

Phylogenetic Assessment of *Garcinia* Species Using RAPD Markers

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The plants in the *Garcinia* species are economically important. Six varieties of *Garcinia* species were investigated for their phylogenetic relationship using Random Amplified Polymorphic DNA (RAPD) markers. Phylogenetic investigation is needed for these tree species to boost breeding and conservation programmes. A standardised procedure was developed for isolation of DNA from the leaf samples of *Garcinia cambogia*, *Garcinia indica*, *Garcinia xanthochymus*, *Garcinia morella*, *Garcinia mangostana* and *Garcinia livingstonei*. The DNA samples were subjected to PCR using 8 random primers. 269 polymorphic bands were obtained and scored to develop the values for the genetic distance. The dendrogram was developed using the software dendroUPGMA and the Cophenetic correlation coefficient of 0.801 is obtained. *G. Cambogia* and *G. livingstonei* are closely placed with a score of 24% followed by *G. morella*. It had a 30% index score to *G. cambogia* and *G. livingstonei* but is followed by just 31% score with *G. indica*. *G. mangostana* is connected at 33.5% dissimilarity to the above groups showing it is an introduced variety. *G. xanthochymus* is the last link with 37% score in the matrix. The genetic relatedness among these species is reported and this can be utilised in marker analysis for other *Garcinia* species.

Keywords: *Garcinia*; RAPD; phylogenetic analysis.

Garcinia is a group of plant species comprising of 250 species belonging to family Clusiaceae. The species are well distributed in the Asian continent. The species of *Garcinia* in India are distributed in the Western Ghats and the North East regions. Of the 30-35 species reported in India from the Western Ghats, few are endemic in nature. The endemic varieties are mostly trees and are very important plant species of the forests. Cultivation of a few species is seen in Kerala and Karnataka states. These trees contain fruits that are

utilised for the economical and pharmacological properties. *Garcinia cambogia*, *Garcinia indica*, *Garcinia xanthochymus*, *Garcinia morella*, *Garcinia mangostana* and *Garcinia livingstonei* are the common varieties that are seen distributed in these regions. *Garcinia mangostana* is an introduced variety from the Indonesian region. *Garcinia cambogia* (Syn. *G. gummi-gutta*) are medium sized trees and the fruits are used to extract Hydroxycitric acid. This is used in anti-obese drug formulations. The fruits from *Garcinia indica*, also

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a medium sized tree is used in culinary, cosmetics and pharmaceutical formulations . *Garcinia xanthochymus* contains important phenolic compounds used in anti-cancer treatment . *Garcinia morella* is utilised for a phytochemical: morellin . *Garcinia mangostana* was introduced in India for its importance as an edible fruit . *Garcinia livingstonei* fruits contain bioflavonoids that are used in treatment for colon cancer in humans .

Randomly Amplified Polymorphic DNA (RAPD) markers are PCR-based markers that amplify random regions on DNA segments. The primers used are short, the techniques are fast, and it uses very less DNA as template. There is no prior knowledge needed of the sequences from these species . PCR involves a procedure to amplify specific segments of DNA. A combination of RAPD primers in PCR help us to get amplified DNA segments from the plant template DNA provided for the investigation. The plants in the *Garcinia* genus of late has been investigated in isolated populations for their phylogenetic relationships . The molecular data is used to validate endemicity of varieties and to rule out possible introductions in the past. Phylogenetic analysis among the species can help researchers understand evolutionary pathways of genes, genomes and species by utilizing genetic differences.

There are different varieties of *Garcinia cambogia* being cultivated in different regions . Breeding programs have been initiated in various areas. Selection based on morphological data is tedious and time taking. Several attempts have been made to identify and develop SCAR based markers for sex determination and plant identification .

The objective of the present investigation was to identify genetic relationship among the six species using RAPD markers. Polymorphism data was analysed and genetic relatedness among the six species was recorded. DNA extraction and PCR protocols have been standardised for *Garcinia indica* and this procedure can be adapted to tree species . The standardisation of DNA extraction, PCR protocols, ISSR and RAPD marker analysis in other *Garcinia* species is also investigated. Genetic diversity in eight *Garcinia cambogia* genotypes was analysed by PCR-RAPD method and genetic relatedness and diversity has been recorded .

