	Inexpensive	Lower sensitivity
	Rapid results (In a few hours)	High cross-reactivity with another antigen/antibody
	Easy to operate	False positive/negative results)
		Require acute sample (0-5 days)
Serology test	Inexpensive	Lower sensitivity
(IgG and IgM)	Rapid results (In a few hours)	High cross-reactivity with other flaviviruses and
	Easy to operate	antigen/antibody
	_	False positive/negative results
		Require febrile sample (5 days above)

Table 3. Summary of pro and cons for current diagnostic tests for dengue infection

Furthermore, the smaller size of nucleic acids than antibodies allow high-density monolayer of aptamer recognition which gives advantages in the miniaturization of the biosensor<sup>96, 97</sup>.

#### **Conclusions and Outlook**

In this review paper, we discuss the pro and cons of existing conventional dengue diagnostic methods. Most conventional diagnostic methods are still based on laboratory methods, timeconsuming, require sophisticated instrumentation and trained person to diagnosis dengue infections which are not suitable in resource-limited condition. Although, dengue rapid test (RDT) based immunochromatography (NS1 antigen and IgM) are available in the market that can eliminate the use of laboratory equipment, but it is more suitable for initial screening to identify whether a person is infected or not before need a further confirmatory test. This is attributed that commercial RDTs have a high risk of cross-reactivity with other flaviviruses leading to false-positive results. Moreover, the lack of sensitivity is often a major challenge in dengue diagnostic based RDT approach. DNA-based biosensor technology has gained an attention by many researchers for its potential to replace the conventional dengue diagnostic method due to its rapid diagnostic as early as one day, cost-effective, highly sensitive and specific, easily miniaturized and can be operated in resource-limited condition. Interestingly, monitoring the level of nucleic acid of dengue virus concentration in patients can provide a valuable information about the progression of dengue virus infection, thus effective treatment could be achieved. To date, there is no DNA-based biosensor for dengue virus cases detection have been commercialized yet. One of the possible reason is the complexity of clinical samples that require bulky instrumentation for sample preparation assays before being detectable by biosensor that could limit the application in pointof-care testing. To date, much effort to put towards the miniaturized sample preparation assays that hold promise to be integrated with DNA biosensor in one single system such as microfluidic or labon-a-chip, giving opportunities for commercialized point-of-care testing for dengue diagnosis. We believe this DNA-based biosensor technology combined miniaturized sample preparation seems compatible with the internet of things (IoT)

application that could revolutionize healthcare especially in dengue diagnostic.

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#### REFERENCES

- Nagoor, K., Babu, S., Kahn, S., Kalluri, R.J., John, K. Knowledge, attitude and practice on dengue fever and its prevention and control measures in urban slums of South India. *International Journal Of Community Medicine And Public Health*, 2017; 4(8):3013-17.
- Husin, F., Chan, Y.Y., Gan, S.H., Sulaiman, S.A., Shueb, R.H. The Effect of hydrocotyle sibthorpioides Lam. extracts on in vitro dengue replication. *Evidence-based complementary and alternative medicine* 2015; (2015): 9. http:// dx.doi.org/10.1155/2015/596109.
- Mustafa, M., Rasotgi, V., Jain, S., Gupta, V. Discovery of fifth serotype of dengue virus (DENV-5): A new public health dilemma in dengue control. *Medical Journal Armed Forces India*, 2015; 71(1):67-70.
- Normile, D. Surprising new dengue virus throws a spanner in disease control efforts. *Science*, 2013; **342**(6157):415-15.
- Villamor, E., Villar, L., Lozano, A., Herrera, V., Herrán, O. Vitamin D serostatus and dengue fever progression to dengue hemorrhagic fever/dengue shock syndrome. *Epidemiology & Infection*, 2017; 145(14):2961-70.
- Buchillet, D. Dengue and dengue-like outbreaks in the past: The case of the Macau epidemic fever of 1874. *Infection, Genetics and Evolution*, 2012; 12(5):905-12.
- 7. Faraz, A., Aadil, M., Shafqat, M.N. Fighting the dengue virus. *Cureus*, 2017; **9**(5):e1271.
- Bhat, V.G., Chavan, P., Ojha, S.,Nair, P.K. Challenges in the Laboratory Diagnosis and Management of Dengue Infections. *The open microbiology journal*, 2015; 9:33-7.
- 9. Guzman, M.G.,Kouri, G. Dengue: an update. Lancet Infect Dis, 2002; 2(1):33-42.
- Lee, H., Rohani, A., Khadri, M., Nazni, W., Rozilawati, H., Nurulhusna, A., Nor Afizah, A., Roziah, A., Rosilawati, R., Teh, C. Dengue

