

Morphological and Molecular Genetic Assessment Of Some Thymus Species

Mesfer M. Alqahtani¹, Mohamed A. Abdein² and Omnia F. Abou El-Leel³

¹Department of Biological Sciences, Faculty of Science and Humanities, Shaqra University, P.O. Box 1040, Ad-Dawadimi, 11911, Saudi Arabia.

²Biology Department, Faculty of Arts and Science, Northern Border University, Rafha, 91911, Saudi Arabia.

³Vegetable and Medicinal and Aromatic Plants Research Departments, Dokki, Giza and Biotechnology Lab. Horticulture Research Institute, Agricultural Research Centre, 12619, Egypt.

<http://dx.doi.org/10.13005/bbra/2815>

(Received: 06 January 2020; accepted: 13 February 2020)

This study aimed to determine the morphological and genetically assessment in five Thymus species: *Thymus vulgaris*, *Origanum vulgare*, *Thymus argenteus*, *Thymus citriodorus* and *Origanum syricum*. Morphological assessment for the five Thymus species were obtained based on some vegetative parameters including: Plant height, Number of branches, Leaves fresh & dry weights and Volatile Oil%. Molecular genetic variability was assessed based on (SCoT-PCR) and (ISSR-PCR) analysis. Growth parameters were illustrated among five Thymus species in all growth parameters were had significant differences. The SCoT-PCR analysis using 5 out of 10 primers tested, the results illustrated that SCoT primers produced 24 Polymorphic bands out of 39 amplified bands with polymorphic average 60.52%, also five ISSR primers out of 14 primers tested, which analysis were generated 14 polymorphic bands out of 23 amplified bands with polymorphic average 60.86%. As well as assessment of SCoT and ISSR molecular marker techniques succeeded in generating reproducible and reliable amplified bands and from obvious results, SCoT-PCR analysis was better than ISSR-PCR analysis in molecular genetics. On the other hand, results obtained from an UPGMA dendrograms resulted in two genetically distinct clusters were determined between Thymus species. This results were conducted that SCoT and ISSR analysis could be useful as tools for identifying Thymes species in breeding programs.

Keywords: Growth Parameters; ISSR; Molecular Genetics; SCoT; Thymus Species.

Thymus genus which belongs to the family Lamiaceae, includes several hundreds of species distributed over world¹, where Mediterranean basin is considered the main center of this herbal plant².

Traditionally, most of plants discriminated on morphological-basis; however, these methods still difficult to apply for an accurate discrimination

and authentication use³. Thymus genus is usually used for flavoring agents, herbal tea, and medicine and the aerial parts and volatile constituents of thyme are used as a medicinal material^{2,4}. reported that, many species in the genus Thymus were polyploidy and disploidy/aneuploidy and further complicate the determination of species

*Corresponding author E-mail: abdeingene@yahoo.com



boundaries and there were have hybrid vigor in the species, probably due to the absence of incompatibility and the presence of a dimorphic breeding program, in genetic populations comprise female and hermaphroditic individuals.⁵ Reported that, Knowledge of genetic diversity within species is necessary for any improvement of cultivars, and biodiversity maintenance and restoration. DNA-based molecular markers, which are not affected by ecological stress have become increasingly important for aromatical biodiversity and genome of aromatical plants⁶. These molecular genetics can also be taxonomically to biodiversity for cytological studies to taxonomically to species and subspecies^{7,8,9,10}.

The start codon targeted (SCoT) polymorphism is a novel, simple and reliable SCoT based on the translation start codon11. Primers for start codon targeted were designed based on the con-served region surrounding the translation initiation codon, ATG. Using a single 18-mer primer as a forward & reversed primers in polymerase chain reaction,¹¹ designed thirty-six primers that were used successfully for plant identification and biodiversity analysis in many medicinal plants. Being characterized by lower recombination levels between its markers and the gene/trait.¹² Conducted that, This ISSR-PCR technique is rapid, simple, inexpensive and more reproducible than RAPD amplification of DNA.. ISSR used to study the genetic diversity of plants for examples; Nepeta¹³, Thyme¹⁴, Salvia¹⁵, Mentha aquatica L.¹⁶, Satureja¹⁷, Salvia¹⁸, Thymus¹⁹, *Phlomis kurdica* and *Phlomis oppositiflora*²⁰ and Ocimum²¹.

Increasingly, the hybridization approach has taken advantage of developments in molecular genetics in order to karyotype of interest in a way that considerably accelerates natural selection and this genotypes approach consist of choosing desired genotypes on the basis of molecular biology, or having prior knowledge of the genes that determine the morphological traits in a plant²².

This work was aimed to study molecular genetics and morphological assessment among different species of this plant using morphological and SCoT and ISSR markers, with a view toward conservation of this endangered species.

