

CCR5 as a Novel Cell and Gene Therapy Strategies Based on Induction of Resistance to HIV

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Cc-chemokine receptor-5 (CCR5) is known as a main co-receptor in human immunodeficiency virus-1 (HIV-1) infection. So, it could be a target for inhibition of HIV-1 entry into CD 4⁺ immune cells. Many studies showed homozygote individual with 32bp deletion in *CCR5* gene had nature resistance to HIV-1. In this manner, recent treatments are focused on inducing this resistance to HIV-1 infected patients with *CCR5*. Berlin and Boston patients transplanted with allogeneic hematopoietic stem cell (HSC) and demonstrated effective cure for HIV-1 infection. In addition, zinc finger nuclease (ZFN) eliminated some problems of Berlin and Boston patients by site-specific *CCR5* gene modification. These recent strategies declined highly-active anti-retroviral therapy (HAART) restrictions such as toxicity, low safety, the side effects following long-term consuming and virus reloading immediately after cut the drugs off. In this review, in addition of introductory biologic and immune-genetic roles of CCR5, we consider novel treatment strategies for HIV-1 infected patient by *CCR5* gene targeted therapy.

Key word: CCR5, HIV-1, hematopoietic stem cell therapy, gene therapy, gene modification, nuclease.

It is demonstrated that a cc-chemokine receptor (CCR)-based targeted therapy can play critical role in more effective cure of patients infected by human immune-deficiency virus-1 (HIV-1)¹⁻⁴. CCRs are a group of G-protein coupled family of receptors (GPCR) and approximately 800 genes have been introduced that encoded functional GPCR and made up about % 1 of human genome^{5, 6}. It is demonstrated that about half of clinical drugs design for GPCRs, with blocking their ligands or increasing ligands accessibility. So their roles in our bodies are very important⁷.

Human GPCRs consist of six families whose major groups are A, B and C and their largest group is A family. One of the most popular

subfamilies of class A is CCR5 which for many years has attracted scientists. The vast majority of investigations have focused to provide the novel approaches to the treatment of HIV based on targeted therapy. We have previously suggested induction of resistance to HIV by *CCR5* gene therapy and stem cell transplantation^{8, 9}. Here we present most recently discovered approaches to the treatment of HIV, based on CCR5, which may define the better treatment, especially target therapy.

CCR5 genetic basis, structure and molecular signaling

CCR5 is encoded by *CMKBR5* gene located on p21.31 region of human chromosome 3¹⁰. Protein product of this gene includes 3 sections^{11, 12}: 1) A seven helical trans-membrane domain which provide 3 extra-cellular and 3 intra-cellular hydrophilic loops. 2) C-terminal residue that

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regulates receptor by serine-threonine phosphorylation. 3) N-terminal residue that is bound to ligand. MIP 1 α , β (also known as CCL3, CCL4), RANTES (also known as CCL5) and the other β -chemokines are ligands that are bound to NT site in CCR5^{12,13}. After ligand binding to NT or 3 extracellular loops, a molecular mechanism activate that cause receptor conformation change and the dissociation of G-protein subunits. G α -GTP and G $\beta\gamma$ subunits are freed and regulate enzymes activity such as adenylate cyclase, phospholipase C isoforms and ion channels. These enzymes regulate protein kinase function, finally increase intracellular ca²⁺ ions and stimulate chemotaxis¹⁴. Pharmacologist have named GPCR as “spare receptor” because full biologic response

happen after 5% occupancy¹⁵. Thus GPCRs associate with cell metabolism, growth, migration, differentiation and death of multiple cell type (apoptosis)^{16, 17}.

Resistance to HIV-1 infection by CCR5

Any change in the sequence of gene variants causes alternation in HIV infection or AIDS process, named as AIDS Restricted Gene (ARG). Some of these variants are detected in special group population which causes natural selection and/or genetic drift different ARG polymorphism/mutation. To this date, more than 35 ARGs have been identified. First of them is CCR5 Δ 32 and¹⁸⁻²². This 32bp deletion (CCR5 Δ 32) in the single-coding exon provides resistance mechanism to HIV/AIDS infection^{12, 23}. CCR5 Δ 32 is caused by replicative

Table 1. CCR5 polymorphisms that affected HIV-1infection

Type of polymorphism	HeterozygoteOr Homozygote	Effect on HIV/AIDS
CCR5* Δ 32/ Δ 32	Homozygote	Create resistance to HIV-1 infection.
CCR5 Δ 32/WT	Heterozygote	Infected by HIV-1 but delay to AIDS for 2-4 years.
CCR5 Δ 32/**m303	Heterozygote	Create resistance to HIV-1 infection.

