

Biodegradation of Ultra-violet Irradiated Waste Polyethylene Bags by Bacterial Community from Soil around Coal-fired Thermal Power Plant

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The current study focused on biotic degradation of waste polyethylene bags using bacterial community from hydrocarbon contaminated soil near coal fired thermal power plant and also the effect of UV irradiation on its biodegradation. The predominant groups in the bacterial community in the hydrocarbon contaminated soil near coal fired thermal power plant were identified by 16s DNA sequencing were Steroidobacter, Flavisolibacter, Planctomyces, Balneimonas, Gemmata, Alicyclobacillus, Lactobacillus, Mycobacterium, Geodermatophilus, Prevotella, Virgisporangium and Adhaeribacter. The native bacterial community from hydrocarbon contaminated soil was capable of polyethylene degradation. The bacterial community in the hydrocarbon contaminated soil metabolized 12.85 ± 0.16 percent of polyethylene (10 g/L) as sole carbon source in mineral salt media within 30 days. The UV irradiation of polyethylene enhanced weight loss of 22.80 percent higher than untreated polyethylene. The improvement in bacterial degradation by UV exposure of waste polyethylene in-vitro for 144 hrs resulted 15.78 ± 0.32 percent weight loss in 30 days. The photo-oxidation by UV irradiation of polyethylene had surface disruption and was confirmed by Field Emission Scanning Electron Microscopy (FE-SEM) and Fourier Transform Infrared Spectroscopy (FTIR). The photochemical reaction induced by UV irradiation of polyethylene resulted in formation of carbonyl peaks on polymer surface and addition as well as shifting of peaks. The morphological changes of polyethylene by UV exposure enhanced colonization, metabolism by and synergistic effect on polyethylene biodegradation by bacterial community from hydrocarbon contaminated soil.

Keywords: Bacterial Community; Carbonyl Index; Waste Polyethylene Bags; Weight Loss, Synergism; UV-Irradiation.

Polyethylene is one of the polymers with versatile applications in packaging, insulation, agricultural mulch films, domestic and industrial uses. The polyethylene is categorized based on branching into low-density polyethylene, high-density polyethylene, linear low-density

polyethylene and cross-linked polyethylene¹. The low-density polyethylene (LDPE) is the prominent component of plastic waste which accounts for 60% of the total plastic production². About 94 percent of plastic waste is thermoset plastic and of great concern to the environment³.

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The conventional methods of disposal of polyethylene waste include recycling, incineration and landfilling. The biodegradation of polyethylene waste is environment friendly and sustainable than the conventional physicochemical breakdown⁴. The biodegradation has been affected by inert and persistent nature of polyethylene for longer time and interference of hydrophobic nature in polymer availability to microorganisms⁵. The microbes with high polyethylene degradation potential were isolated from plastic contaminated sites such as garbage soil, crude oil spilled sites, plastic dumping site and soil from volcano crater^{6,7}.

The most important step in microbial degradation of polyethylene is the surface attachment of bacterial cells and biofilm formation on the polymer surface⁸. The alterations in polyethylene properties make easy availability of polyethylene for microbial biodegradation. The researchers have studied the additive effect of prior abiotic pretreatments of the polymer such as thermal oxidation, UV irradiation and chemical disintegration on microbial biodegradation of polyethylene⁹. The UV treatment of polyethylene resulted from photo-oxidative reactions by the absorption of ultraviolet radiation¹⁰. The enzymes secreted by the microbes initiate the biological degradation of petro-based polymers and break the polymer chain into oligomers and to monomers. These smaller monomeric products are metabolized in the microbial cells as carbon source¹¹. In the present study, the bacterial community from hydrocarbon contaminated soil around coal fired thermal power plant was evaluated for their ability to degrade polyethylene in liquid media in-vitro and also the effect of UV-radiation on its biodegradation.

MATERIAL AND METHODS

Collection of Waste polyethylene bags

The waste polyethylene bags were collected from the local waste dumping site in Bathinda, Punjab, India. The plastic bags were translucent and of 20 microns thickness. The plastic bags were thoroughly washed with deionized water followed by ethanol sterilization (70% v/v), drying overnight and stored aseptically for further use.