MATERIALS AND METHODS

Genetic Resources

The seedlings of the five Thymus species were obtained from two countries, Egypt and Kingdom of Saudi Arabia, as a commonly known species showed in Table 1. Thymus plants were collected during two seasons of 2017/2018 and 2018/2019, at Vegetable and Medicinal and Aromatic Plants Research Departments, Dokki, Giza and Biotechnology Research Lab., H.R.I., A.R.C., Egypt. Two months old seedlings of five Thymus Sp. were obtained from the greenhouse of Vegetable and Medicinal and Aromatic Plants Research Dep., Egypt.

Vegetative Parameters

Plant height (cm), Number of branches, Leaves fresh & dry weights and Volatile Oil %.

Molecular Genetic Assessment

DNA Extraction

The DNA extraction of the five species of Thymus was performed as described by²³. DNA extractions were checked by means of absorbance ratios a260:a280 through a UV-spectrophotometer where Deoxyribo Nucleic Acid is pure with a same ratio from 1.8 - 2.0. Moreover, using electrophoresis in 1% agarose gel with ethidium bromide.

SCoT and ISSR Analysis

Obtaining clear reproducible amplification products require a number of factors were included polymerase chain reaction temperature cycle profile and concentration which were optimized according to24 and 25 respectively, in the PCR reaction using 5 SCoT primers and 5 ISSR primers in molecular genetic analysis for the five Thymus species. ISSR primers procured from Bio Basic Company Canada. On the other hand, SCoT primers sequence were designed and derived from the investigations studies by²⁶ and^{27,11} and procured from Biobasic Company.

inter-simple sequence repeat and start codon targeted assays were performed as described by^{24,25 and 28}.

Gel Electrophoresis

PCR products were running throw mini agarose unit at 100 V for one 30min. and 1.5 % agarose gel using 100bp Ladder DNA marker to study the molecular variation between the Thymus species.

Statistically Analysis

RCBD was adopted for the present study data were Statistical analysis by the standard methods according to²⁹. The new Least Significant Difference (LSD) test was used for comparison between means. The bands of DNA generated by each primer were counted and their molecular sizes were compared with those of the ISSR and SCoT assays. The bands scored from DNA profiles generated by each primer were pooled together. Then the presence or absence for each band of DNA was treated as a binary character in a data matrix (coded one & zero, respectively) to calculate genetic similarity and to construct dendrogram tree among the studied five *Thymus* Species. Calculation was achieved using Dice similarity coefficients³⁰ as implemented in the computer program SPSS-10.

RESULTS AND DISCUSSION

Vegetative Composition Diameter

The vegetative parameters results were including plant height (cm), number of branches/plant and leaves fresh & dry weights (g), of (*Thymus* species) seedlings in both two seasons are shown in Table (2).

Plant Height

Table (2) represented that plant height, *Origanum vulgare* was the highest plant in first season in the two cuts were as follow (31.39 and 35.37cm) and increased in the second season was cuts as follows (35.90 and 32.31cm) and this followed by *Thymus citriodorus*, *Thymus vulgaris* and *Origanum syricum*. While, the lowest in the *Thymus* Sp. in plant height was *Thymus 3* which results in first season in the two cuts were as follows (6.82 and 8.91cm) and in the second season data

in the tow cuts were as follows (9.62 and 9.19cm), respectively.

Branch Number

Table (2) results revealed that, the number of branches it was clear from that the greatest branches number were revealed by *Thym.1* in the first season: in two cuts were as follow (16 and 39) and in the second season: in the two cuts were as follow (39 and 42.67) and this results were followed by *Thymus argenteus*, *Thymus citriodorus* and *Origanum syricum*. While, the lowest number of branches were recorded in *Origanum vulgare* which were in the first season: in the two cuts as follow (3.67 and 4) and in the second season: in the two cuts ere as follow (4.33 and 5.33), respectively.

Fresh and Dry Weight of Leaves /Plant

the results of leave fresh & dry weights in the five sp. of *Thymus* in the two seasons data were revealed in Table (2), *Thymus argenteus* results were recorded as the highest data in all *Thymus* sp. under study and results were as follow, in the first season : fresh weight of leaves/plant in the two cuts were as follow (422.4 and 519 gm) and in dry weight of leaves/plant were as follow in the two cuts (49.25 and 53.98gm). While, in the second season: fresh weight of leaves/plant in the two cuts were as follow (522.14 and 659.99 gm) and in dry weight of leaves/plant were as follow in the two cuts (52.34 and 65.20gm) and this results were followed by *Origanum vulgare*, *Thymus vulgaris* and *Thymus citriodorus*, respectively. On the other hand, *Origanum syricum* was the lowest in both fresh and dry weight of leaves/plant in the two seasons and the results were as follow, the first season: fresh weight of leaves/plant in the two cuts were as follow (23.74 and 27.51 gm) and in dry weight of leaves/plant were as follow in the two cuts (8.41 and 9.57gm). While, in the second