vector control in malaysia-challenges and recent advances. *International Medical Journal Malaysia*, 2015; **14**(1):11-16.

- Muller, D.A., Depelsenaire, A.C., Young, P.R. Clinical and laboratory diagnosis of dengue virus infection. *The Journal of infectious diseases*, 2017; 215(suppl\_2):S89-S95. https:// doi.org/10.1093/infdis/jiw649
- Cheah, W., Ng, K., Marzilawati, A., Lum, L. A review of dengue research in malaysia. *The Medical journal of Malaysia*, 2014; 69: 59-67.
- Nisalak, A. Laboratory diagnosis of dengue virus infections. Southest Asian J Trop Med Public Health, 2015; 46(1):55-76.
- Samuel, P.P., Tyagi, B. Diagnostic methods for detection & isolation of dengue viruses from vector mosquitoes. *Indian journal of medical research*, 2006; **123**(5):615-18.
- 15. Kao, C.-L., King, C.-C., Chao, D.-Y., Wu, H.-L., Chang, G. Laboratory diagnosis of dengue virus infection: current and future perspectives in clinical diagnosis and public health. *J Microbiol Immunol Infect*, 2005; **38**(1):5-16.
- Wasik, D., Mulchandani, A., Yates, M.V. A heparin-functionalized carbon nanotube-based affinity biosensor for dengue virus. *Biosensors* and Bioelectronics, 2017; 91:811-16.
- Jyothi, P., Metri, B.C. Correlation of serological markers and platelet count in the diagnosis of dengue virus infection. *Advanced biomedical research*, 2015; 4. 10.4103/2277-9175.150396
- Gubler, D., Kuno, G., Sather, G., Velez, M., Oliver, A. Mosquito cell cultures and specific monoclonal antibodies in surveillance for dengue viruses. *The American Journal of Tropical Medicine and Hygiene*, 1984; 33(1):158-65.
- Henchal, E., Gentry, M., McCown, J., Brandt, W. Dengue virus-specific and flavivirus group determinants identified with monoclonal antibodies by indirect immunofluorescence. *Am J Trop Med Hyg*, 1982; **31**(4):830-6.
- Vaughn, D.W., Green, S., Kalayanarooj, S., Innis, B.L., Nimmannitya, S., Suntayakorn, S., Endy, T.P., Raengsakulrach, B., Rothman, A.L., Ennis, F.A. Dengue viremia titer, antibody response pattern, and virus serotype correlate with disease severity. *Journal of Infectious Diseases*, 2000; 181(1):2-9.
- Gubler, D., Sather, G., Fonseca da Cunha, F., 1988. Laboratory diagnosis of dengue and dengue hemorrhagic fever, Simposio Internacional sobre Febre Amarela e Dengue, pp. 291-322.
- Vickers, I., Harvey, K., Nelson, K., Brown, M., Bullock-DuCasse, M., Lindo, J. Evaluation of onestep dengue NS1 rapidip<sup>TM</sup> instatest and onestep dengue fever IgG/IgM rapicard<sup>TM</sup>

instatest during the course of a dengue type 1 epidemic. *Diagnostic Microbiology and Infectious Disease*, 2017; **89** (4): 271-275.