*CCR5 Δ 32: 32bp deletion in exon 1 CCR5 gene that cause second extracellular loop be defected and truncate protein created.

**CCR5 m303 (C101X): Transversion of T to A in 303 nucleotide open reading frame that created nonsense mutation in 101 amino-acid at first extracellular loop CCR5.

Table 2. Some of the current drugs for HIV treatment

Drug(s)	Function	Reference
Maraviroc	HIV entry inhibitor (CCR5 antagonist)	[40]
Enfuvirtide	HIV entry inhibitor (fusion inhibitor)	[40]
Abacavir, Didanosine, Emtricitabine, Lamivudine, Stavudine, Tenofovir disoproxil fumarate (DF), Zalcitabine, Zidovudine	Nucleotide reverse transcriptase (RT) inhibitors	[75]
Rilpivirine, Nevirapine, Etravirine, Efavirenz, Delavirdine	Non-nucleotide reverse transcriptase (RT) inhibitors	[75]
Raltegravir and Elvitegravir	Integrase inhibitor	[75]
Vorinostat	Histone deacetylase inhibitor	[76]
Atazanavir, Darunavir, Fosamprenavir, Indinavir, Lopinavir/ritonavir, Nelfinavir, Ritonavir, Saquinavir, Tipranavir	Protease inhibitor	[75]
Elvitegravir/Cobicistat/Emtricitabine/Tenofovir	combinations of reverse transcriptase and integrase inhibitors	[75]
Abacavir/Lamivudine	Reverse transcriptase inhibitors (fixed-	[75]
Abacavir/Lamivudine/Zidovudine	dose combinations of	
Emtricitabine/Tenofovir DF	nucleotide analogues)	
Lamivudine/Zidovudine	Reverse transcriptase inhibitors (fixed-	[75]
Efavirenz/Emtricitabine/Tenofovir DF	dose combinations of both types of inhibitors)	
Emtricitabine/Rilpivirine/Tenofovir DF		

slippage by RNA pol due to the presence of a direct repeat flanking the deleted region which leads to elimination of second extracellular loop on receptor. Therefore, HIV-1 enters to bloodstream but could not infect CD4⁺ T cell and macrophage¹². A number of studies have shown that individual with homozygote *CCR5Δ32* allele exhibits a natural resistance to HIV and Acquired immunodeficiency syndrome (AIDS)^{12,23,24}. It is also worth mentioning that heterozygote individual after exposing to human immunodeficiency virus display slower progression to AIDS than homozygote individual for the wild type allele^{25, 26}. In Northern Europe, Caucasian population indicate the highest *CCR5Δ32*, 20% of whom are heterozygote and 1% homozygote for this mutation, respectively²⁷. Because of these remarkable detections, CCR5 is known as a major co-receptor, even when immune-cells present CD4 on their surface. If CCR5 is not presented on cell surface (following knock-out it or when using its inhibitors) HIV-1 could not contaminate cells^{28,29}.

CCR5Δ32 is highly resistant to HIV-1 infection, but not completely. Since CXCR4 is another co-receptor for HIV-1 entries. So CCR5 is not the only co-receptor for HIV-1 infection and one of the reasons that HIV/AIDS have epidemiological heterogeneity is genetic variants in host receptor and co-receptors (CCR5 or CXCR4) that HIV can switch itself to per variant³⁰.

Resistance to HIV-1 infection or delay to AIDS process by Another polymorphisms

In addition to *CCR5Δ32*, Some other mutations/polymorphisms have been discovered which provide the resistance mechanism, such as *CCR5/m303* (table 1)³¹, *IDH1C*³². But although some of the others like *CCR5Δ32* heterozygote are susceptible to HIV infection, they delay the AIDS progression, such as *CCR2 64F*³³, *SDF1 3'A*²⁵, *HLA B*57*¹⁸, *HLA B27*³⁴, *KIR 3DS1*^{35, 36}, *PROX1 Hap-CGT*³⁷, *ACSM4 A*³⁸.