Table 1. The *Thymus* species numbers and the names of the five studied species

Cultivar Number	<i>Thymus</i> species	Common name	Origin
1	<i>Thymus vulgaris</i>	Balady	Egypt
2	<i>Origanum vulgare</i>	Syrian	Saudi Arabia
3	<i>Thymus argenteus</i>	Oregano	Egypt
4	<i>Thymus citriodorus</i>	Jordanian	Saudi Arabia
5	<i>Origanum syricum</i>	Gabaly	Saudi Arabia

Table 2. Vegetative parameters, plant height, branches number, fresh weight, dry weight and volatile oil % of five *Thymus* species through two cuts and two seasons.

	First cut					Second Cut				
	Plant height (cm)	Branches number	Fresh weight (g/plant)	Dry weight (g/plant)	Volatile oil %	Plant height (cm)	Branches number	Fresh weight (g/plant)	Dry weight (g/plant)	Volatile oil %
First season										
<i>Thymus vulgaris</i>	21.53	16.00	33.74	11.47	0.10	23.76	39.00	49.59	17.20	0.12
<i>Origanum vulgare</i>	31.39	3.67	36.42	12.39	0.27	35.37	4.00	51.04	14.28	0.29
<i>Thymus argenteus</i>	6.82	6.67	422.46	49.25	0.07	8.91	8.67	519.81	53.98	0.08
<i>Thymus citriodorus</i>	29.24	5.33	29.10	11.42	0.22	32.22	9.00	41.26	14.36	0.26
<i>Origanum syriacum</i>	12.30	4.33	23.74	8.41	0.05	13.72	5.67	27.51	9.57	0.06
New L.S.D. (0.05) =	1.42	1.88	86.27	9.87	0.02	1.11	3.16	93.45	14.08	0.01
Second season										
<i>Thymus vulgaris</i>	26.76	39.00	36.51	12.85	0.12	24.47	42.67	52.34	18.79	0.14
<i>Origanum vulgare</i>	35.90	4.33	39.99	13.82	0.31	31.31	5.33	58.29	15.94	0.33
<i>Thymus argenteus</i>	9.62	8.33	522.14	52.34	0.11	9.19	8.67	659.99	56.20	0.13
<i>Thymus citriodorus</i>	32.76	7.33	36.10	13.08	0.27	30.48	9.33	43.92	12.36	0.29
<i>Origanum syriacum</i>	14.25	5.00	29.36	11.13	0.07	12.73	6.67	30.40	11.07	0.12
New L.S.D. (0.05) =	0.70	2.63	81.85	9.90	0.02	1.42	1.17	114.93	12.81	0.03

season: fresh weight of leaves/plant in the two cuts were as follow (29.36 and 30.40 gm) and in dry weight of leaves/plant were as follow in the two cuts (11.13 and 11.07gm).

Volatile Oil %

The results were observed in Table (2) illustrated the largest amount of volatile oil % was in *Origanum vulgare* in both two seasons as follow, the first season: results in the two cut were as follow (0.27 and 0.29%) and in the second season: results in the two cuts were as follow: (0.31 and 0.33%) respectively, and this results were followed by *Thymus citriodorus*, *Thymus vulgaris* and *Thymus argenteus*. While, the lowest amount of volatile oil

% was observed in *Origanum syriacum* in both two seasons and the results as follow, the first season: the two cut were as follow (0.05 and 0.06%) and in the second season: results in the two cuts were as follow: (0.07 and 0.12%), respectively.

Molecular Genetics Assessment

This results of the genetic variability in five species of *Thymus* using SCoT-PCR and ISSR-PCR analysis. Where five SCoT primers out of ten tested primers were succeeded on the five different *Thymus* Species, and five ISSR primers out of fourteen tested primers generated reproducible amplified bands.