UV irradiation of polyethylene bags

The plastic bags were cut into small strips

of 1.5 × 1.5 cm, sterilized with 70% v/v ethanol for 30 minutes. The polyethylene strips were irradiated under UV lamps (16W) placing at 5 cm distance for 144 h in UV protected glass chamber.

Soil sample for polyethylene degrading bacterial community

Soil was collected from coal fired Thermal Power Plant, Bathinda, Punjab (30°13' 59.57" N and 74°55' 48.92" E) from the depth of approximately 0–10 cm in sterile zip lock plastic bags. The bacterial community used in the present study was prepared by soil enrichment method using waste polyethylene as sole carbon source in Mineral Salt media (MSM)¹². The bacterial community with mixed culture of bacteria was preserved at 4°C and used in further degradation studies. The dominant bacterial groups in the hydrocarbon contaminated soil were identified by 16s rDNA sequencing.

Biodegradation of waste polyethylene by bacterial populations

The degradation study was performed with untreated and UV irradiated waste polyethylene (1% w/v) added to mineral salt media as a sole source of carbon and energy at 30°C and continuous agitation of 150 rpm for an incubation period of 1 month. The 16 h old bacterial populations were grown in Luria-Bertani broth was used as the inoculum for degradation studies. The bacterial growth and biodegradation study was investigated by culturing polyethylene enriched medium containing NH₄NO₃ (1.0 gL⁻¹); K₂HPO₄ (1.0 gL⁻¹); KCl (0.15 gL⁻¹); MgSO₄·7H₂O (0.2 gL⁻¹); CaCl₂·2H₂O (0.1 gL⁻¹) and yeast extract (0.1 gL⁻¹) along with 1.0 mgL⁻¹ of ZnSO₄·7H₂O, FeSO₄·6H₂O, and MnSO₄¹³. The samples were collected at 5 days intervals for estimation of biomass growth of bacterial consortium. The weight loss of polyethylene samples from the culture media was estimated¹⁴ after washing with sodium dodecyl sulfate (2% (v/v) followed by distilled water¹⁵. All the experiments were conducted in triplicates and the results were the mean values with standard deviations. The results were statistically analyzed by single factor Analysis of Variance followed by Post-hoc Tukey test.

Quantification of bacterial adherence on the polyethylene

The bacterial populations on the polyethylene surface were measured indirectly

by determining the concentration of extractable protein. The protein estimation by Lowry's method was followed for the supernatant obtained from boiling polyethylene samples in 0.5 N NaOH for 30 min¹⁶.

Structural changes on polyethylene surface

The Fourier Transform Infrared Spectroscopy (FTIR) and Scanning Electron Microscope (SEM) analysis of untreated polyethylene, UV irradiated polyethylene and samples of polyethylene films after biotic degradation for 30 days were studied for understanding structural changes. The change in the polyethylene surface and generation of new peaks were obtained by FTIR spectroscopy (FTIR Bruker, Model: Tensor 27) from 400 to 4000 cm^{-1} . Using the FTIR spectrum, carbonyl residue as carbonyl index and double bond index were measured on the basis of ratio of relative intensities of the carbonyl band (1715 cm^{-1}) and double bond band (1650 cm^{-1}) to that of methylene scissoring band at 1460 cm^{-1} . The bacterial growth and structural changes on polyethylene was checked by SEM analysis as per the method given by Tribedi and Dey¹⁷.

RESULTS AND DISCUSSION

The hydrocarbon contaminated soil from the coal fired Thermal Power Plant, Bathinda used for screening polyethylene degrading bacterial community was slightly alkaline with pH 8.10 ± 0.08 and electric conductivity of $556 \pm 1.8 \mu\text{S/cm}$. The total petroleum hydrocarbon content present

in contaminated soil affected the physicochemical properties¹⁸. The soil sample had bacterial population of 3.11×10^9 CFU/g (Figure 1).