Thus, the variant of host genotype may change the function or alternate gene expression after HIV-1 contamination, known as HIV-dependency factors (HDFs). HDFs are needed for HIV-1 infection process, transmission, viral loading and challenge with immune-system³⁹. However, host genetic background is a part of factors that affect HIV-1 infection. The others are HIV acquisition, immune-system condition and

HAART (highly-active-anti-retroviral therapy) results.

Current therapeutic methods: benefits and limitations

To date, several drugs have been introduced for inhibition of AIDS progression, like enfuvirtide (T20) and maraviroc. Maraviroc is a co-receptor antagonist blocks interaction between CCR5 and envelope (*env*) protein coating HIV-1 surface⁴⁰. Enfuvirtides act as fusion inhibitor disrupting conformation change in glycoprotein-41 (gp41)⁴⁰. In addition to, highly active anti-retroviral therapy (HAART) is emerging which targets virus enzymes (Table 2). It is showed that ART have significant positive effect on the reduction of latent reservoir virus in immune system at an early stage or immediately after informing of HIV-1 infection. For example, when an infant was born from a mother that infected by HIV-1, after 30 hours of birth undergone ART and continued until 18 months. In the months 30, no pro-viral DNA or plasma RNA of HIV was detected in peripheral mononuclear cells⁴¹.

One of the problems in HIV-1 treatment by ART is “drugs resistance”. This phenomenon mainly caused by new mutation patterns in HIV-1 genes that virus need for its essential proteins like protease⁴². Hence, HAART could not eradicate virus, although reduce its replication significance. In addition, they should be used long time and maybe discontinue of them in any time cause RNA and DNA virus rebounding from latent reservoirs. Moreover, HAART cannot act in different individuals by the same efficiency. High cost and some side effects following long-term therapy are another restriction the use of HAART^{43, 44}.

Recent treatment strategies are focused on gene therapy especially against HIV and other refractory disease, we present some study in this field previously^{8, 9, 45-49}. Here, we present most recently discovered approaches to the treatment of HIV, based on CCR5, which may define the better treatment, especially target therapy.

Novel therapeutics approaches: stem cell transplantation and gene-modification

Novel treatment strategies with stem cell transplantation (SCT) overcome to some problems that observed in ART^{8,9}. Successful allogeneic SCT have performed for acute myeloid leukemia (AML) in a patient co-infected by HIV and HCV and had

been undergone HAART in 2002⁵⁰. Following this, Hutter *et al.* in 2009 transplanted allogeneic (*CCR5* $\Delta 32/\Delta 32$) stem cell as a treatment for “Berlin patient” who was suffering from AML and HIV infection¹. The patient was undergoing HAART for 10 years and discontinued them after SCT¹. Rebounding of HIV may be observed if HAART discontinued because of another CXCR4 co-receptor existence⁵¹, but in this patient after 20 months not observed signs of virus activating or replicating¹. Hutter *et al.* sequenced CXCR4 patient and checked its variants, but not detected any trail of CXCR4 and concluded HIV-1 in this patient was not binding with CXCR4 as a co-receptor. Number of CD 4⁺ T-cells after HSCT for *CCR5* $\Delta 32/\Delta 32$ increased such as normal range in health people and HIV DNA or RNA was undetectable gradually¹. Result of this treatment showed no existence of RNA or DNA virus after 3.5 years, even when HAART had been discontinued^{2, 52}. Likewise, it was reported Berlin patient body remained free of HIV-1 after 5.5 years⁵³.

In 2012, Henrich *et al.* performed HSCT for two patients, Boston patients, who have heterozygote genotype for *CCR5* (*CCR5* WT/*CCR5* $\Delta 32$) and transplanted with WT *CCR5* cells. Result of this study suggested that replication of DNA or RNA virus had been suppressed and HIV-1 reservoir reduced after transplantation³.

Although results of Hutter *et al.* and Henrich *et al.* made a revolution in HIV-1 treatment, but there were problems such as low frequency of homozygote *CCR5* $\Delta 32$ in population and founding a suitable HLA match donor with target patient. These problems encouraged Duarte *et al.* in 2015 to transplanted hematopoietic stem cell of umbilical cord blood (CB) to a patient infected by HIV-1 from *CCR5*^{-/-} donor that have AML⁵⁴⁻⁵⁶. HCT from CB not needed to stringent HLA matching like as HSCT with bone marrow. This study showed peripheral mononuclear cells were resistance to HIV-1 infection.