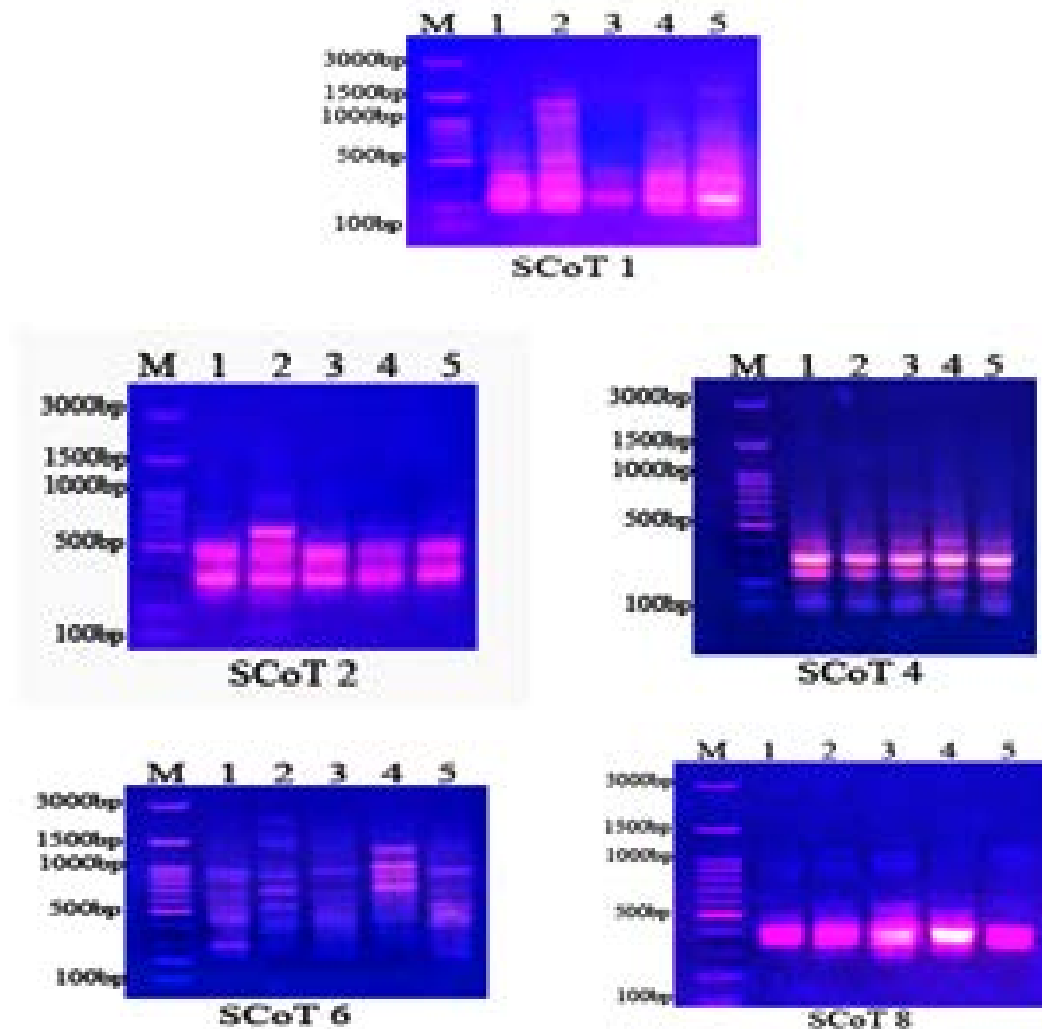


Fig. 1. SCoT-PCR Profile for five species of *Thymus* amplified with five primers for each analysis

SCoT and ISSR Analysis Assessment

Molecular genetic data produced by SCoT and ISSR analysis were shown in Figs (1 and 2) and Tables (3 and 4). These data showed that, in SCoT results, primer (SCoT-6) was resulted in the highest number of amplified bands and primer (SCoT-4) was represented the lowest number of amplified bands compared with other SCoT primers. On the other hand, in ISSR data, primer 44B resulted in the highest number of amplified bands and primer (HB-14) showed the lowest number of amplified bands in all ISSR primers.

On the other hand, SCoT primers except SCoT 4 and SCoT-8 generated 10 unique bands

out of 39 amplified bands and ISSR primers except (44B, HB-10 and HB-14) generated 4 unique bands out of 23 amplified bands, May be these unique bands were useful as unique markers as explained by 31 in cymbopogon; 32 in canolla; 33 in tomato and 34 in pumpkin.

Also, Table 5 showed that five species of Thymus, (*Thymus vulgaris*, *Origanum vulgare*, *Thymus argenteus*, *Thymus citriodorus* and *Origanum syricum*) characterized by five SCoT primers and five ISSR primers data, 23 polymorphic bands from 38 amplified bands were produced by SCoT primers with polymorphic average 60.52%. While, 14 polymorphic bands from 23 amplified