Genetic diversity relates to understanding the species ability to adapt to the environment . In the present investigation bands obtained from the PCR-RAPD gels were scored and a dendrogram constructed based on the unweighted pair group method with arithmetic mean (UPGMA) clustering method is developed. Genetic relatedness among the selected species of *Garcinia* is obtained and further investigations involving DNA that can be carried out is reported.

MATERIAL AND METHODS

Collection of DNA from leaf samples

The trees of *Garcinia cambogia*, *Garcinia indica*, *Garcinia xanthochymus*, *Garcinia morella*, *Garcinia mangostana* and *Garcinia livingstonei* used in the study were located and the GPS coordinates of the collection spots are recorded in Table 1. The collection areas recorded are from sites which are close to each other. The collection herbarium is recorded at the department of Biotechnology, St. Joseph's college, Bangalore. Leaf samples were collected and stored in liquid nitrogen.

DNA extraction

500 mg of the leaf sample were homogenised using tissue homogenizer with 15ml of lysis buffer consisting of 2% Hexadecyltrimethylammonium bromide (CTAB), 100mM Tris pH 8, 20mM EDTA, 1.4M NaCl, 2% polyvinylpyrrolidone 40 (PVP) and 0.2% Beta-mercaptoethanol. The extract was transferred to a tube and were incubated at 65°C for 1 hour in a water bath. A set of six tubes with the six species was obtained. The tubes are centrifuged at 10000 rpm for 10 minutes. Supernatant was transferred to a fresh tube. 12ml is recovered from the supernatant. Equal volume of Chloroform was added, and the tubes are mixed. The tubes are then centrifuged at 10000 rpm for 15 minutes. The aqueous layer is pipetted out into fresh centrifuge tube without disturbing the interface. Equal volume of Isopropanol and 2 ml of 3M Sodium acetate was added. The contents with the tube are kept at room temperature for 10 minutes. The tube is then centrifuged at 10000 rpm for 15 minutes. The supernatant was discarded. The pellet was washed with 600µl of 75% ethanol. The pellet air dried and stored in 600µl of 1X Tris- EDTA buffer. The

method was used to extract the DNA from all the six species of *Garcinia* leaf samples.

Column purification

Plant genomic purification kit, with the catalogue no 2115700021730 from Genei laboratories, Bangalore India was used to purify the extracted DNA samples. The quality of isolated genomic DNA was checked by Agarose gel electrophoresis. 2µl of DNA was mixed with 1µl of 1X loading dye and loaded in a slot of 0.8% agarose gel containing 0.015 µg/ml of Ethidium Bromide. The purified samples were subjected to RAPD-PCR reactions.

PCR-RAPD analysis

The PCR reaction was set with the components detailed in table 2. 37µl of this reaction

mixture was aliquoted into 48 different labeled PCR vials and to this 2µl of different template DNA and 1µl random primer were added. The details of the eight RAPD primers used is detailed in table 3. The PCR was set in Eppendorf Mastercycler Gradient unit with reaction conditions detailed in table 4. After the reaction the components in the vial were loaded on agarose gels.

Detection by Agarose gel electrophoresis

The RAPD-PCR products were separated based on their molecular weight, by agarose gel electrophoresis. 1% agarose gel with 0.015 µg/ml of Ethidium Bromide were used. The DNA were run using electrophoresis buffer and in electrophoresis units and observed under gel documentation units. DNA ladder ranging from

Table 1. GPS coordinates of plant specimens.

| Plant species | GPS coordinates | |
|------------------------------|-----------------|--------------|
| <i>Garcinia cambogia</i> | 12°05'26.0"N | 76°02'03.1"E |
| <i>Garcinia indica</i> | 12°05'30.8"N | 76°01'59.3"E |
| <i>Garcinia xanthochymus</i> | 12°05'33.0"N | 76°01'48.5"E |
| <i>Garcinia morella</i> | 11°59'41.5"N | 76°04'01.9"E |
| <i>Garcinia mangostana</i> | 11°59'40.2"N | 76°04'02.6"E |
| <i>Garcinia livingstonei</i> | 12°05'24.9"N | 76°01'56.5"E |