As describe above, artificial disruption methods with “*CCR5* gene modification” were exerted widely and eliminated some problems of allogeneic HSCT⁵⁷. Gene modification is permanent, inheritable and transmitted HIV-1 resistant cells to next generation.

For example for site-specific nuclease, zinc finger protein (ZFP) surveyed extensive. ZFP

is a transcription factor that binding to Fok I restriction enzyme domains with their zinc finger motifs and provide zinc finger nuclease (ZFN) [58, 59]. Theoretically, following DSB in DNA by Fok I, repair DNA mechanism applied error prone non-homologous end joining (NHEJ) [60] or homology directly repair by homologous recombination (HR)⁶¹. Then, small nucleotide deletion or addition observed and result in disruption of reading frame and gene expression. It could engineer with nucleases artificially. Predominant repair with ZFN is error-prone NHEJ.

First in 2005, alternation of *CCR5* with ZFN in *in vitro* condition was showed⁶². Then *CCR5* disruption by ZFN in mouse model was applied and exhibited specify and sufficient *in vivo* functions for induction of resistance to HIV⁶³. In this manner, genetic modification with ZFNs utilized in various living organism and cell lines; Including primary T cell, HSC and humanize mice⁶⁴⁻⁶⁷. Afterwards, *CCR5* was modified by ZFN with adenovirus vector on *ex vivo* condition and performance was efficiently in healthy and HIV infected CD4⁺ T cell in 2013⁶⁸. Knock out of *CCR5* by ZFP modification artificially and infusion of autologous CD4⁺ T cell transplantation showed safety and immune reconstitution with increasing in CD4⁺ T cells, In 2014⁴. In this study DNA virus level decreased in most patients and RNA virus level were undetectable in one patient and modified T cells for *CCR5* were stable⁴.

Another kind of site-specific nuclease is TALEN (Transcription Activator-like Effectors Nuclease). In comparison to ZFN, TALEN has lower cytotoxicity and reduce off-target activity in *CCR5* locus. But both of them could disrupt genes about 45%⁶⁹. In comparison to ZFN more delivery problems observed, due to large TALEN protein size. But had been showed that TALEN expressed by adenoviral vector⁷⁰.

In addition to ZFN and TALEN, CRISPR/Cas9 is another targeted gene disruption in HIV therapy. Clustered regularly interspaced palindromic repeats (CRISPRs) are short direct repeat (21-47 nt) with vary intervening spacer sequence that surround by CRISPR associated gene (Cas9) in bacteria. When CRISPR is transcribed, pre-crRNA converts to crRNA by RNase III and associate with trans-acting RNA (tracrRNA) that diagnose target DNA. Then, this

RNA duplex bind to Cas9 and the crRNA guide complex to target DNA that is complementary to spacer sequence. This ribonucleotide complex cleave target DNA with Cas9 and cause DSB⁷¹. Some studies suggested CRISPR/Cas9 could efficiently ablate the viral genome from latently HIV-1 infected cells^{72,73}. *CCR5* in primary human CD4⁺ T cells and CD34⁺ hematopoietic is targeted progenitor and stem cells and demonstrated ablate viral genes with minimal off-target mutagenesis by CRISPR/Cas9, In 2014⁷⁴.

T cell recovery and suppression of HIV-1 are achieved by gene modification and/or cell therapy. Recent approaches provide effective cure of HIV-1 but there are some challenges in *CCR5* candidate for HIV treatment and need to more investigations.

CONCLUSION

Results of many studies have shown that *CCR5* can be a therapeutic target for treatment of HIV-1. A new horizon of stem cell therapy (such as cord blood stem cell) is shifted to obtain more effective and easier methods that can apply for many people with no HLA-matching problems and donor finding restrictions. We suggest identification of another unknown polymorphism to apply more effective treatment of HIV by find out more genetic factors that promote or restrict HIV replication. We think also it would be interesting to study the therapeutic effect of autologous embryonic stem cell transplantation that modify by human artificial chromosome (HAC) and *ZFN* gene. It can cleavages *CCR5* gene specifically and induces the resistance to HIV-1 infection.

