Acid Black 172 Dye Decolorization and Bioelectricity Generation by Microbial Fuel Cell with Filamentous Fungi on Anode

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A microbial fuel cell (MFC) has great potential for azo dyes decolorization and electricity generation by using filamentous fungi as biocatalysts. In this study, *Aspergillusniger* and *Trichodermaharzianum* were inoculated in anode chamber of double-chamber MFC to decolorize azo dye acid black 172 with Potassium Ferricyanide in the cathode chamber. During MFC operations, Acid black 172 oxidized and produced a maximum open-circuit voltage of 890 mV, and maximum current density of 163 mA/m² with an external resistance of 1000Ù. Also, variable parameters such as dye concentration, Co-substrate and dye as a sole carbon source were studied to improve microbial fuel cell performance.

Keywords: Microbial fuel cell, Decolorization, bioelectricity, Acid black 172.

Azo dyes are a group of poorly biodegradable synthetic colorants that are often found in the textile wastewater (Cai*et al.*,2014). These compounds are highly resistant to aerobic biodegradation by microorganisms but can be easily reduced by anaerobic microorganisms to form aromatic amines, which are a group of carcinogens that are stable under anaerobic conditions and must be returned to aerobic conditions before they can be further degraded and mineralized (Garcia *et al.*, 2015). A range of physicochemical methods exists to remove color from dye containing effluents (Mu *et al.*, 2004). The most extensively used are coagulation and flocculation processes. They require significant quantities of chemicals and produce notable amounts of sludge, requiring further handling and disposal. Enzymatic decolorization is now widely used for the decolorization of dye wastewater. However, this method is also facing several problems such as cost of enzymes, enzymes stability and product inhibition (Husain, 2010). On the other hand, biological processes provide a low-cost and efficient alternative for simultaneous color removal. In recent years, microbial fuel cell (MFC) technology has explored extensively for their innovative features and environmental benefits (Rozendalet al., 2009). This work presents an efficient and cost-effective approach to establish an MFC-assisted electrochemical oxidation process for the decolorization of azo dye.

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METHODOLOGY

Dyes

Five model textile dyes (obtained from Moket Mac Company at 10th of Ramadan City, Egypt) were used for testing decolorization capacity of *Aspergillusniger* and *Trichodermaharzianum* including: Acid red 399, Acid yellow 235, Acid yellow 218, Acid blue 296, and Acid black 172. **Sample collection and cultivation conditions**

A fungal species was collected from wastewater of an Egyptian Company for artificial Carpet at 10th of Ramadan City, in sterile clean glass bottles then, stored at 4°C, cultivated in petridishes containing Dox¹/₂s agar medium at 25°C and stocked in slant tubes containing Czapek¹/2s yeast agar solid medium under refrigeration at 4°C. Initial selection was based on maximum voltage production and dye decolorization. For submerged cultures, a pre-inoculum of young mycelium disk of 0.5cm of diameter was obtained from 2 daysold colonies for Aspergillusniger, while in case of Trichodermaharzianumtwo young mycelium disks of 0.5cm of diameter were obtained from 4 days-old colonies in solid medium. Disks were inoculated into flasks with 50 ml of fresh medium containing 15g/L glucose, 2g/L NaNo₃, 1g/L KH₂PO₄, 0.5g/L KCL and 5g/L yeast extract in 50 mM phosphate buffer (PH7) and incubated at 25 UC for 96h for Trichodermaharzianum, while Aspergillusniger was incubated at 35 UC for 48h. After this period of time, flasks were used for inoculation of 250 ml flasks (bottles) containing 50 ml Acid black 172 dye (which has been prepared using 10mg/L)of anodic compartment of fungal microbial fuel cells. Each culture was incubated under maximum incubation temperature.

Construction of fungal microbial fuel cell

The dual chambered MFC was constructed using two air tight laboratory glass bottles (anode and cathode) of 250 ml capacity connected via a glass tube (LengthЧ11 cm, width= 6.6 and diameterЧ0.5 cm) that is heated and bent into a u-shaped, filled with 2% agar gel in saturated KCL solution and inserted through the lid of each bottle. The anodic and cathodic electrodes were graphite rods (surface areaЧ5.5cm). Copper (Cu) wires were soldered to the electrodes by using a conductive epoxy. A multimeter with a data logger system (MT-1820 ProstKit, Taiwan) was used as the automated data acquisition system.