The 16s rDNA sequencing of native bacterial community from hydrocarbon contaminated soil near coal fired thermal power plant identified predominant bacterial genera as *Steroidobacter*, *Flavisolibacter*, *Planctomyces*, *Balneimonas*, *Gemmata*, *Alicyclobacillus*, *Lactobacillus*, *Mycobacterium*, *Geodermatophilus*, *Prevotella*, *Virgisporangium* and *Adhaeribacter*. The bacterial populations in the hydrocarbon contaminated soil were able to grow using polyethylene as sole carbon and energy source. It was clear from the protein concentration at 5 days with an increase of 3.69 times than 0 h. The continuous increase in protein concentration over time from the colonization of microbial populations on the polyethylene surface. The increase of protein concentration was 12.93 times and 25.98 times at 15 days and 30 days respectively than the inoculation. But the increase in biomass growth was declined to 44 percent in 10th day than 5th day (Graph 1). The increase was 2.44 times in the next 5 days which was again reduced to 59 percent in 20th day of biodegradation study. Further the percent increase was reduced to 9.69 percent in 30th day of incubation.

The biomass growth using polyethylene as carbon source and biodegradation was confirmed from the 12.85 ± 0.16 percent weight loss of waste polyethylene bags in 30 days. The range of weight loss in polyethylene after

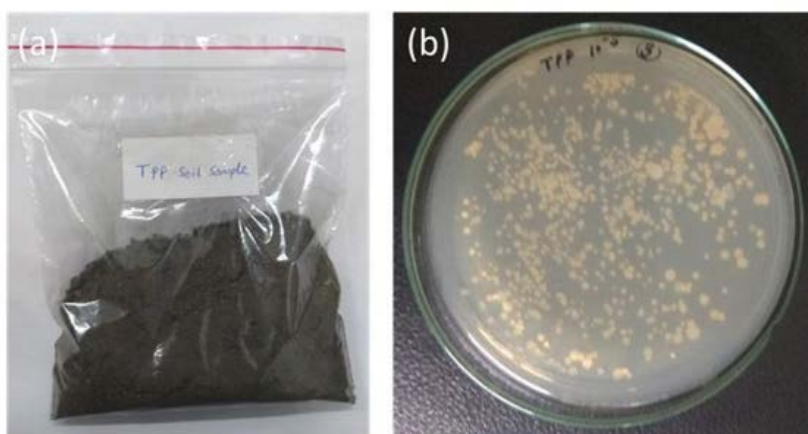


Fig. 1. (a) Hydrocarbon contaminated soil from Thermal power plant for polyethylene degrading bacteria (b) Viable bacteria from hydrocarbon contaminated soil on nutrient agar plates.

biodegradation of 30 days was from 12.67 to 13.05 percent. This weight loss of polyethylene by bacterial community from hydrocarbon contaminated soil from coal fired thermal power plant was higher than LDPE biodegradation in 60 days by *Pseudomonas aeruginosa* (ISJ14) from waste dumping site¹⁹. The polyethylene degradation was increased by different microbes than pure culture²⁰. The microbial populations present in the hydrocarbon contaminated sites had high biosurfactant production capability and presence of alkane hydroxylase gene for competent biodegradation²¹. The degradation of polyethylene was due to interaction between different polyethylene degrading microorganisms²².

The UV irradiation increased the bacterial biomass (protein concentration) to 61.89 percent than untreated polyethylene films in 5 days and 33.82 percent in 30 days of degradation. The protein concentration of bacterial community from utilizing UV treated polyethylene as carbon source was 24.30 and 38.82 times higher in 15th day and 30th day of degradation respectively than the day of inoculation. The initial growth was also higher in UV treated polyethylene with 6.67 times in 5 days of inoculation. The rate of growth in UV treated polyethylene was doubled in 10 days which was reduced by 65.56, 24.45, 15.41 and 11.2 percent respectively for 15, 20, 25 and 30 -days incubation period. The UV exposure of waste polyethylene