REFERENCES

- Hutter G, Nowak D, Mossner M, Ganepola S, Mussig A, et al. (2009) Long-term control of HIV by *CCR5* Delta32/Delta32 stem-cell transplantation. *N Engl J Med*. 360: 692-8.
- Hutter G, Thiel E (2011) Allogeneic transplantation of *CCR5*-deficient progenitor cells in a patient with HIV infection: an update after 3 years and the search for patient no. 2. *AIDS*. 25: 273-4.
- Henrich TJ, Hu Z, Li JZ, Sciaranghella G, Busch MP, et al. (2013) Long-term reduction in peripheral blood HIV type 1 reservoirs following reduced-intensity conditioning allogeneic stem cell transplantation. *J Infect Dis*. 207: 1694-702.
- Tebas P, Stein D, Tang WW, Frank I, Wang SQ, et al. (2014) Gene editing of *CCR5* in autologous CD4 T cells of persons infected with HIV. *N Engl J Med*. 370: 901-10.
- Lander ES, Linton LM, Birren B, Nusbaum C, Zody MC, et al. (2001) Initial sequencing and analysis of the human genome. *Nature*. 409: 860-921.
- Venter JC, Adams MD, Myers EW, Li PW, Mural RJ, et al. (2001) The sequence of the human genome. *Science*. 291: 1304-51.
- Flower DR (1999) Modelling G-protein-coupled receptors for drug design. *Biochim Biophys Acta*. 1422: 207-34.
- Esmailzadeh A, Farshbaf A, Erfanmanesh M (2015) Autologous Hematopoietic Stem Cells transplantation and genetic modification of *CCR5* m303/m303 mutant patient for HIV/AIDS. *Medical Hypotheses*. 84: 216-18.
- Esmailzadeh A, Farshbaf A (2015) Novel Approaches Based on Autologous Stem Cell Engineering and Gene-Modification; Evidence for the Cure of HIV/AIDS. *Genetic Syndromes & Gene Therapy*. 6:1.
- Samson M, Soularue P, Vassart G, Parmentier M (1996) The genes encoding the human CC-chemokine receptors CC-CKR1 to CC-CKR5 (CMKBR1-CMKBR5) are clustered in the p21.3-p24 region of chromosome 3. *Genomics*. 36: 522-6.
- Samson M, Labbe O, Mollereau C, Vassart G, Parmentier M (1996) Molecular cloning and functional expression of a new human CC-chemokine receptor gene. *Biochemistry*. 35: 3362-7.
- Samson M, Libert F, Doranz BJ, Rucker J, Liesnard C, et al. (1996) Resistance to HIV-1 infection in caucasian individuals bearing mutant alleles of the *CCR-5* chemokine receptor gene. *Nature*. 382: 722-5.
- Cocchi F, DeVico AL, Garzino-Demo A, Arya SK, Gallo RC, et al. (1995) Identification of RANTES, MIP-1 alpha, and MIP-1 beta as the major HIV-suppressive factors produced by CD8+ T cells. *Science*. 270: 1811-5.
- Abbas W, Herbein G (2014) Plasma membrane signaling in HIV-1 infection. *Biochim Biophys Acta*. 1838: 1132-42.
- Luttrell LM (2008) Reviews in molecular biology and biotechnology: transmembrane signaling by G protein-coupled receptors. *Mol Biotechnol*. 39: 239-64.
- Murdoch C, Finn A (2000) Chemokine receptors