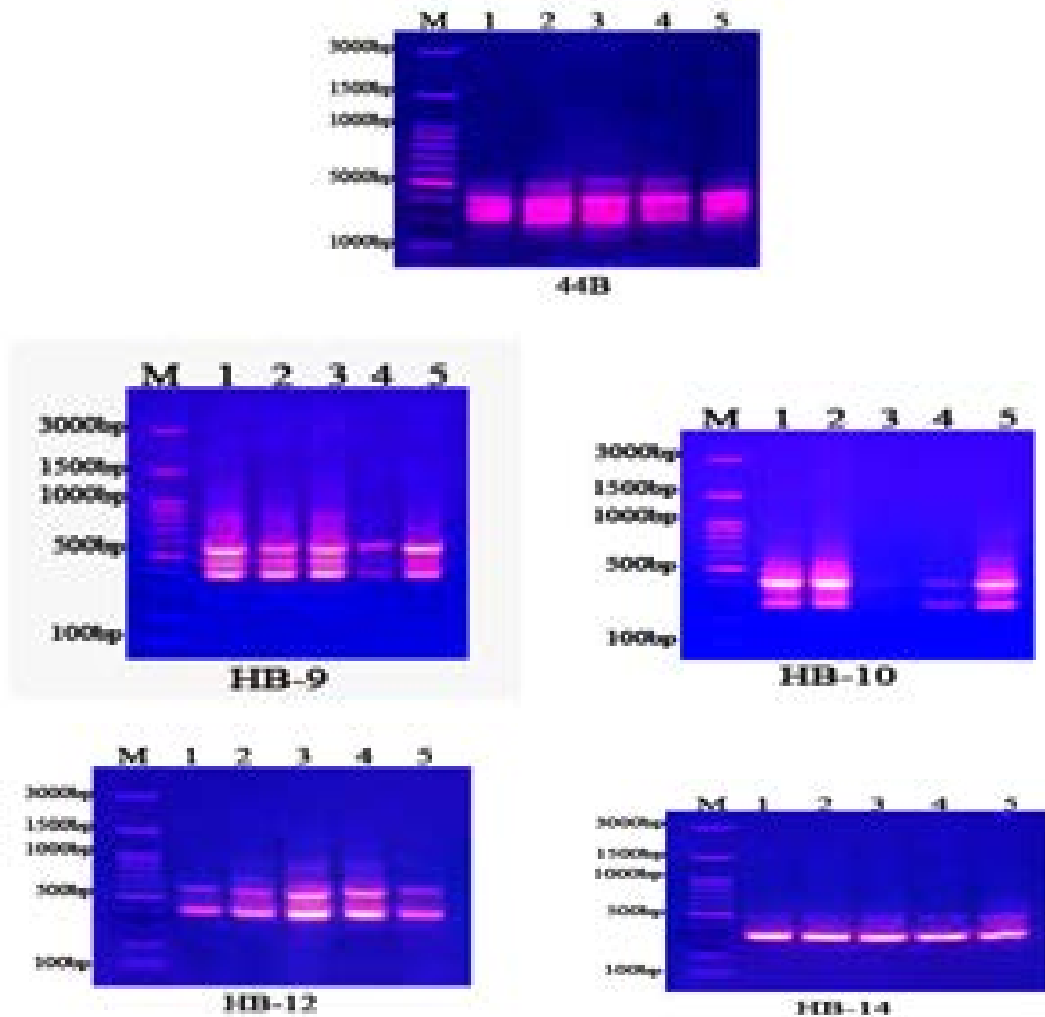


Fig. 2. ISSR-PCR Profile for five species of Thymus amplified with five primers for each analysis

Table 3. Molecular genetic data produced from amplified banding patterns of SCoT technique

Primer Name	Sequence (5→3')	Molecular size range	Total Amplified Band	Monomorphic Band	Polymorphic band	Unique Band	Polymorphic %
SCoT 1	CAA CAATGGCTACCCACC	180:1470	8	3	5	2	62.50%
SCoT 2	CAACAATGGCTACCCACC	135:920	7	3	4	1	50%
SCoT 4	CAACAATGGCTACCCACC	135:48001	5	4	1	-	20%
SCoT 6	CAACAATGGCTACCCACC	193:1063	12	2	10	7	83.33%
SCoT 8	CAACAATGGCTACCCACC	195:587	7	1	4	-	14.28%
Total			38	15	24	10	60.52%

Table 4. Molecular genetic data produced from amplified banding patterns of ISSR technique

Primer Name	Sequence (5→3')	Molecular size range	Total Amplified Band	Monomorphic Band	Polymorphic band	Unique Band	Polymorphic %
44B	(CT) ₈ GC	150:560	6	3	3	-	50%
HB-09	(GT) ₆ GC	380:760	5	1	4	3	80%
HB-10	(GA) ₆ CC	300:560	4	1	3	-	75%
HB-12	(CAC) ₃ GC	300:840	5	1	4	1	80%
HB-14	(CT) ₃ GC	380:480	3	3	-	-	-
Total			23	9	14	4	60.86%

bands with polymorphic average 60.86% were generated by ISSR primers. On the other hand, in the combined results there were 37 polymorphic bands from total 61 amplified bands with total polymorphic average 60.65%. These obtained data indicates that SCoT-PCR and ISSR-PCR techniques were succeeded in differentiate between five *Thymus* species studied.