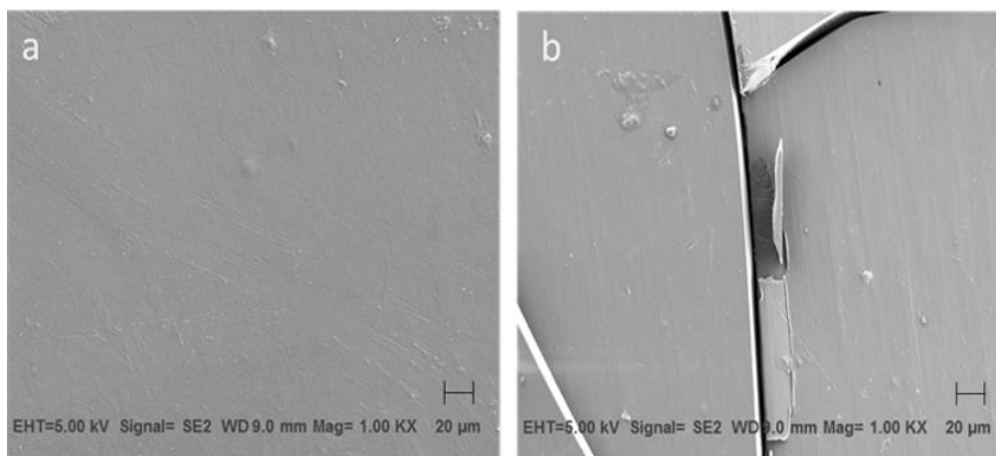


Fig. 2. SEM images of (a) Untreated polyethylene (b) UV irradiated polyethylene for 144 h

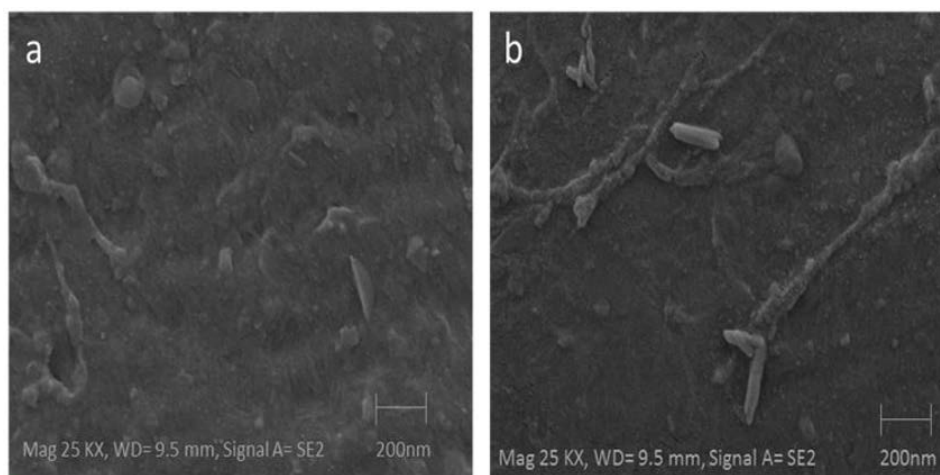


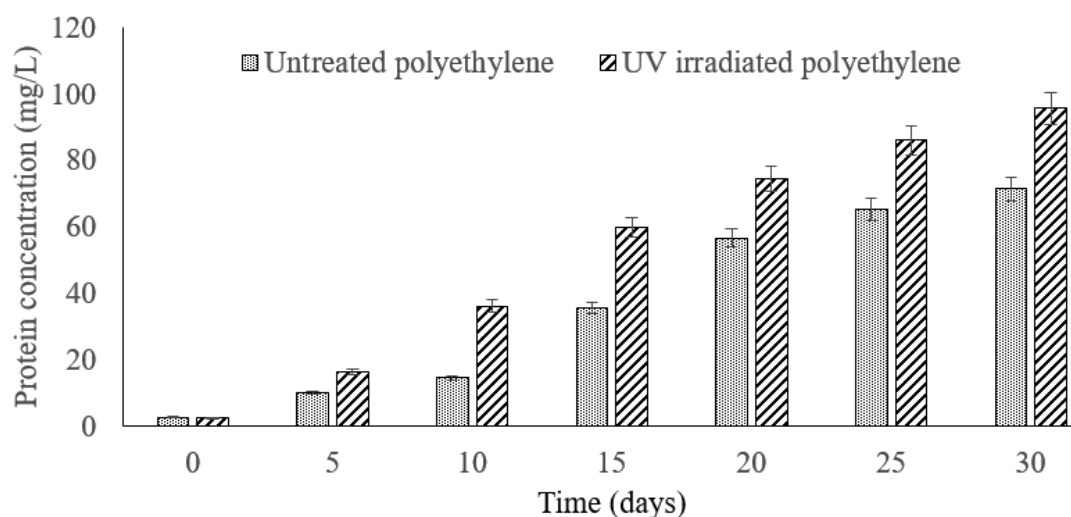
Fig. 3. SEM images of bacterial growth in 30 days on (a) untreated polyethylene (b) UV irradiated polyethylene.

bags improved initial growth and more bacterial colonization on polymer surface.

