

Classification and Replication Mechanism of *Staphylococcus* Phage

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The use of antibiotics is causing high resistance in bacterial cell. One of the major pathogen that cause of dangerous infection is *staphylococcus aureus*, these bacteria can urgently resistance to new generation of antibiotic such methicillin. Hence recognize the bacterial genome and mobile element such lysogenic phage and understanding the pathogenesis of pathogen is important. In this review we investigated the *Staphylococcus aureus* phages and their role in virulence gene transfer.

Key word: Phage, classification of Phage, staphylococcus Phage

Staphylococcus aureus is a Gram-positive bacterium that causes a serious infection such as, pneumonia¹, bovine mastitis, morbidity and mortality to humans². *S. aureus* are resistant to multiple antibiotics and reports that some had acquired high level to new antibiotic generation such vancomycin³⁻⁴. *S. aureus* encoded virulence factors such a coagulase⁵, accessory gene regulator A (*agrA*), staphylococcal protein A (*spa*) and *sarA* to the development of disease⁶. Staphylococci are classified via different methods, one of the major categories on the basis of coagulase. Coagulase is a virulence factor that causes blood clotting in the proximity of the pathogen bacteria (see figure 1)⁷.

Methicillin-resistant *S. aureus* (MRSA) are more challenging because is resistance to commonly used antibacterial agents. The *mecA*

gene is liable for the increased antibiotic resistance of MRSA and encodes PBP2a, which is a penicillin-binding protein with low-binding affinity and which mediates methicillin resistance⁸.

Phage

Phages are kind of viruses that attack and destroy of the bacteria. There are at least 12 distinct groups of phages and each phage is specific to its bacterial host. The morphology and genetic material (DNA or RNA) varies according to the phage species. Phage are commonly find in the environment: there are more than 10¹⁰ phage per liter of surface seawater⁹. phage have been characterized by their host range and the physical characteristics of the free virion, including capsid size, resistance to organic solvents, shape, structure and genome size and type such as single-stranded DNA (ssDNA), ssRNA, double-stranded (dsDNA) and dsRNA [10]. phages are known to augment wherever their bacterial hosts exist¹¹ but the phage can exist freely outside the bacterial host, however all phages like mostly virus are obligate intracellular parasites and need their host to propagate¹². The International Committee for Taxonomy of Viruses (ICTV) require phage capsid

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morphology to be established for their formal classification¹⁰ but the phage may exist as a lysogenic prophage that does not produce mature virions¹³. Bacterial classification easily done by examining the con-served 16S ribosomal genes¹⁴ but phage lack ribosomal DNA and there are no conserved gene common to all phage on which to base a classification¹⁰. According to the ICTV system, bacteriophages are classified into one order, *Caudovirales* which consists of three related families. Phage virions can be tailed, filamentous, polyhedral and pleomorphic and most of them contain dsDNA¹⁵.

Life cycle of bacteriophage

Phages can be categorized into Lytic and Lysogenic phages. This is specified by the events that follow injection of nucleic acid into the bacterial cell. Lytic phages usually lead to the release of phage through the host cell bursts¹⁶. Lysogenic phages have two pathways: first they have ability to undergo lysis in their host cell, where by their new generation of phage is released into the environment. However, they can establish stable

relationships with the host in which lytic genes are not expressed and their genome becomes integrated into the bacterial chromosome, and is replicated along with the host cell DNA¹⁷. Phage can also constitute major vehicles for horizontal gene transfer¹⁸. Phage also has a major role in virulence by encoding numerous virulence factors and by their movements within genomes¹⁹.