Molecular Distance of Combination of SCoT and ISSR Analysis

On the other hand, Table (6) illustrated that, results of molecular distance (MD) matrix between all five species of *Thymus* studied based on SCoT and ISSRs combined results.

Molecular distances based on SCoT analysis data were ranged from 0.633 (between *Thymus vulgaris* and *Thymus citriodorus* species) to 0.843 (between *Thymus vulgaris* and *Origanum vulgare* species) was lower than molecular distance based on ISSR ranged from 0.603 (between *Thymus vulgaris* and *Thymus citriodorus* species) to 0.942

(between *Thymus vulgaris* and *Origanum vulgare* species). While in molecular distance combination data were ranged from 0.164 to 0.404 among the same genotypes obtained by SCoT analysis.

Previously data represented the important of SCoT-PCR technique in molecular genetic assessment in *Thymus* species in comparison with ISSR-PCR technique. These results were in agreement with Nepeta¹³, Thyme¹⁴, *Mentha aquatica* L.¹⁶, *Satureja*¹⁷, *Salvia*¹⁸ and *Thymus*¹⁹.

Dendrogram Analysis of Combination Between SCoT and ISSR Analysis

Fig. 3. illustrated Dendrogram tree of SCoT and ISSR analysis combination data were divided the five *Thymus* Species into two main clusters: The first cluster contained two *Thymus* sp. (*Thymus argenteus* and *Thymus citriodorus*) and the second cluster was divided into two sub-clusters: the first sub-cluster included *Origanum syriacum* only. On the other hand, the second sub-cluster included the other species (*Thymus vulgaris* and *Origanum vulgare*).

Table 5. Polymorphic, Monomorphic, Unique bands and Polymorphic percentage generated by the (ISSR and SCoT) analysis

Primer Name	Total Amplified Band	Monomorphic Band	Polymorphic band	Unique Band	Polymorphic %
SCoT	38	15	23	4	60.52%
ISSR	23	9	14	4	60.86%
Total	61	24	37	8	60.65%

Table 6. Molecular distances (MD) between five *Thymus* Species based on Dice dissimilarity index for SCoT &ISSR and combined data

MD	<i>Thymus vulgaris</i>	<i>Origanum vulgare</i>	<i>Thymus argenteus</i>	<i>Thymus citriodorus</i>	<i>Origanum syriacum</i>
<i>Origanum vulgare</i>	ISSR	0.942			
	SCoT	0.843			
	Comb	0.872			
<i>Thymus argenteus</i>	ISSR	0.743	0.813		
	SCoT	0.752	0.732		
	Comb	0.733	0.753		
<i>Thymus citriodorus</i>	ISSR	0.603	0.684	0.813	
	SCoT	0.633	0.702	0.732	
	Comb	0.602	0.680	0.763	
<i>Origanum syriacum</i>	ISSR	0.902	0.853	0.684	0.661
	SCoT	0.753	0.733	0.761	0.771
	Comb	0.822	0.792	0.721	0.721

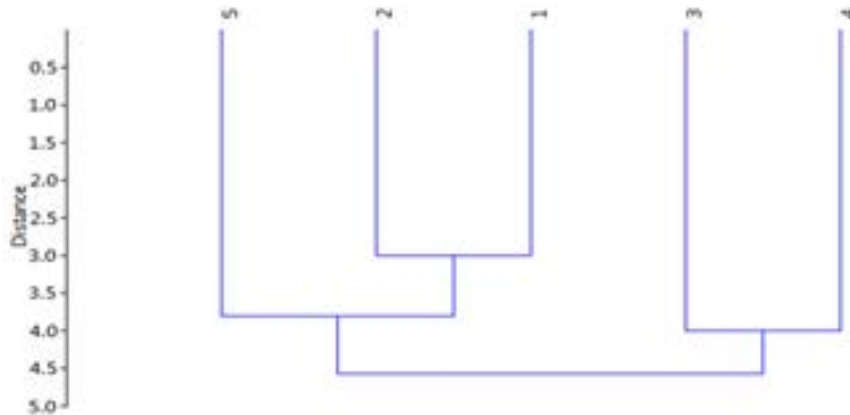


Fig. 3. Dendrogram tree for combination data of SCoT and ISSR analysis for the five *Thymus* species

This results were conducted that SCoT and ISSR analysis could be defined as tools for identifying *Thymus* species in breeding programs and combination data from SCoT and ISSR analysis were suitable for the genetic relationships evaluation between the five *Thymus* species and this results were in agreement with genetic analysis has been conducted by^{35,36,37}, *Salvia*¹⁸ and *Thymus*¹⁹ *Phlomis kurdica* and *Phlomis oppositiflora*²⁰ and *Ocimum*^{21,38}. Revealed that, by using of ISSR-PCR technique of some accessions of *Thymus daenensis*, was obtained two geographically diverse groups were generated by dendrogram.