Table 2. PCR reaction components

| Components | Master mix | |
|-------------------|------------|--------|
| | 1X | 48X |
| D.D.H2O | 17µl | 816µl |
| 2X PCR Master MIX | 20 µl | 960µl |
| Random Primer | 1µl | 48µl |
| Template DNA | 2µl | 96µl |
| Total Volume | 40 µl | 1920µl |

Table 3. List of RAPD Primer with sequence details

| Sl.no | Primers | Sequence |
|-------|---------|------------|
| 1. | OPA-02 | TGCCGAGCTG |
| 2. | OPB-10 | CTGCTGGGAC |
| 3. | OPD-02 | GGACCCAACC |
| 4. | OPC-06 | GAACGGACTC |
| 5. | OPD-08 | GTGTGCCCCA |
| 6. | OPC-07 | GTCCCGACGA |
| 7. | OPB-07 | GGTGACGCAG |
| 8. | OPB-08 | GTCCACACGG |

0.1, 0.2, 0.3, 0.6, 1.5, 2, 2.5, 3 and 3.5kb. was used as molecular size marker. The gel was documented in Gel documentation unit and photographs of gels displaying the bands were taken for data analysis.

Data analysis

Each amplified product from the gel was scored as a unit character and the populations were recorded for the presence (1) or absence (0) of a band. By using the concept of calculating all possible pair-wise genetic distances, the values were obtained from the following formula: $D_{ab} = 1 - (2n_{ab} / (n_a + n_b))$, Where, n_a and n_b are the

Table 4. PCR Cycle conditions

| Temperature | Time | No. of cycles |
|-------------|--------------|---------------|
| 94°C | 5 minutes | 1 |
| 94°C | 30 second | 40 |
| 45°C | 1 minute | |
| 72°C | 1.30 minute | |
| 72°C | 7 minutes | 1 |
| 4°C | Till loading | |

numbers of bands amplified in individuals a and b respectively and 2nab is the number of bands shared by those individuals. The percent disagreement between the populations was calculated based on Unweighted Pair Group Method with Arithmetic Mean (UPGMA). The resulting dissimilarity index was used to evaluate the relationship among various populations of this species with cluster analysis, using Unweighted pair-group average. All computations were carried out using the phylogenetic software, DendroUPGMA, . The molecular size of the amplified products is recorded for unique bands of DNA fragments by comparing with the 0.1, 0.2, 0.3, 0.6, 1.5, 2, 2.5, 3 and 3.5kb ladder.

RESULT AND DISCUSSION

The DNA was successfully extracted from all the six *Garcinia* species. The extracted DNA samples were column purified and checked on agarose gel (Figure 1). RAPD amplicons were resolved on agarose gels (Figure 2,3). A total of 271 bands were obtained and scored from 8 primers of which, 269 were polymorphic (Table 5). The results showed 98% polymorphic

bands for the primer OPD-02 and 100% for the other primers. The only monomorphic band was of 2.4kb and was obtained from the OPD-02 primer having a sequence GGACCCAACC. The scoring data from the bands were analysed for the values for pair wise distance matrix calculation. The generated values are reported in a distance matrix format in Table 6. From the values of the distance matrix, the software DendroUPGMA generated the dendrogram as represented in Figure 4. The dendrogram is also represented in Newick format, a way of representing dendrograms as a text, using parentheses and commas. All the tree-visualization programs accept this format. The Newick representation for the dendrogram obtained is: (((((Gu:0.243,li:0.243):0.067,mo:0.309):0.009,in:0.318):0.017,ma:0.335):0.034,xa:0.370). The Cophenetic correlation coefficient (CP) from the data of RAPD scoring for was found to be 0.80. The CP value for this dendrogram is a measure of how faithfully a dendrogram preserves the pairwise distances between the original unmodeled data points. It is a value between 0 and 1, where 1 represents a perfect match in the dendrogram representation. The value here is at 80% and is a good score for a dendrogram. The dendrogram