- and their role in inflammation and infectious diseases. *Blood*. 95: 3032-43.
17. Vlahakis SR, Villasis-Keever A, Gomez T, Vanegas M, Vlahakis N, et al. (2002) G protein-coupled chemokine receptors induce both survival and apoptotic signaling pathways. *J Immunol*. 169: 5546-54.
 18. Carrington M, O'Brien SJ (2003) The influence of HLA genotype on AIDS. *Annu Rev Med*. 54: 535-51.
 19. An P, Winkler CA (2010) Host genes associated with HIV/AIDS: advances in gene discovery. *Trends Genet*. 26: 119-31.
 20. Hutcheson HB, Lautenberger JA, Nelson GW, Pontius JU, Kessing BD, et al. (2008) Detecting AIDS restriction genes: from candidate genes to genome-wide association discovery. *Vaccine*. 26: 2951-65.
 21. O'Brien SJ, Nelson GW (2004) Human genes that limit AIDS. *Nat Genet*. 36: 565-74.
 22. O'Brien SJ, Hendrickson SL (2013) Host genomic influences on HIV/AIDS. *Genome Biol*. 14: 201.
 23. Liu R, Paxton WA, Choe S, Ceradini D, Martin SR, et al. (1996) Homozygous defect in HIV-1 coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. *Cell*. 86: 367-77.
 24. Zagury D, Lachgar A, Chams V, Fall LS, Bernard J, et al. (1998) C-C chemokines, pivotal in protection against HIV type 1 infection. *Proc Natl Acad Sci U S A*. 95: 3857-61.
 25. Dean M, Carrington M, Winkler C, Huttley GA, Smith MW, et al. (1996) Genetic restriction of HIV-1 infection and progression to AIDS by a deletion allele of the CKR5 structural gene. Hemophilia Growth and Development Study, Multicenter AIDS Cohort Study, Multicenter Hemophilia Cohort Study, San Francisco City Cohort, ALIVE Study. *Science*. 273: 1856-62.
 26. Huang Y, Paxton WA, Wolinsky SM, Neumann AU, Zhang L, et al. (1996) The role of a mutant CCR5 allele in HIV-1 transmission and disease progression. *Nat Med*. 2: 1240-3.
 27. Naif HM (2013) Pathogenesis of HIV Infection. *Infect Dis Rep*. 5: e6.
 28. Deng H, Liu R, Ellmeier W, Choe S, Unutmaz D, et al. (1996) Identification of a major co-receptor for primary isolates of HIV-1. *Nature*. 381: 661-6.
 29. Dragic T, Litwin V, Allaway GP, Martin SR, Huang Y, et al. (1996) HIV-1 entry into CD4+ cells is mediated by the chemokine receptor CC-CKR-5. *Nature*. 381: 667-73.
 30. Arenzana-Seisdedos F, Parmentier M (2006) Genetics of resistance to HIV infection: Role of co-receptors and co-receptor ligands. *Semin Immunol*. 18: 387-403.
 31. Quillent C, Oberlin E, Braun J, Rousset D, Gonzalez-Canali G, et al. (1998) HIV-1-resistance phenotype conferred by combination of two separate inherited mutations of CCR5 gene. *Lancet*. 351: 14-8.
 32. Chinn LW, Tang M, Kessing BD, Lautenberger JA, Troyer JL, et al. (2010) Genetic associations of variants in genes encoding HIV-dependency factors required for HIV-1 infection. *J Infect Dis*. 202: 1836-45.
 33. Smith MW, Dean M, Carrington M, Winkler C, Huttley GA, et al. (1997) Contrasting genetic influence of CCR2 and CCR5 variants on HIV-1 infection and disease progression. Hemophilia Growth and Development Study (HGDS), Multicenter AIDS Cohort Study (MACS), Multicenter Hemophilia Cohort Study (MHCS), San Francisco City Cohort (SFCC), ALIVE Study. *Science*. 277: 959-65.
 34. Gao X, Bashirova A, Iversen AK, Phair J, Goedert JJ, et al. (2005) AIDS restriction HLA allotypes target distinct intervals of HIV-1 pathogenesis. *Nat Med*. 11: 1290-2.
 35. Martin MP, Gao X, Lee JH, Nelson GW, Detels R, et al. (2002) Epistatic interaction between KIR3DS1 and HLA-B delays the progression to AIDS. *Nat Genet*. 31: 429-34.
 36. Martin MP, Qi Y, Gao X, Yamada E, Martin JN, et al. (2007) Innate partnership of HLA-B and KIR3DL1 subtypes against HIV-1. *Nat Genet*. 39: 733-40.
 37. Herbeck JT, Gottlieb GS, Winkler CA, Nelson GW, An P, et al. (2010) Multistage genomewide association study identifies a locus at 1q41 associated with rate of HIV-1 disease progression to clinical AIDS. *J Infect Dis*. 201: 618-26.
 38. Hendrickson SL, Lautenberger JA, Chinn LW, Malasky M, Sezgin E, et al. (2010) Genetic variants in nuclear-encoded mitochondrial genes influence AIDS progression. *PLoS One*. 5: e12862.
 39. Hutter G, Bluthgen C, Neumann M, Reinwald M, Nowak D, et al. (2013) Coregulation of HIV-1 dependency factors in individuals heterozygous to the CCR5-delta32 deletion. *AIDS Res Ther*. 10: 26.
 40. Haqqani AA, Tilton JC (2013) Entry inhibitors and their use in the treatment of HIV-1 infection. *Antiviral Res*. 98: 158-70.
 41. Persaud D, Gay H, Ziemniak C, Chen YH, Piatak M, Jr., et al. (2013) Absence of detectable HIV-1 viremia after treatment cessation in an infant. *N Engl J Med*. 369: 1828-35.