ACKNOWLEDGMENT

The authors appreciate the constant help and work support provided by Shaqra University & Northern Border University, Saudi Arabia and Agricultural Research Centre, Egypt.

REFERENCES

1. Akcin, A.T. Numerical taxonomic studies on some species of the genus *Thymus* L. (Labiatae) in Turkey. *Asian Journal of Plant Science*. 2006; **5**(5): 782-788.
2. Stahl-Biskup, E. and F. Saez. *Thyme, the genus Thymus*. Taylor and Francis, London and New York, 2002; p. 330.
3. Arif, A., M.A. Bakir, H.A. Khan, A.H. Al-Farhan, A.A. Al-Homaidan, A.H. Bahkali, M. Al-Sadoon and M. Shobrak. Application of RAPD for molecular characterization of plant species of medicinal value from an arid environment. *Genetic and Molecular Research*. 2010; **9**(4): 2191-2198.
4. Sostaric, I., Liberz, Gradisa M, Martin PD, Stevanovic Z.D., Satovicz. Genetic diversity and relationships among species of the genus *Thymus* L. (section Serphyllum). *Flora*. 2012; **207**: 654-661.
5. Karp, A., K. Edwards, M. Bruford, B. Vosman, M. Morgante, O. Seberg, A. Kremer, P. Boursot, P. Arctander, D. Tautz and G. Hewitt. Newer molecular technologies for bio-diversity evaluation: opportunities and challenges. *Nature Biotechnol.*, 1997; **15**: 625-628.
6. Nybom, H. and K. Weising. DNA profiling of plants. *Medicinal Plant Biotechnology*. 2007; **9**: 73-95.
7. Lynch, M. and B.G. Milligan. Analysis of population genetic-structure with RAPD markers. *Mol. Ecol.* 1994; **3**: 91-99.
8. Mulcahy, D.L., M. Cresti, H.F. Linskens, C. Intriери, O. Silverstoni, R. Vignani and M. Pancaldi. DNA fingerprinting of Italian grape varieties: a test of reliability in RAPDs. *Advanced Horticultural Science.*, 1995; **9**: 185-187.
9. Baigi, M.N.R., S. Grewal and S. Dhillon. Molecular characterization and genetic diversity analysis of citrus cultivars by RAPD markers. *Turk J. Agric. For.*, 2009; **33**: 375-384.
10. Alamdary, S.B.L., A. Safarnejad and M. Rezaee. Evaluation of genetic variation between *Thymus* accessions using molecular markers. *J. Basic. Appl. Sci. Res.*, 2011; **1**(12): 2552-2556.