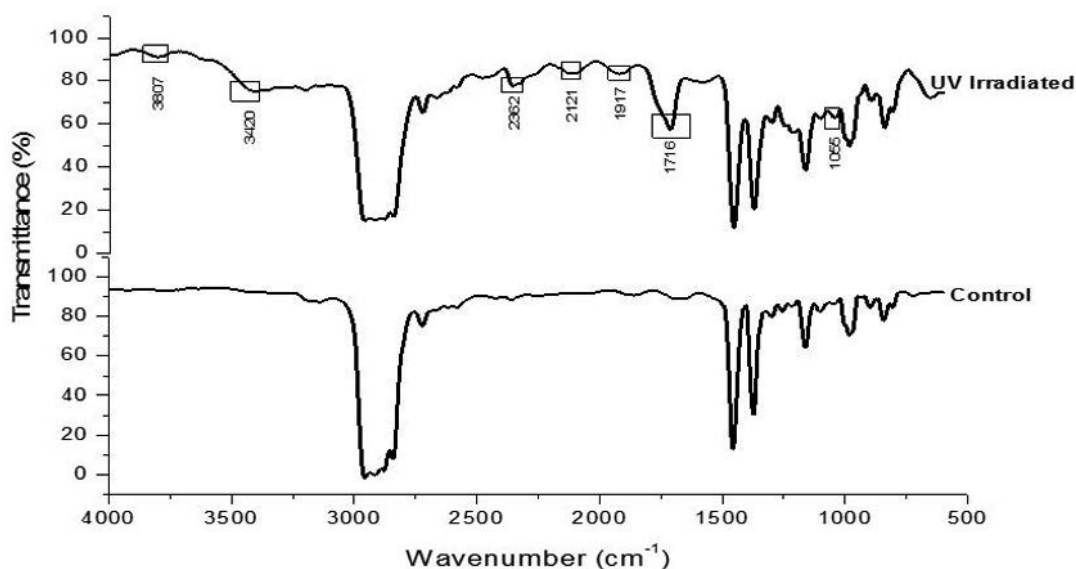
The increase in biomass growth of bacterial community from hydrocarbon contaminated soil utilizing UV irradiated polyethylene as carbon and energy source was confirmed with biodegradation of 15.81 ± 0.32 percent weight loss of waste polyethylene bags in 30 days. The polyethylene after biodegradation had a weight loss ranged from 15.43 to 16.22 percent in 30 days. The

UV irradiation on bacterial biodegradation of polyethylene bags was statistically found highly significant at P value <0.01 by one way ANOVA with Post-hoc Tukey test. The biomass growth was also found significant (P value <0.01) with the UV treatment of polyethylene bags.

The UV irradiation increased the carbonyl index and caused significant alteration on the polymer structure that favored the enhanced microbial attachment on polymer surface. The



Graph 1. Protein concentration of bacterial population as biomass indicator on polyethylene surface using nontreated and UV irradiated polyethylene films.



Graph 2. FTIR spectra of non-treated and UV irradiated polyethylene films.

increased degradation of polyethylene was also reported by Jeon and Kim²³ and Montazer and coworkers²⁴.

The structural modifications of UV treated polyethylene was clear from FTIR spectra. FTIR spectra of nontreated polyethylene films and UV irradiated films is shown in Graph 2. The infrared spectrum of the waste polyethylene bags after photooxidation (UV irradiation for 144 h) showed a peak at 1716 cm^{-1} and 1055 cm^{-1} representing carbonyl groups (-C=O) and ether groups (-C-O-C-) respectively. The generation of additional peaks at 3807 cm^{-1} , 3420 cm^{-1} , 2362 cm^{-1} , 2121 cm^{-1} and 1917 cm^{-1} after 144 h of UV irradiation also confirmed the structural changes in the polymer. The functional groups identified in the FTIR spectra were indicator precursors in the photochemical reactions of the polyethylene²⁵. The prior UV pretreatments of polyethylene increased surface hydrophilicity by formation of additional carbonyl groups. The radical was formed initially from absorption of UV-radiation on polyethylene and further to hydro-peroxide formation and then terminal carbonyl groups²⁶.