- Pal, S., Dauner, A.L., Mitra, I., Forshey, B.M., Garcia, P., Morrison, A.C., Halsey, E.S., Kochel, T.J., Wu, S.-J.L. Evaluation of dengue NS1 antigen rapid tests and ELISA kits using clinical samples. 2014. <u>https://doi.org/10.1371/journal.pone.0113411</u>
- Blacksell, S.D. Commercial dengue rapid diagnostic tests for point-of-care application: recent evaluations and future needs? *BioMed Research International*, 2012; 2012. http:// dx.doi.org/10.1155/2012/151967.
- Young, P.R., Hilditch, P.A., Bletchly, C., Halloran, W. An antigen capture enzyme-linked immunosorbent assay reveals high levels of the dengue virus protein NS1 in the sera of infected patients. *Journal of clinical microbiology*, 2000; 38(3):1053-57.
- Libraty, D.H., Young, P.R., Pickering, D., Endy, T.P., Kalayanarooj, S., Green, S., Vaughn, D.W., Nisalak, A., Ennis, F.A., Rothman, A.L. High circulating levels of the dengue virus nonstructural protein NS1 early in dengue illness correlate with the development of dengue hemorrhagic fever. *Journal of Infectious Diseases*, 2002; **186**(8):1165-68.
- Peeling, R.W., Artsob, H., Pelegrino, J.L., Buchy, P., Cardosa, M.J., Devi, S., Enria, D.A., Farrar, J., Gubler, D.J., Guzman, M.G. Evaluation of diagnostic tests: dengue. *Nature Reviews Microbiology*, 2010; 8: S30-8. doi: 10.1038/ nrmicro2459.
- Hunsperger, E.A., Yoksan, S., Buchy, P., Nguyen, V.C., Sekaran, S.D., Enria, D.A., Vazquez, S., Cartozian, E., Pelegrino, J.L., Artsob, H. Evaluation of commercially available diagnostic tests for the detection of dengue virus NS1 antigen and anti-dengue virus IgM antibody. 2014; 8: e3171. doi: <u>10.1371/journal.</u> pntd.0003171.
- 29. Blacksell, S.D., Mammen, M.P., Thongpaseuth, S., Gibbons, R.V., Jarman, R.G., Jenjaroen, K., Nisalak, A., Phetsouvanh, R., Newton, P.N., Day, N.P. Evaluation of the Panbio dengue virus nonstructural 1 antigen detection and immunoglobulin M antibody enzyme-linked immunosorbent assays for the diagnosis of acute dengue infections in Laos. *Diagnostic microbiology and infectious disease*, 2008; **60**(1):43-49.
- Kumarasamy, V., Wahab, A.A., Chua, S., Hassan, Z., Mohamad, M., Chua, K. Evaluation of a commercial dengue NS1 antigen-capture ELISA for laboratory diagnosis of acute dengue virus

infection. *Journal of virological methods*, 2007; **140**(1):75-79.