However temperate phages have an alternative life-cycle that is absent from the reproduction of lytic bacteriophages, whereby the bacterial host have the phage genome and then replicates it during bacterial cell division. This phage genome in host's chromosome expresses resistance to infection by the same phage, but not to infection by heterologous phages²⁰. During lysogeny, which follows recombination of the phage genome into the bacterial chromosome; most of the phage genes are repressed²¹. In some prophages with low G+C, phage conversion genes proposed virulence and fitness genes²². Lysogeny is not a constant state and during bacterial growth phage arises due to spontaneous prophage

Table 1. *Staphylococcus aureus* phages studied and classified

Family	Author- year	Title	Conclusion
<i>Siphoviridae</i>	Aswani VH et al. 2014	Complete Genome Sequence of a <i>Staphylococcus epidermidis</i> Bacteriophage Isolated from the Anterior Nares of Humans [33]	By examining the entire genome of the phage was observed that the phage has an icosahedral capsid and unusually long non-contractile tail.
	Hongying Jia et al. 2013	Complete Genome Sequence of <i>Staphylococcus aureus</i> Siphovirus Phage JS01 [34]	Isolated phage from milk and use the TEM showed that the phage has a long non-contractile tail and an icosahedral head.
<i>Podoviridae</i>	Swift SM et al. 2014	Complete Genome Sequence of <i>Staphylococcus aureus</i> Phage GRCS [35]	By examining the hole genome of the phage suggested that these could be used as phage-therapy.
<i>myoviridae</i>	Leila Kvachadze et al. 2014	Evaluation of lytic activity of staphylococcal bacteriophage Sb-1 against freshly isolated clinical pathogens [36]	The Phage Sb-1 has high spectrum of lytic activity against staphylococcus.
	Katrien Vandersteegen et al. 2013	Romulus and Remus, Two Phage Isolates Representing a Distinct Clade within the Twortlikevirus Genus, Display Suitable Properties for Phage Therapy Applications [37]	Two phages have a lytic activity against 70% of staphylococcus aureus isolates and both phage shown biofilm-degrading capacity.

induction. The condition that which prophages enter the lytic cycle depend on chemical or physical agents that damage DNA, including oxidants, antibiotics like mitomycin C, and UV radiation can induce prophage entry into the lytic cycle²³. The first unit of the prophage is the leftward-transcribed integrase/cI region for the maintenance of phage in lysogeny phase. A large rightward-transcribed region encoding proteins for the lytic pathway including replication, head/tail morphogenesis, packaging and lysis functions is followed by a leftward-transcribed region.

Phages have several potential conversion genes, also called cargo²⁴ and appears to have a set of lysins immediately followed by three leftward-transcribed genes, two of which encode a supposed membrane protein and a lysM domain-containing protein²⁴. These proteins can locate on the host cell surface. Expression of these proteins during growth represents phage conversion genes. If the phage has a serine recombinase (ORF1) a repressor (ORF6) and anti-repressor (ORF7) indicating that the phage is temperate²⁵. Repressor is a self-assembling dimer also known as the cI

protein²⁶ and binds to DNA in the helix-turn-helix binding motif. cI protein regulates the transcription of the cI protein and the Cro protein. The life cycle of phages is controlled by cI and Cro proteins and phage will remain in the lysogenic state if cI proteins predominate, but can transformed into the lytic cycle if cro proteins predominate. Auto-negative regulation causes a stable minimum concentration of the repressor molecule and, if SOS signals arise, allows for more efficient prophage induction²⁷. That mean In the presence of cI proteins, only the cI gene transcribed and in contrast in absence of cI protein the cro gene is transcribed.

Staphylococcus phages

Phages are widespread in *Staphylococcus aureus* genus and have been extensively studied²⁸. The phage was firstly used for the typing of clinical *S. aureus* isolates²⁹. The majority of *S. aureus* phages known so far are double-stranded DNA belonging to the *Siphoviridae* family of the *Caudovirales* order³⁰. Staphylococcal *Siphoviridae* are composed of an icosahedral capsid and a non-contractile tail³¹. *Staphylococcal Podoviridae*, such SAP-2 phage are composed of