42. Kozyryev I, Zhang J (2015) Bayesian Analysis of Complex Interacting Mutations in HIV Drug Resistance and Cross-Resistance. *Adv Exp Med Biol.* 827: 367-83.
43. Zaccarelli M, Tozzi V, Lorenzini P, Trotta MP, Forbici F, et al. (2005) Multiple drug class-wide resistance associated with poorer survival after treatment failure in a cohort of HIV-infected patients. *AIDS.* 19: 1081-9.
44. Carr A (2003) Toxicity of antiretroviral therapy and implications for drug development. *Nat Rev Drug Discov.* 2: 624-34.
45. Erfan Manesh M, Esmaeilzadeh A, Hajikhan Mirzaei M (2015) IL-24: A Novel Gene Therapy Candidate For Immune System Up-Regulation in Hodgkin's Lymphoma. *Journal of Medical Hypotheses and Ideas.* 9: 61-6.
46. Hajikhan Mirzaei M, Esmaeilzadeh A (2014) Overexpression of MDA-7/IL-24 as an anticancer cytokine in gene therapy of thyroid carcinoma. *Journal of Medical Hypotheses and Ideas.* 8: 7-13.
47. Piri Z, Esmaeilzadeh A, Hajikhanmirzaei M (2012) Interleukin-25 as a candidate gene in immunogene therapy of pancreatic cancer. *Journal of Medical Hypotheses and Ideas.* 6: 75-9.
48. Mazaheri T, Esmaeilzadeh A, Mirzaei MH (2012) Introducing the immunomodulatory effects of mesenchymal stem cells in an experimental model of Behçet's disease. *Journal of Medical Hypotheses and Ideas.* 6: 23-7.
49. Mirzamohammadi F, Esmaeilzadeh A (2007) A Novel Method in Gene Therapy of HIV. *iranian journal of public health.* 36:
50. Sora F, Antinori A, Piccirillo N, De Luca A, Chiusolo P, et al. (2002) Highly active antiretroviral therapy and allogeneic CD34(+) peripheral blood progenitor cells transplantation in an HIV/HCV coinfecting patient with acute myeloid leukemia. *Exp Hematol.* 30: 279-84.
51. Jubault V, Burgard M, Le Corfec E, Costagliola D, Rouzioux C, et al. (1998) High rebound of plasma and cellular HIV load after discontinuation of triple combination therapy. *AIDS.* 12: 2358-9.
52. Allers K, Hutter G, Hofmann J, Loddenkemper C, Rieger K, et al. (2011) Evidence for the cure of HIV infection by CCR5Delta32/Delta32 stem cell transplantation. *Blood.* 117: 2791-9.
53. Hutter G, Ganepola S (2011) Eradication of HIV by transplantation of CCR5-deficient hematopoietic stem cells. *ScientificWorldJournal.* 11: 1068-76.
54. Duarte RF, Salgado M, Sanchez-Ortega I, Arnan M, Canals C, et al. (2015) CCR5 Delta32 homozygous cord blood allogeneic transplantation in a patient with HIV: a case report. *Lancet HIV.* 2: e236-42.
55. Petz LD, Redei I, Bryson Y, Regan D, Kurtzberg J, et al. (2013) Hematopoietic cell transplantation with cord blood for cure of HIV infections. *Biol Blood Marrow Transplant.* 19: 393-7.
56. Petz L (2013) Cord blood transplantation for cure of HIV infections. *Stem Cells Transl Med.* 2: 635-7.
57. Manjunath N, Yi G, Dang Y, Shankar P (2013) Newer gene editing technologies toward HIV gene therapy. *Viruses.* 5: 2748-66.
58. Pavletich NP, Pabo CO (1991) Zinc finger-DNA recognition: crystal structure of a Zif268-DNA complex at 2.1 Å. *Science.* 252: 809-17.
59. Bitinaite J, Wah DA, Aggarwal AK, Schildkraut I (1998) FokI dimerization is required for DNA cleavage. *Proc Natl Acad Sci U S A.* 95: 10570-5.
60. Lieber MR (2010) The mechanism of double-strand DNA break repair by the nonhomologous DNA end-joining pathway. *Annu Rev Biochem.* 79: 181-211.
61. Moynahan ME, Jasin M (2010) Mitotic homologous recombination maintains genomic stability and suppresses tumorigenesis. *Nat Rev Mol Cell Biol.* 11: 196-207.
62. Mani M, Kandavelou K, Dy FJ, Durai S, Chandrasegaran S (2005) Design, engineering, and characterization of zinc finger nucleases. *Biochem Biophys Res Commun.* 335: 447-57.
63. Perez EE, Wang J, Miller JC, Jouvenot Y, Kim KA, et al. (2008) Establishment of HIV-1 resistance in CD4+ T cells by genome editing using zinc-finger nucleases. *Nat Biotechnol.* 26: 808-16.
64. Wilen CB, Wang J, Tilton JC, Miller JC, Kim KA, et al. (2011) Engineering HIV-resistant human CD4+ T cells with CXCR4-specific zinc-finger nucleases. *PLoS Pathog.* 7: e1002020.
65. Yuan J, Wang J, Crain K, Fearn C, Kim KA, et al. (2012) Zinc-finger nuclease editing of human cxcr4 promotes HIV-1 CD4(+) T cell resistance and enrichment. *Mol Ther.* 20: 849-59.
66. Doyon Y, Vo TD, Mendel MC, Greenberg SG, Wang J, et al. (2011) Enhancing zinc-finger-nuclease activity with improved obligate heterodimeric architectures. *Nat Methods.* 8: 74-9.
67. Voit RA, McMahan MA, Sawyer SL, Porteus MH (2013) Generation of an HIV resistant T-cell line by targeted "stacking" of restriction factors. *Mol Ther.* 21: 786-95.
68. Maier DA, Brennan AL, Jiang S, Binder-Scholl GK, Lee G, et al. (2013) Efficient clinical scale