- 31. Valdés, K., Alvarez, M., Pupo, M., Vázquez, S., Rodríguez, R., Guzmán, M.G. Human dengue antibodies against structural and nonstructural proteins. *Clinical and diagnostic laboratory immunology*, 2000; 7(5):856-57.
- 32. Bessoff, K., Delorey, M., Sun, W., Hunsperger, E. Comparison of two commercially available dengue virus (DENV) NS1 capture enzymelinked immunosorbent assays using a single clinical sample for diagnosis of acute DENV infection. *Clinical and vaccine immunology*, 2008; **15**(10):1513-18.
- Phuong, H.L., Thai, K.T., Nga, T.T., Giao, P.T., Hung, L.Q., Binh, T.Q., Nam, N.V., Groen, J., de Vries, P.J. Detection of dengue nonstructural 1 (NS1) protein in Vietnamese patients with fever. *Diagnostic microbiology and infectious disease*, 2009; 63(4):372-78.
- 34. Fry, S.R., Meyer, M., Semple, M.G., Simmons, C.P., Sekaran, S.D., Huang, J.X., McElnea, C., Huang, C.-Y., Valks, A., Young, P.R. The diagnostic sensitivity of dengue rapid test assays is significantly enhanced by using a combined antigen and antibody testing approach. *PLoS Negl Trop Dis*, 2011; 5(6):e1199. doi: 10.1371/ journal.pntd.0001199
- Shenoy, B., Menon, A., Biradar, S. Diagnostic utility of dengue NS1 antigen. *Pediatric Infectious Disease*, 2014; 6(3):110-13.
- Wang, S.M., Sekaran, S.D. Early diagnosis of Dengue infection using a commercial Dengue Duo rapid test kit for the detection of NS1, IGM, and IGG. *The American journal of tropical medicine and hygiene*, 2010; 83(3):690-95.
- Pan-ngum, W., Blacksell, S.D., Lubell, Y., Pukrittayakamee, S., Bailey, M.S., de Silva, H.J., Lalloo, D.G., Day, N., White, L.J., Limmathurotsakul, D. Estimating the true accuracy of diagnostic tests for dengue infection using bayesian latent class models. *PloS one*, 2013; 8(1):e50765. <u>doi.org/10.1371/journal.</u> pone.0050765.
- Zainah, S., Wahab, A.A., Mariam, M., Fauziah, M., Khairul, A., Roslina, I., Sairulakhma, A., Kadimon, S., Jais, M.M., Chua, K. Performance of a commercial rapid dengue NS1 antigen immunochromatography test with reference to dengue NS1 antigen-capture ELISA. *Journal of virological methods*, 2009; 155(2):157-60.
- Osorio, L., Ramirez, M., Bonelo, A., Villar, L.A., Parra, B. Comparison of the diagnostic accuracy of commercial NS1-based diagnostic tests for early dengue infection. *Virological Journal*, 2010; 7(361):7-361.

- 40. Wilson, H.L., Tran, T., Druce, J., Dupont-Rouzeyrol, M., Catton, M. Neutralization Assay for Zika and Dengue Viruses by Use of Real-Time-PCR-Based Endpoint Assessment. *Journal* of Clinical Microbiology, 2017; **55**(10):3104-12.
- Decker, D.S., Vray, M., Sistek, V., Labeau, B., Enfissi, A., Rousset, D., Matheus, S. Evaluation of the diagnostic accuracy of a new dengue IgA capture assay (Platelia Dengue IgA Capture, Bio-Rad) for dengue infection detection. *PLoS neglected tropical diseases*, 2015; 9(3):e0003596-e96.
- Najioullah, F., Viron, F., Césaire, R. Evaluation of four commercial real-time RT-PCR kits for the detection of dengue viruses in clinical samples. *Virology journal*, 2014; 11(1):1-5.
- Azhar, E., Kao, M., Niedrig, M., Masri, B., Godus, A., Badierah, R., Khan, N., Almazrooa, A., Ashshi, A., Jamjoom, G. Virological diagnosis of dengue fever in Jeddah, Saudi Arabia: Comparison between RT-PCR and virus isolation in cell culture. 2010; 2(2):24-29.
- Grobusch, M., Niedrig, M., Göbels, K., Klipstein Grobusch, K., Teichmann, D. Evaluation of the use of RT PCR for the early diagnosis of dengue fever. *Clinical microbiology and infection*, 2006; 12(4):395-97.
- Sa-ngasang, A., Wibulwattanakij, S., Chanama, S., O-rapinpatipat, A., A-nuegoonpipat, A., Anantapreecha, S., Sawanpanyalert, P., Kurane, I. Evaluation of RT-PCR as a tool for diagnosis of secondary dengue virus infection. *Japanese journal of infectious diseases*, 2003; 56(5/6):205-09.
- Teles, F., Prazeres, D., Lima Filho, J. Trends in dengue diagnosis. *Reviews in medical virology*, 2005; 15(5):287-302.
- Chien, L.-J., Liao, T.-L., Shu, P.-Y., Huang, J.-H., Gubler, D.J., Chang, G.-J.J. Development of real-time reverse transcriptase PCR assays to detect and serotype dengue viruses. *Journal of clinical microbiology*, 2006; 44(4):1295-304.
- Huhtamo, E., Hasu, E., Uzcátegui, N.Y., Erra, E., Nikkari, S., Kantele, A., Vapalahti, O., Piiparinen, H. Early diagnosis of dengue in travelers: comparison of a novel real-time RT-PCR, NS1 antigen detection and serology. *Journal of Clinical Virology*, 2010; 47(1):49-53.
- Gurukumar, K., Priyadarshini, D., Patil, J., Bhagat, A., Singh, A., Shah, P., Cecilia, D. Development of real time PCR for detection and quantitation of Dengue Viruses. *Virology journal*, 2009; 6(1):10.
- 50. Johnson, B.W., Russell, B.J., Lanciotti, R.S. Serotype-specific detection of dengue viruses in a fourplex real-time reverse transcriptase PCR