Table 5. Percentage polymorphism of primers

| Primers | No of bands -A | No of polymorphic bands -B | % polymorphism B/A x 100 |
|---------|----------------|----------------------------|--------------------------|
| OPA-02 | 21 | 21 | 100 |
| OPB-10 | 29 | 29 | 100 |
| OPD-02 | 50 | 49 | 98 |
| OPC-06 | 31 | 31 | 100 |
| OPD-08 | 28 | 28 | 100 |
| OPC-07 | 49 | 49 | 100 |
| OPB-07 | 26 | 26 | 100 |
| OPB-08 | 37 | 37 | 100 |

Table 6. Representation data of distance matrix obtained by pair wise distance values

| | <i>G.cambogia</i> | <i>G.mangostana</i> | <i>G.morella</i> | <i>G.indica</i> | <i>G.livingstonei</i> | <i>G.xanthochymus</i> |
|-----------------------|-------------------|---------------------|------------------|-----------------|-----------------------|-----------------------|
| <i>G.cambogia</i> | 0 | 0.763 | 0.662 | 0.65 | 0.486 | 0.794 |
| <i>G.mangostana</i> | | 0 | 0.653 | 0.663 | 0.603 | 0.714 |
| <i>G.morella</i> | | | 0 | 0.670 | 0.576 | 0.820 |
| <i>G.indica</i> | | | | 0 | 0.59 | 0.695 |
| <i>G.livingstonei</i> | | | | | 0 | 0.674 |
| <i>G.xanthochymus</i> | | | | | | 0 |

indicates that the species are divided into 2 clusters: one that is solely *G. xanthochymus* and all other species in the other cluster with a genetic distance of 37%. Within the cluster with all other species, *G. cambogia* and *G. livingstonei* are most similar to each other with a 24% dissimilarity index. Both the species produce hydroxy citric acid and

guttiferones as their main chemical component in fruits, *G. morella* and *G. indica* have a 30% and 31% dissimilarity index to both *G. cambogia* and *G. livingstonei* respectively. Both the plants have bright red fruits and are phenotypically similar. This is attributed to the presence of anthocyanin pigments in them. The oil from both the plants are utilised in cosmetic industries. *G. mangostana* is the least similar in the group with a dissimilarity index of 33.5% in the group. This variety is introduced to India and the dendrogram justifies the same. The research on the origin of the Malaysian varieties of *G. mangostana* says this species may be the first in origins among the *Garcinia* varieties. *G. xanthochymus* is the most dissimilar with a score of 37% and is placed by itself in the dendrogram. The phenotypic characters of this plant are very different to the group in terms of fruit and leaf size. The chemical composition of these fruits is exclusive