Table 2. Toxins and their functional status

Name of gene or toxin	Performance
sak	immune modulator
chp	chemotaxis inhibitory protein CHIP
sea	Enterotoxins (cause diarrhea, arthritis, atopic dermatitis and toxic shock) [40]
Seg, seh, sei	Enterotoxins [41]
Panton-Valentine leukocidin (PVL)	Cytotoxin [42]
eta (exfoliative toxin A)	staphylococcal scalded-skin syndrome [43]

Table 3. Classification of phage family of *Staphylococcus aureus* (all data extracted from NCBI with *Staphylococcus aureus* phage keyword)

Phage family	Author-Name of phage- journal
Siphoviridae	Deghorain M_ StB12, StB27, and StB20_ Journal of Bacteriology [46]
	Mariem BJ_ phi7401PVL_ BMC Microbiology [47]
	Zhang M_φ7247PVL, φ5967PVL_ FEMS Microbiology Letter [48]
	Kim MS_SA11_ Journal of Virology [49]
	Diana Gutiérrez_vB_SepiS-phiIPLA5 and vB_SepiS-phiIPLA7_ BMC Genomics [50]
Myoviridae	G.E. Christie_80 and 80φ_ Virology [51]
	Zelin Cui_JD007_ Journal of Virology [52]
Podoviridae	Jingmin Gu_GH15_ Journal of Virology [53]
	Tony Kwan_3A and 26 phage_ PNAS[54]
	Son JS_SAP-2_ Appl Microbiol Biotechnol [32]

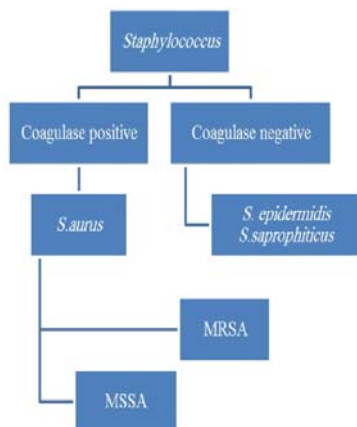



Fig. 1. Differentiation of *S. aureus* based on Coagulase test



Fig. 2. Modular organization of phage genome, if the suppressor  expressed the phage remains in lysogenic phase but if de-repressor activated (SOS response or oxidative stress) phage can be activated and enter the lytic phase

Phages are carrying single virulence factor genes, however some exceptions have been reported³⁹ and this gene not strictly associated to a specific phage and appear to be transfer horizontally⁴⁴. Virulence genes are often located near the attachment site (att) of the prophage in host chromosome. Phage responsible for the mobilization of *Staphylococcus aureus* Pathogenicity Islands (SaPIs), which encodes major toxin genes such as the toxic shock syndrome toxin 1 and other super-antigens⁴⁵. SaPIs are not mobile by themselves and need the helper phage for moving. The mechanism for induction is the specific interaction of a SaPI repressor and a de-repressor encoded by the helper phage. Different proteins of a helper phage may be involved in induction of different SaPIs.

CONCLUSION

One of the major problems by modern medicine is the spread of antibiotic resistant genes among pathogenic bacteria, as is seen with methicillin resistance in the species

a small icosahedral capsid, short, non-flexible and non-contractile tail³². Finally *Myoviridae* are characterized by an icosahedral capsid and a long contractile tail.

Virulence factor and phage

Phages encode a some of *S. aureus* virulence factors and variety of toxins, mainly allowing bacterial cell escaping from host immune system³⁸ such a widespread immune modulator staphylokinase (sak) responsible for host tissue destruction, the staphylococcal inhibitor of complement SCIN (scn) and chemotaxis inhibitory protein CHIP(chp). Phage also encode a several super-antigen such a: (sea) and (seg), these super-antigens are enterotoxins causing food poisoning, necrotizing fasciitis and toxic shock syndrome³⁹.(To more information see table 2)

Staphylococcus aureus. *Staphylococcus aureus* has an extraordinary range of virulence factors that allows it to survive extreme conditions within the human host. This bacterium has a lot of toxins that some of them are transferred by phage. Hence identification of phage types and toxins moved by their phage is important. Phages are the primary vehicles for horizontal gene transfer and mobilization of SaPIs.