assay. Journal of clinical microbiology, 2005; **43**(10):4977-83.

- Jittmittraphap, A., Thammapalo, S., Ratanasetyuth, N., Wongba, N., Mammen, M.P., Jampangern, W. Rapid detection of dengue viral RNA in mosquitoes by nucleic acidsequence based amplification (NASBA). 2006; 37(6):1117-24.
- Lau, Y.-L., Lai, M.-Y., Teoh, B.-T., Abd-Jamil, J., Johari, J., Sam, S.-S., Tan, K.-K., AbuBakar, S. Colorimetric Detection of Dengue by Single Tube Reverse-Transcription-Loop-Mediated Isothermal Amplification. *PloS one*, 2015; 10(9):e0138694.
- 53. van Meer, M.P., Mögling, R., Klaasse, J., Chandler, F.D., Pas, S.D., van der Eijk, A.A., Koopmans, M.P., Reusken, C.B., GeurtsvanKessel, C.H. Re-evaluation of routine dengue virus serology in travelers in the era of Zika virus emergence. *Journal of Clinical Virology*, 2017; **92**:25-31.
- Zhang, B., Salieb-Beugelaar, G.B., Nigo, M.M., Weidmann, M., Hunziker, P. Diagnosing dengue virus infection—rapid tests and the role of micro/nanotechnologies. *Nanomedicine: Nanotechnology, Biology and Medicine,* 2015; 11(7):1745-61.
- 55. Berlioz-Arthaud, A., Gurusamy, A. Comparison of PanBio dengue IgM ELISA assay with pentax dengue IgM particle agglutination assay to evaluate factors affecting false positive results. *Southeast Asian Journal of Tropical Medicine and Public Health*, 2008; **39**(1):55-61.
- Hunsperger, E.A., Yoksan, S., Buchy, P., Nguyen, V.C., Sekaran, S.D., Enria, D.A., Pelegrino, J.L., Vázquez, S., Artsob, H., Drebot, M. Evaluation of commercially available anti-dengue virus immunoglobulin M tests. *Emerging infectious diseases*, 2009; 15(3):436-40.
- 57. Anders, K.L., Nguyet, N.M., Quyen, N.T.H., Van Ngoc, T., Van Tram, T., Gan, T.T., Tung, N.T., Dung, N.T., Chau, N.V.V., Wills, B. An evaluation of dried blood spots and oral swabs as alternative specimens for the diagnosis of dengue and screening for past dengue virus exposure. *The American journal of tropical medicine and hygiene*, 2012; **87**(1):165-70.
- Pal, S., Dauner, A.L., Valks, A., Forshey, B.M., Long, K.C., Thaisomboonsuk, B., Sierra, G., Picos, V., Talmage, S., Morrison, A.C. Multicountry Prospective Clinical Evaluation of Two Enzyme-Linked Immunosorbent Assays and Two Rapid Diagnostic Tests for Diagnosing Dengue Fever. *Journal of clinical microbiology*, 2015; **53**(4):1092-102.
- 59. Nga, T.T.T., Thai, K.T., Phuong, H.L., Giao, P.T., Hung, L.Q., Binh, T.Q., Mai, V.T.C., Van