Operation of a fungal microbial fuel cell

The fuel cell compartments were cleaned and the electrodes were autoclaved for 20 min. The anode solution (50mL) contained the following components(per liter) 15g $C_6H_{12}O_6$, 2g NaNO₃, 1g KH₂PO₄, 0.5g KCL ,5g yeast extract in 50 Mm phosphate buffer(pH 7) and fungal biomasses. The compartment was inoculated with 50 ml acid black 194 which prepared as mentioned previously, finally anode volume (100 ml).

The cathode solution on the other hand, consisted of 100ml of Potassium Ferricyanide $(K_3(Fe(CN)_6)$ solution (10mmol/L)prepared in distilled water. The initial pH of Potassium Ferricyanidesolution (pH 2) at cathode was adjusted using 0.5M HCLwhich acted as the electron acceptor.

The MFCs with inoculation were first incubated with electrolytes described above for 4 days, then evaluated the decolorization rate of Acid black 172 and electricity generation. The external resistance was fixed at 1000&! and in some case the MFCs were operated under different external resistances to obtain the polarization curves. The temperature of MFCs was kept at 35°C for *Aspergillus niger* and at 25°C for *Trichodermaharzianum* using the laboratory incubators both the two inoculated MFCs and the two control sets were operated under identical conditions respectively, and average results were reported.

Electrochemical analyses

Cell voltages were recorded every 20 minute after a stable open circuit potential was achieved using a data logger system. Current (*I*) was calculated at a resistance(*R*) from the voltage (V) by I=V/R, and current density (I/A²), where A is the projected anode surface area.

Acid black 172 decolorization

The absorption of A.black 172 in the visible light range peaks at a wavelength of 572 nm was measured using a T60 UV-VIS spectrophotometer and used to calculate A.black 172 from Eq. (1) where A_0 and A are the initial and final absorptions, respectively.

Decolorization (%) = $(A_{a} \times A)/X 100$

RESULTS AND DISCUSSION

Decolorization of dyes using Aspergillus niger and Trichoder maharzianum

Aspergillus niger and Trichoderma harzianum were selected to investigate their effect on decolorization rate of the following dyes:(1) Acid red 399, Acid yellow 235, Acid yellow 218, Acid blue 296, and Acid black 172. The initial concentration was (10mg/L) amended in flasks containing 100 ml of Dox's broth medium. The flasks were inoculated as previously mentioned. The results in Fig (2) show variation in decolorization abilities. Aspergillusniger was able to remove more than 50% of these acid dyes. On other hand, *Trichodermaharzianum*was able to remove less than 50% of acid red 399,acid yellow 235, acid yellow 218 and acid blue 296 dyes. The results showed that,*Aspergillusniger*and *Trichodermaharzianum* were effectively able to remove more than 85% of acid black 172. Accordingly, it is selected for further investigations. These results may be due to chemical structures of dyes significantly affecting the performance of dye biodecolorization. For similar structures of azo dyes, monoazodyes are easily biodecolrizedthan polyazo dyes (Hsueh*et al.*, 2009).Also, our results agree with Mohamed*et al.*,20105 who state that

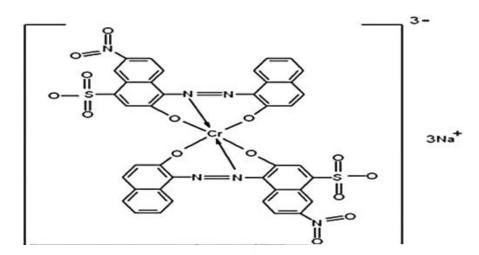


Fig. 1.

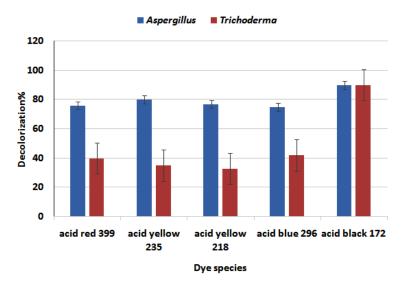


Fig. 2. Decolorization of dyes using Aspergillusniger and Trichodermaharzianum

the best decolorizing dye was acid black 194 by *Aspergillusflavus*, *Aspergillustamarii* and *Aspergillusparasiticus*.