The carbonyl index showed the oxidation of polyethylene by UV radiation from the carbonyl species in the FTIR spectra. The UV pretreatment of the polyethylene for 144 h enhanced the ester carbonyl index and keto carbonyl index by 7.97 and 9.49 percent respectively. The increase in carbonyl index from UV treatment was due to photo-oxidative reaction of plasticizer and stabilizers present in polyethylene that later leads to chain scission²⁷.

The UV irradiation of waste polyethylene bags also increased the brittleness of the polyethylene, surface roughness, cracks and depressions on the polymer surface as clear from scanning electron microscopy (SEM) images (figure 2). The structural changes on the UV irradiated polyethylene enhanced bacterial consortium from hydrocarbon contaminated soil to improve degradation by 22.80 percent higher than nontreated waste polyethylene films *in vitro*.

The SEM images after 30 days biodegradation study was also confirmed the bacterial biodegradation of polyethylene (figure 3). The adherence of microorganisms on the surface of plastics followed by the colonization of the exposed surface are the major mechanisms involved in the

microbial degradation of plastics²⁸. The microbes adhered to the surface increased biodegradation and cause cracks and cavities²⁹.

The decrease in pH in the growth media with polyethylene evidenced the metabolism and the biodegradation process in four weeks period by bacterial populations from hydrocarbon contaminated soil. This was also in accordance with reports of Das and Kumar³⁰.

The weight loss of untreated polyethylene without pro-oxidant additives and oxidative pretreatment by UV irradiation of polyethylene confirmed the biodegradation potential of bacterial community from hydrocarbon contaminated soil. The bacterial populations in the community from hydrocarbon contaminated soil had the polyethylene degrading ability from surface adhesion, bacterial proliferation and exopolysaccharide production³¹. The interactions among bacterial populations in the community with different oxidative enzymes resulted in higher polyethylene biodegradation^{32,33}.

The improvement in weight loss of the waste polyethylene by UV treatment and biotic degradation confirmed the synergistic effect from photooxidation by UV radiation and biodegradation by bacterial community from hydrocarbon contaminated soil²⁴. The similar results were also reported by Albertsson et al.²⁶ and Esmaeili et al.³⁴.

The UV light exposure induced the photooxidation of the polymer and generated the free radicals that facilitate the microbial attack^{26, 35,36}. The photooxidation by UV irradiation increased carbonyl bond and terminal double-bond index which in turn increased microbial utilization of carbonyl residues^{17,37,38}. The biodegradation was initiated with adherence of microbes on surface of polyethylene³⁹. The biodegradation of these recalcitrant pollutants was also enhanced by the surfactants produced by the microbes which increase bioavailability⁴⁰. The enzymes present in the polyethylene degrading microorganisms from hydrocarbon contaminated soil cleave the polymer chain into monomers and smaller fragments that can be easily used up by the microorganism^{31,41}. This depolymerization process with intracellular and extracellular depolymerases helped microbes from hydrocarbon contaminated soil to utilize polyethylene as carbon and energy sources^{23, 42,43}. The products were easily metabolized by

microbes via the α -oxidation pathway and citric acid cycle⁴⁴. The biodegradation efficiency was improved in UV irradiated polyethylene and with microbial communities resulted in more microbial colonization and synergistic effect of photooxidation and biodegradation^{26,45,46}. Microbial biodiversity inhabiting contaminated sites have already proved high efficiency to degrade polyethylene. The nature and type of polyethylene, types of additive present in the polymer and extent of prior photooxidation pretreatment play a crucial role in biodegradation⁴⁷.

CONCLUSION

The hydrocarbon contaminated soil from coal fired thermal power plant was a good habitat for different efficient bacterial populations capable of degrading polyethylene wastes. The prior UV irradiation resulted in increased bacterial adherence, colonization and synergism in polyethylene biodegradation. The combination of photo-oxidation and biodegradation helps in easy metabolism of polyethylene and in plastic waste management without any ecological impacts.

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Conflict of interest

The authors declare that there is no conflict of interest.

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