Comparison of pathogen *Staphylococcus aureus* with non-toxicogenic shown Phage-encoded virulence factors responsible for *S. aureus* pathogenesis are absent in non-*S.aureus*. With identify and classify the phage and how they move and change the genetic of bacterial host, their movements can be investigated. Hence phage can move the virulence factor from pathogen bacteria to non-pathogen bacteria and conversion it to pathogen bacterial cell.

REFERENCES

1. Pannaraj, P.S., *et al.*, Infective pyomyositis and myositis in children in the era of community-

- acquired, methicillin-resistant *Staphylococcus aureus* infection. *Clin Infect Dis*, 2006; **43**(8): p. 953-60.
2. Otto, M., Basis of virulence in community-associated methicillin-resistant *Staphylococcus aureus*. *Annu Rev Microbiol*, 2010; **64**: p. 143-62.
 3. Weigel, L.M., *et al.*, Genetic analysis of a high-level vancomycin-resistant isolate of *Staphylococcus aureus*. *Science*, 2003; **302**(5650): 1569-71.
 4. LG., M., *et al.*, Necrotising fasciitis caused by community-associated methicillin resistant *Staphylococcus aureus* in Los Angeles. *N Engl J Med*, 2005; **352**: 1445-1453.
 5. Sawai, T., K. Tomono, K. Yanagihara, Y. Yamamoto, M. Kaku, Y. Hirakata, and T.T. H. Koga, and S. Kohno., Role of coagulase in a murine model of hematogenous pulmonary infection induced by intravenous injection of *Staphylococcus aureus* enmeshed in agar beads. *Infect. Immun*, 1997; **65**: 466-471.
 6. Gomez, M.I., *et al.*, *Staphylococcus aureus* protein A induces airway epithelial inflammatory responses by activating TNFR1. *Nat. Med*, 2004; **10**: 842-848.
 7. Fatemeh Rezaei, *et al.*, Using Phage as A Highly Specific Antibiotic Alternative Against Methicillin Resistance *Staphylococcus aureus* (MRSA). *Biosciences Biotechnology Research Asia*, 2014; **11**(2): 523-529.
 8. Chambers, H.F., The changing epidemiology of *Staphylococcus aureus*? *Emerg Infect Dis*, 2001; **7**(2): 178-82.
 9. Bergh, Ø., *et al.*, High abundance of viruses found in aquatic environments. *Nature* 1989; **340**: 467-468.
 10. Rohwer, F. and R. Edwards, The Phage Proteomic Tree: a genome-based taxonomy for phage. *J Bacteriol*, 2002; **184**(16): 4529-35.
 11. Hendrix, R.W., *et al.*, Evolutionary relationships among diverse bacteriophages and prophages: All the world's a phage. *Proc Natl Acad Sci*, 1999; **96**: 2192-2197.
 12. Jensen, E.C., *et al.*, Prevalence of broad-host-range lytic bacteriophages of *Sphaerotilus natans*, *Escherichia coli*, and *Pseudomonas aeruginosa*. *Appl Environ Microbiol*, 1998; **64**(2): 575-80.