Nam, N., de Vries, P.J. Evaluation of two rapid immunochromatographic assays for diagnosis of dengue among Vietnamese febrile patients. *Clinical and Vaccine Immunology*, 2007; **14**(6):799-801.

- Moorthy, M., Chandy, S., Selvaraj, K., Abraham, A. Evaluation of a rapid immunochromatographic device for the detection of IgM & IgG antibodies to Dengue viruses (DENV) in a tertiary care hospital in South India. *Indian journal of medical microbiology*, 2009; 27(3):254-6.
- 61. Sánchez-Vargas, L.A., Sánchez-Marce, E.E., Vivanco-Cid, H. Evaluation of the SD BIOLINE dengue duo rapid test in the course of acute and convalescent dengue infections in a Mexican endemic region. *Diagnostic microbiology and infectious disease*, 2014; **78**(4):368-72.
- Johnson, S., Krauss, T.F: Label-free affinity biosensor arrays: novel technology for molecular diagnostics, Taylor & Francis, 2017.
- Rashid, J.I.A., Abdullah, J., Yusof, N.A., Hajian, R. The development of silicon nanowire as sensing material and its applications. *Journal of Nanomaterials*, 2013; 2013(2013). http://dx.doi. org/10.1155/2013/328093
- 64. Gao, J., Jeffries, L., Mach, K.E., Craft, D.W., Thomas, N.J., Gau, V., Liao, J.C., Wong, P.K. A Multiplex Electrochemical Biosensor for Bloodstream Infection Diagnosis. SLAS TECHNOLOGY: Translating Life Sciences Innovation, 2017; 22(4):466-74.
- 65. Omar, A., Rashid, J.I.A., Latif, A.A.A., Karim, K.A., Bakar, O.C., Bakar, M.A., Yunus, W.M.Z.W. Development of cortisol immunosensor based reduced graphene oxide (rGO) for future application in monitoring stress levels among military personnel. Defence S and *T Technical Bulletin*, 2017; 10(2):142-148.
- Lerner, M.B., Dailey, J., Goldsmith, B.R., Brisson, D., Johnson, A.C. Detecting Lyme disease using antibody-functionalized singlewalled carbon nanotube transistors. *Biosensors* and Bioelectronics, 2013; 45:163-67.
- 67. Sarangadharan, I., Hsu, C.-P., Chu, C.-H., Regmi, A., Chen, Y.W., Wang, Y.-L. Blood based biomarker detection using FET biosensor: towards self-health management. *ECS Transactions*, 2017; 77(7):11-15.
- García, A.A., Franco, L.S., Pirez-Gomez, M.A., Pech-Pacheco, J.L., Mendez-Galvan, J.F., Machain-Williams, C., Talavera-Aguilar, L., Espinosa-Carrillo, J.H., Duarte-Villaseñor, M.M., Be-Ortiz, C. Feasibility study of an optical caustic plasmonic light scattering sensor for human Serum Anti-Dengue Protein E Antibody Detection. *Diagnostics*, 2017; 7(3):47.