- gene modification via zinc finger nuclease-targeted disruption of the HIV co-receptor CCR5. *Hum Gene Ther*. 24: 245-58.
69. Mussolino C, Morbitzer R, Lutge F, Dannemann N, Lahaye T, et al. (2011) A novel TALE nuclease scaffold enables high genome editing activity in combination with low toxicity. *Nucleic Acids Res*. 39: 9283-93.
70. Holkers M, Maggio I, Liu J, Janssen JM, Miselli F, et al. (2013) Differential integrity of TALE nuclease genes following adenoviral and lentiviral vector gene transfer into human cells. *Nucleic Acids Res*. 41: e63.
71. Wiedenheft B, Lander GC, Zhou K, Jore MM, Brouns SJ, et al. (2011) Structures of the RNA-guided surveillance complex from a bacterial immune system. *Nature*. 477: 486-9.
72. Ebina H, Misawa N, Kanemura Y, Koyanagi Y (2013) Harnessing the CRISPR/Cas9 system to disrupt latent HIV-1 provirus. *Scientific Reports*. 3: 2510.
73. Hu W, Kaminski R, Yang F, Zhang Y, Cosentino L, et al. (2014) RNA-directed gene editing specifically eradicates latent and prevents new HIV-1 infection. *Proc Natl Acad Sci U S A*. 111: 11461-6.
74. Mandal PK, Ferreira LM, Collins R, Meissner TB, Boutwell CL, et al. (2014) Efficient ablation of genes in human hematopoietic stem and effector cells using CRISPR/Cas9. *Cell Stem Cell*. 15: 643-52.
75. Menendez-Arias L (2013) Molecular basis of human immunodeficiency virus type 1 drug resistance: overview and recent developments. *Antiviral Res*. 98: 93-120.
76. Archin NM, Liberty AL, Kashuba AD, Choudhary SK, Kuruc JD, et al. (2012) Administration of vorinostat disrupts HIV-1 latency in patients on antiretroviral therapy. *Nature*. 487: 482-5.