 13. Ferretti, J.J., *et al.*, Complete genome sequence of an M1 strain of *Streptococcus pyogenes*. *Proc Natl Acad Sci U S A*, 2001; **98**(8): 4658-63.
 14. Nelson, D., Phage Taxonomy: We Agree To Disagree. *J. Bacteriol*, 2004; **186**(21): 7029-7031.
 15. Kakoma, K., Isolation and Characterisation of Bacteriophages and Their Potential Use for the Control of Bacterial Infections in Poultry, in Department of Microbial, Biochemical and Food Biotechnology University of the Free State. Bloemfontein: South Africa 2009; 158.
 16. Engelkirk, P.G. and G.R. Burton, Burton's microbiology for the health sciences, ed. t. Edition. Lippincott Williams & Wilkins, 2006.
 17. Little, J.W., Lysogeny, prophage induction, and lysogenic conversion. Phages, ASM Press: 2005; 37-54.
 18. Lindsay, J.A., Genomic variation and evolution of *Staphylococcus aureus*. *Int J Med Microbiol*, 2010; **300**(2-3): 98-103.
 19. Goerke, C., *et al.*, Extensive phage dynamics in *Staphylococcus aureus* contributes to adaptation to the human host during infection. *Mol Microbiol*, 2006; **61**(6): 1673-85.
 20. EA., B., Bacterial and Bacteriophage Genetics, ed. r. ed. New York: Springer-Verlag, 1994.
 21. M., P., A Genetic Switch: Phage Lambda Revisited. 3rd ed ed., New york: Cold Spring Harbor, 2004.
 22. Desiere, F., *et al.*, Comparative genomics of phages and prophages in lactic acid bacteria. *Antonie van Leeuwenhoek*, 2002; **82**(1-4): 73-91.
 23. Breck A. Duerkop, K.L.P., and Malcolm J. Horsburgh, Enterococcal Bacteriophages and Genome Defense. *Europe Pubmed Central*. 2014.
 24. Stevens RH, Ektefaie MR, and F. DE., The annotated complete DNA sequence of *Enterococcus faecalis* bacteriophage phiEf11 and its comparison with all available phage and predicted prophage genomes. *FEMS Microbiol Lett*, 2011; **317**(1): 9-26.
 25. Keary, R., *et al.*, Genome analysis of the staphylococcal temperate phage DW2 and functional studies on the endolysin and tail hydrolase. *Bacteriophage*, 2014. **4**: e28451.
 26. Burz, D.S., *et al.*, Self-assembly of bacteriophage lambda cI repressor: effects of single-site mutations on the monomer-dimer equilibrium. *Biochemistry*, 1994; **33**(28): 8399-405.
 27. Ptashne, M., A Genetic Switch. New York: Cold Spring Harbor Laboratory Press, 2004.
 28. Feng, Y., *et al.*, Evolution and pathogenesis of *Staphylococcus aureus*: lessons learned from genotyping and comparative genomics. *FEMS Microbiol Rev*, 2008; **32**(1): 23-37.
 29. Rosenblum, E.D. and S. Tyrone, Serology, Density, and Morphology of Staphylococcal Phages. *J Bacteriol*, 1964; **88**: p. 1737-42.
 30. Canchaya, C., *et al.*, Prophage Genomics. *Microbiol. Mol Biol Rev*, 2003. **67**: p. 238-276.