11. Collard, B.C.Y. and D.J. Mackill. Start Codon Targeted (SCoT) polymorphism: A simple novel DNA marker technique for generating gene-target markers in plants. *Plant Molecular Biology*. 2009; **27**: 86-93.
12. Pharmawati, M., G. Yan and I.J. McFarlane. Application of RAPD and ISSR markers to analyses molecular relationships in *Grevillea* (Proteaceae). *Aust. Syst. Bot.* 2004; **17**(1): 49-60.
13. Smolik, M.D. Jadczyk and A. G³ówczyk. Assessment of morphological and genetic variability in chosen *Nepeta* accessions. *Herba Polonica.*, 2008; **54**(4): 68-78.
14. Smolik, M.D. Jadczyk and S. Korzeniewska. Assessment of morphological and genetic variability in some *Thymus* accessions using molecular markers. *Not. Bot. Hort. Agrobot. Cluj*. 2009; **37**(2): 234-240.
15. Javan, Z.S., F. Rahmani and R. Heidar. Assessment of genetic variation of genus *Salvia* by RAPD and ISSR markers. *AJCS.*, 2012; **6**(6):1068-1073.
16. Schanzer, I.A.; M.V. Semenova, O.V. Shelepova and TV. Voronkova. Genetic diversity and natural hybridization in populations of clonal plants of *Mentha aquatica* (Lamiaceae). *Wulfenia*. 2012; **19**: 131-139.
17. Kameli, M., S.M. Hejazi and M. Ebadi. Assessment of genetic diversity on populations of three *Satureja* species in Iran using ISSR markers. *Annals of Biological Research*. 2013; **4**(3): 64-72.
18. Yousefiazarkhanian, M.A. Asghari, J. Ahmadi, B. Asghari and A.A. Jafari. Genetic Diversity Assessment of some *Salvia* sp. ecotypes based on ISSR markers. *Biological Forum An International Journal*. 2015; **7**(1): 286-288.
19. Yousefi, V., A. Najaphy, A. Zabarjadi and H. Safari. Molecular characterization of *Thymus* species using ISSR Markers. *The Journal of Animal & Plant Sciences*. 2015; **25**(4):1087-1094.
20. Evren, OH., E.Y. Uzbasioglu and M.Y. Dadand. Determination of intra-specific genetic variation of *Phlomis kurdica* and *Phlomis oppositiflora* and investigation for the hybridity of P. x melitenense (Lamiaceae) by means of molecular markers. *Institute of Botany, Slovak Academy of Scienc. Uesn*. 2015; **70**(9): 1159-1171.
21. Patel, H.K., R.S. Fougat, S. Kumar, J.G. Mistry and M. Kumar. Detection of genetic variation in *Ocimum* species using RAPD and ISSR markers. *3 Biotech.*, 2016; **5**(5): 697-707.
22. Pradeep Reddy, M., N. Sarla, and E.A. Siddiq. Inter simple sequence repeat (ISSR) polymorphism and its application in plant breeding. *Euphytica*. 2002; **128**: 9-17.
23. Dellaporta, S.L.; J. Wood and J.B. Hicks. A plant DNA mini preparation. Version III. *Plant Mol. Biol., Rep*, 1983; **1**: 19-21.
24. Fathi, M.A.; SH.M. Hussein and S.Y. Mohamed (2013). Horticultural and molecular genetic evaluation of some peach selected strains cultivated under Kalubiah governorate conditions. *9(1st):12-23*.
25. Xiong, F.Q.; R.C. Zhong; Z.Q. Han and J. Jiang. Start codon targeted polymorphism for evaluation of functional genetic variation and relationships in cultivated peanut (*Arachis hypogaea*) genotypes. *Mol. Biol. Rep*, 2011; **38**(5): 3487- 3494.
26. Joshi, C.P.; H. Zhou; X. Huang and V.L. Chiang. Context sequences of translation initiation codon in plants. *Plant Mol. Biol*. 1997; **35**: 993-1001.
27. Sawant, S.V.; P.K. Singh; S.K. Gupta; R. Madnala and R. Tuli. Conserved nucleotide sequences in highly expressed genes in plants. *J. Genet.*, 1999; **78**:123 131.
28. Ozyurt, I.K ; Y.Akca and S. Ercisli. Molecular characterization of *Prunus mahaleb* rootstock candidates by ISSR markers. *Genetika*. 2013; **45**(3):717-726.
29. Snedecor, G.W. and W.G. Cochran (1980). "Statistical Methods". Oxford and J.B.H. publishing comm., 6th "edition".
30. Dice, L.R. Measures of the amount of ecologic association between species. *Ecology*. 1945; **26**: 297-302.
31. Adhikari, S.; S. Saha; T.K. Bandyopadhyay and P. Ghosh. Efficiency of ISSR marker for characterization of cymbopogon germplasm and their suitability in molecular barcoding. *Plant systematic and Evaluation*. 2015; **301**: 439-450.
32. Abd El-Aziz, M.H. and R.M. Habiba. Molecular assessment of genetic diversity in some canola homozygous lines. *Egyptian Journal of Genetics and Cytology*. 2016; **45**: 129- 145.
33. Abdein, M.A., D. Abd El-Moneim, S.S. Taha, W.S.M. Al-Juhani and S.E. Mohamed. Molecular characterization and genetic relationships among some tomato genotypes as revealed by ISSR and SCoT markers. *Egyptian Journal of Genetics and Cytology.*, 2018; **47**(1) Jun., 2018.
34. Abdein, M.A. Genetic Diversity between Pumpkin Accessions Growing in the Northern Border Region in Saudi Arabia Based on Biochemical and Molecular Parameters. *Egyptian Journal Botany*. 2018; **58**(3) , pp. 463-476.
35. Fracaro, F. and S. Echeverrigaray. Genetic variability in *Hesperozygis ringens* (Lamiaceae), an endangered aromatic and medicinal plant of Southern Brazil. *Biochem. Genet*. 2006; **44**: 479-

- 490.
36. Liu J., L. Wang, Y. Geng, Q. Wang, L. Luo and Y. Zhong. Genetic diversity and population structure *Lamiophlomis rotata* (Lamiaceae), an endemic species of Qinghai-Tibet Plateau. *Genetica*. 2006; **128**: 385-394.
37. Agostini G., S. Echeverrigaray and T.T. Souza-Chies. Genetic relationships among South American species of *Cunila* D. Royen ex L. based on ISSR. *Plant Sys. Evol.* 2008; **274**: 135-141.
38. Rahimmalek, M., B. Bahreininejad, M. Khorrami and S.B.E. Tabatabaei. Genetic variability and geographic differentiation in *Thymus daenensis* *daenensis*, an endangered medicinal plant, as revealed by inter simple sequence repeat (ISSR) markers. *Biochem. Genet.*, 2009; **47**: 831-842.