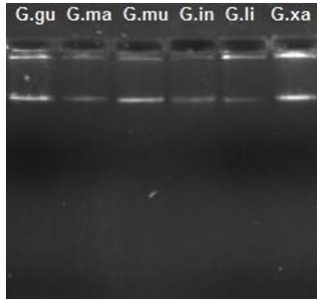


Fig. 1. DNA bands after column purification

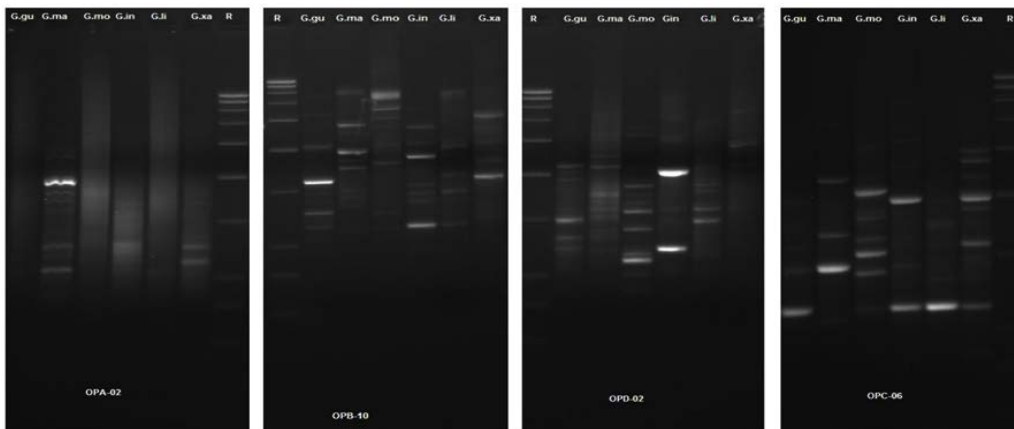


Fig. 2. RAPD profile generated by the PCR reactions from Primers-0PA-02; OPB-10;OPD-2;OPC-6

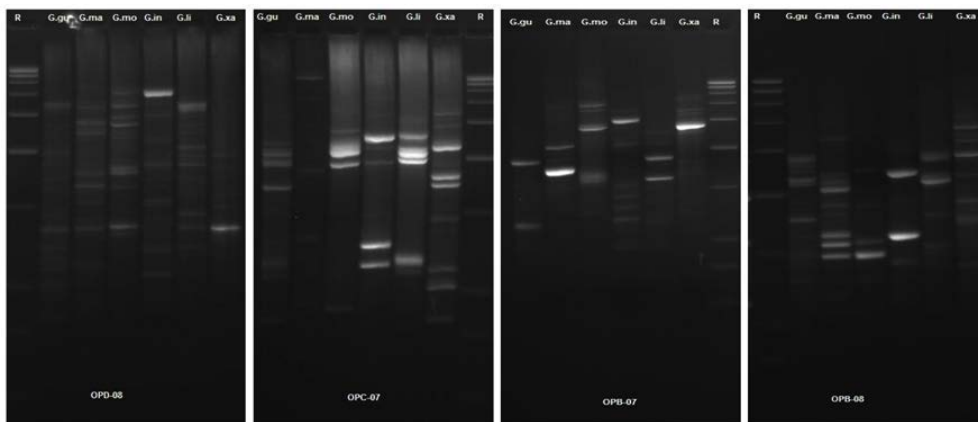


Fig. 3. RAPD profile generated by the PCR reactions from Primers:0PD-08; OPC-7; OPB-7; OPB-8

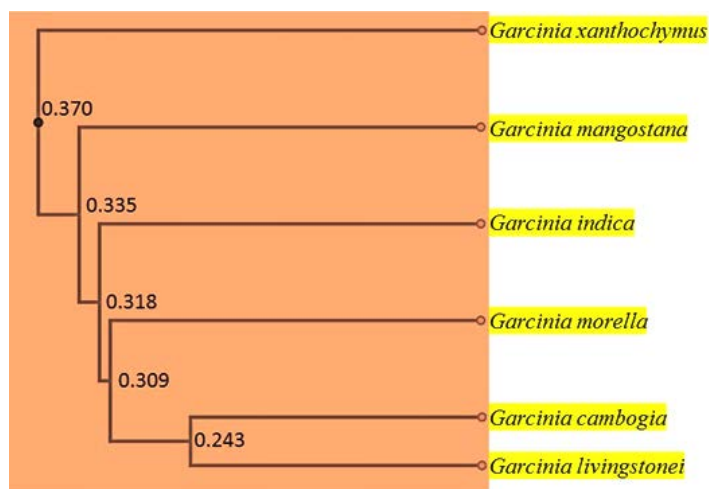


Fig. 4. UPGMA Dendrogram representation of relationship among various populations of *Garcinia* species

and includes xanthochymols. The dendrogram representation outlines the relationships between and among these plant species. The CP value also validates the accuracy of the analysis. With 98% of polymorphism in these markers, the sequences of the primers can be utilised for generating specific markers for future work. Genetic diversity study has shown relatedness to the phenotypic characterisation. The species being economically important, further investigation at the marker level can boost breeding and conservation programmes of these tree species. The genetic diversity studies can be expanded to the *Garcinia* varieties in India.

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