31. Ackermann, H.W., Tailed bacteriophages: the order caudovirales. *Adv Virus Res*, 1998; **51**: 135-201.
32. Son, J.S., *et al.*, Antibacterial and biofilm removal activity of a podoviridae *Staphylococcus aureus* bacteriophage SAP-2 and a derived recombinant cell-wall-degrading enzyme. *Appl Microbiol Biotechnol*, 2010; **86**(5): 1439-49.
33. VH., A., *et al.*, Complete Genome Sequence of a *Staphylococcus epidermidis* Bacteriophage Isolated from the Anterior Nares of Humans. *Genome Announc*, 2014; **7**(2).
34. Hongying Jia, *et al.*, Complete Genome Sequence of *Staphylococcus aureus* Siphovirus Phage JS01. *Genome Announcements*, 2013; **1**(6).
35. SM., S. and N. DC., Complete Genome Sequence of *Staphylococcus aureus* Phage GRCS. *Genome Announc*, 2014; **10**(2).
36. Leila Kvachadze, *et al.*, Evaluation of lytic activity of staphylococcal bacteriophage Sb-1 against freshly isolated clinical pathogens. *Microbial Biotechnology*, 2011; **4**(5): p. 643-650.
37. Katrien Vandersteegen., *et al.*, Romulus and Remus, Two Phage Isolates Representing a Distinct Clade within the Twortlikevirus Genus, Display Suitable Properties for Phage Therapy Applications. *Journal of Virology*, 2013; **87**: 3237-3247.
38. Goerke, C., *et al.*, Diversity of prophages in dominant *Staphylococcus aureus* clonal lineages. *J Bacteriol*, 2009; **191**(11): p. 3462-8.
39. Deghorain, M. and L. Van Melderen, The *Staphylococci* phages family: an overview. *Viruses*, 2012; **4**(12): 3316-35.
40. Rasooly, R., P.M. Do, and M. Friedman, Inhibition of biological activity of staphylococcal enterotoxin A (SEA) by apple juice and apple polyphenols. *J Agric Food Chem*, 2010; **58**(9): 5421-6.
41. Zschock, M., *et al.*, Pattern of enterotoxin genes seg, seh, sei and sej positive *Staphylococcus aureus* isolated from bovine mastitis. (0378-1135 (Print)).
42. Endo, Y., *et al.*, Phage conversion of exfoliative toxin A in *Staphylococcus aureus* isolated from cows with mastitis. *Vet Microbiol*, 2003; **96**(1): p. 81-90.
43. Amagai, M., *et al.*, *Staphylococcal Exfoliative Toxin B Specifically Cleaves Desmoglein 1*. 2002; **118**(5): 845-850.
44. Brussow, H., C. Canchaya, and W.D. Hardt, Phages and the evolution of bacterial pathogens: from genomic rearrangements to lysogenic conversion. *Microbiol Mol Biol Rev*, 2004; **68**(3): 560-602, table of contents.
45. Novick, R.P., G.E. Christie, and J.R. Penades, The phage-related chromosomal islands of Gram-positive bacteria. *Nat Rev Microbiol*, 2010; **8**(8): 541-51.
46. Deghorain, M., *et al.*, Characterization of novel phages isolated in coagulase-negative staphylococci reveals evolutionary relationships with *Staphylococcus aureus* phages. *J Bacteriol*, 2012; **194**(21): 5829-39.
47. Mariem, B.J., *et al.*, Molecular characterization of methicillin-resistant Panton-valentine leukocidin positive *Staphylococcus aureus* clones disseminating in Tunisian hospitals and in the community. *BMC Microbiol*, 2013; **13**(2): p. 2.
48. Meng Zhang, *et al.*, Identification of the third type of PVL phage in ST59 methicillin-resistant *Staphylococcus aureus* (MRSA) strains. *FEMS Microbiol Lett*, 2011; **323**: p. 20-28.
49. Min Soo Kim and Heejoon Myung, Complete Genome of *Staphylococcus aureus* Phage SA11. *Journal of Virology*, 2012. **86**.
50. Gutiérrez D, *et al.*, Genomic characterization of two *Staphylococcus epidermidis* bacteriophages with anti-biofilm potential. *BMC Genomics*, 2012; **13**.
51. G.E. Christia, *et al.*, The complete genomes of *Staphylococcus aureus* bacteriophages 80 and 80á– implications for the specificity of SaPI mobilization. *Virology*, 2010; **407**(2): 381-390.
52. Zelin Cui, *et al.*, Complete Genome Sequence of Wide-Host-Range *Staphylococcus aureus* Phage JD007. *Journal of Virology*, 2012; **86**.
53. Jingmin Gu, *et al.*, Complete Genome Sequence of *Staphylococcus aureus* Bacteriophage GH15. *Journal of Virology*, 2012; **86**: 8914–8915.
54. Tony Kwan, *et al.*, The complete genomes and proteomes of 27 *Staphylococcus aureus* bacteriophages. *PNAS*, 2005; **102**.