- Xu, X., Liu, X., Li, Y., Ying, Y. A simple and rapid optical biosensor for detection of aflatoxin B1 based on competitive dispersion of gold nanorods. *Biosensors and Bioelectronics*, 2013; 47:361-67.
- Su, L., Zou, L., Fong, C.-C., Wong, W.-L., Wei, F., Wong, K.-Y., Wu, R.S., Yang, M. Detection of cancer biomarkers by piezoelectric biosensor using PZT ceramic resonator as the transducer. *Biosensors and Bioelectronics*, 2013; 46:155-61.
- Priye, A., Bird, S.W., Light, Y.K., Ball, C.S., Negrete, O.A., Meagher, R.J. A smartphonebased diagnostic platform for rapid detection of Zika, chikungunya, and dengue viruses. *Scientific Reports*, 2017; 7. doi:10.1038/srep44778.
- Oliveira, N., Souza, E., Ferreira, D., Zanforlin, D., Bezerra, W., Borba, M.A., Arruda, M., Lopes, K., Nascimento, G., Martins, D. A Sensitive and Selective Label-Free Electrochemical DNA Biosensor for the Detection of Specific Dengue Virus Serotype 3 Sequences. *Sensors*, 2015; 15(7):15562-77.
- Senapati, S., Slouka, Z., Shah, S.S., Behura, S.K., Shi, Z., Stack, M.S., Severson, D.W., Chang, H.-C. An ion-exchange nanomembrane sensor for detection of nucleic acids using a surface charge inversion phenomenon. *Biosensors and Bioelectronics*, 2014; 60:92-100.
- Deng, J., Toh, C.-S. Impedimetric DNA biosensor based on a nanoporous alumina membrane for the detection of the specific oligonucleotide sequence of dengue virus. *Sensors*, 2013; 13(6):7774-85.
- Rai, V., Hapuarachchi, H.C., Ng, L.C., Soh, S.H., Leo, Y.S., Toh, C.-S. Ultrasensitive cDNA detection of dengue virus RNA using electrochemical nanoporous membrane-based biosensor. *PloS one*, 2012; 7(8):e42346.
- 76. Souza, E., Nascimento, G., Santana, N., Danielly, F., Lima, M., Natividade, E., Martins, D., Lima-Filho, J. Label-Free Electrochemical Detection of the Specific Oligonucleotide Sequence of Dengue Virus Type 1 on Pencil Graphite Electrodes. *Sensors*, 2011; **11**:5616-29.
- Zhang, G.-J., Zhang, L., Huang, M.J., Luo, Z.H.H., Tay, G.K.I., Lim, E.-J.A., Kang, T.G., Chen, Y. Silicon nanowire biosensor for highly sensitive and rapid detection of Dengue virus. *Sensors and Actuators B: Chemical*, 2010; 146(1):138-44.
- Zaytseva, N.V., Montagna, R.A., Baeumner, A.J. Microfluidic biosensor for the serotype-specific detection of dengue virus RNA. *Analytical chemistry*, 2005; 77(23):7520-27.
- 79. Baeumner, A.J., Schlesinger, N.A., Slutzki, N.S., Romano, J., Lee, E.M., Montagna,

R.A. Biosensor for dengue virus detection: sensitive, rapid, and serotype specific. *Analytical chemistry*, 2002; **74**(6):1442-48.