Acid black 172 concentration effect ondye decolorization and voltage output (OCV)

Comparison on performance of voltage output in a fungal microbial fuel cell at various concentrations of acid black 172 (i.e., 5, 10, 30 and 50mg/L; Table.1) showed that the decolorization percent and voltage output(OCV) for *Aspergillusniger* and *Trichodermaharzianum*reached its maximum value (95%) and (890mV) respectively at dye concentration 10mg/L. Also showed that increasing in the dye concentration resulted in a decrease in the decolorization percent and voltage output (OCV). The results are in agreement with the findings of

Boret al.(2010)that observed the decreased peak output voltage at high loading of Reactive Blue 160 might explain that decolorization and bioelectricity generation of Proteus hauseriare competitive to each other as both reactions all utilized electrons released from bacterial oxidation of organic matter. In addition, possibly due to accumulation of the decolorized intermediates of Reactive Blue 160 as mediators, the steady-state (SS) cell-voltages were stabilized at slightly higher levels than the SS-voltage in Reactive blue 160 free cases. In general, the performance of an MFC depends on the concentration and the type of dye used; Mu et al. (2009) showed that during closed-circuit operation, decolorization efficiency decreased from 78.7% to 35% with an increase in influent dye concentration. Sun et al. (2009) reported that

 Table 1. Acid black 172 concentration effect on dye decolorization and voltage output (OCV)

Dye concentration	Decolorization (%)		Voltage(OCV)	
(mg/L)	Aspergillus	Trichoderma	Aspergillus	Trichoderma
5	93	94	850 mV	870 mV
10	95	95	890 mV	890 mV
30	80	75	810 mV	800 mV
50	65	61	750 mV	720 mV

 Table 2. Co-substrate effect on decolorization

 of Acid black172 and voltage output (OCV)

Co-substrate	Decolorization(%D)		Voltage (OCV)	
	Aspergillus	Trichoderma	Aspergillus	Trichoderma
Glucose	95	98	899 mV	895 mV
Sucrose	90	92	820 mV	800 mV
Control	80	75	700 mV	620 mV

Table 3. Simultaneous dye decolorization and current generation

Time (I		Decolorization (%)		Current density (mA/m ²)	
	Aspergillus	Trichoderma	Aspergillus	Trichoderma	
0	95	94	163	160	
12	93	94	149	154	
24	92	93	147	154	
36	92	92	145	145	
48	90	90	145	136	
60	90	90	136	131	
72	88	85	133	129	

the percent decolorization decreased with increase in Active Brilliant Red X-3B concentration.

Co-substrate effect on decolorization of Acid black 172 and voltage output (OCV)

Different co-substrates such asglucose, and sucrosewere individually added as equimolecular to 30g sucrose per liter were tested to determine the optimum co-substrate which supported the maximum decolorization and voltage output of the tested fungi. The results in Table (2) showed that, the decolorization percent for Aspergillusniger and Trichodermaharzianum reached its maximum value(95% and 98%) respectively using glucose as co-substrate, also showed voltage output value reached its maximum value (899 mV and 895 mV) for Aspergillusniger and Trichodermaharzianum respectively. The results similar to those obtained by Sun etal. (2009) who stated that, the maximum decolorization rate of ABRX3 observed with glucose, also in agreement with our results Cao et al. (2010) found that, the power density was highest for glucose during decolorization of Congo red. Simultaneous dye decolorization and current generation

Dye decolorization rate and current generation weremeasured after operation for three days. The results in Table (3) showed that decolorization rate and current density were decreased after sixty hours of operation. These results in contrast with Chen (2006) and Chen *et al.*(2009) who mentioned that impulse additions of C.I. reactive red 141 significantly enhanced increasing the rate of oxidative phosphorylation of*P.hauseri*to accelerate electron transport in the respiratory chain of immobilized cells on the anodic biofilm for bioelectricity production in MFC.

CONCLUSION

The microbial fuel cell (MFC) with a fungal anode achieved more than 84% decolorization of azo dye acid black 172 in three days of continuous operation with current density more than 128 MA/ m².Also, a maximum open circuit voltage(OCV) was obtained with 10 g/L of dye concentration was 890 mV and maximumopen circuit voltage(OCV) was obtained with Co-substrate with glucose was more than 894mV for the two fungal species.

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