- Rashid, J.I.A., Yusof, N.A., Abdullah, J., Hashim, U., Hajian, R. A novel disposable biosensor based on SiNWs/AuNPs modifiedscreen printed electrode for dengue virus DNA oligomer detection. *IEEE Sensors Journal*, 2015; 15(8):4420-27.
- Rashid, J.I.A., Yusof, N.A., Abdullah, J., Hashim, U., Hajian, R. Surface modifications to boost sensitivities of electrochemical biosensors using gold nanoparticles/silicon nanowires and response surface methodology approach. *Journal* of materials science, 2016; 51(2):1083-97.
- Mir, M., Katakis, I., Vreeke, M. Aptamer biosensors: an alternative to immunosensors. *IVD Technology*, 2007; 13(4):
- 83. Rashid, J.I.A., Yusof, N.A. The strategies of DNA immobilization and hybridization detection mechanism in the construction of electrochemical DNA sensor: A review. *Sensing and Bio-Sensing Research*, 2017; **16**:19-31.
- Rashid, J.I.A., Yusof, N.A., Abdullah, J., Hashim, U., Hajian, R. The utilization of SiNWs/AuNPsmodified indium tin oxide (ITO) in fabrication of electrochemical DNA sensor. *Materials Science* and Engineering: C, 2014; 45: 270-76.
- Kwakye, S., Goral, V.N., Baeumner, A.J. Electrochemical microfluidic biosensor for nucleic acid detection with integrated minipotentiostat. *Biosensors and Bioelectronics*, 2006; 21(12):2217-23.
- Nascimento, H.P., Oliveira, M.D., de Melo, C.P., Silva, G.J., Cordeiro, M.T., Andrade, C.A. An impedimetric biosensor for detection of dengue serotype at picomolar concentration based on gold nanoparticles-polyaniline hybrid composites. *Colloids and Surfaces B: Biointerfaces*, 2011; 86(2):414-19.
- Giakoumaki, E., Minunni, M., Tombelli, S., Tothill, I.E., Mascini, M., Bogani, P., Buiatti, M. Combination of amplification and postamplification strategies to improve optical DNA sensing. *Biosensors and Bioelectronics*, 2003; 19(4):337-44.
- Chua, A., Yean, C.Y., Ravichandran, M., Lim, B., Lalitha, P. A rapid DNA biosensor for the molecular diagnosis of infectious disease. *Biosensors and Bioelectronics*, 2011; 26(9):3825-31.
- Bora, U., Sett, A., Singh, D. Nucleic Acid Based Biosensors for Clinical Applications. *Biosens J*, 2013; 1;104. DOI: 10.4172/2090-4967.1000104.
- 90. Figueiredo, A., Vieira, N.C., Dos Santos, J.F., Janegitz, B.C., Aoki, S.M., Junior, P.P., Lovato,

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R.L., Nogueira, M.L., Zucolotto, V., Guimarães, F.E. Electrical detection of dengue biomarker using egg yolk immunoglobulin as the biological recognition element. *Scientific reports*, 2015; **5**. doi:10.1038/srep07865

- Dias, A.C.M., Gomes-Filho, S.L., Silva, M.M., Dutra, R.F. A sensor tip based on carbon nanotube-ink printed electrode for the dengue virus NS1 protein. *Biosensors and Bioelectronics*, 2013; 44:216-21.
- 92. Atias, D., Liebes, Y., Chalifa-Caspi, V., Bremand, L., Lobel, L., Marks, R.S., Dussart, P. Chemiluminescent optical fiber immunosensor for the detection of IgM antibody to dengue virus in humans. *Sensors and Actuators B: Chemical*, 2009; **140**(1):206-15.
- Mikkelsen, S.R. Electrochecmical biosensors for DNA sequence detection. *Electroanalysis*, 1996;

**8**(1):15-19.

- 94. Hayat, A., Marty, J.L. Aptamer based electrochemical sensors for emerging environmental pollutants. *Frontiers in chemistry*, 2014; **2**:41.
- Han, K., Liang, Z., Zhou, N. Design strategies for aptamer-based biosensors. *Sensors*, 2010; 10(5):4541-57.
- Bagni, G., Osella, D., Sturchio, E., Mascini, M. Deoxyribonucleic acid (DNA) biosensors for environmental risk assessment and drug studies. *Analytica chimica acta*, 2006; 573:81-89.
- 97. Wang, J., Rivas, G., Cai, X., Palecek, E., Nielsen, P., Shiraishi, H., Dontha, N., Luo, D., Parrado, C., Chicharro, M. DNA electrochemical biosensors for environmental monitoring. A review. *Analytica Chimica Acta*, 1997; **347**(1):1-8.