Antifungal Activity of Verbenaceae

Rogimon P. Thomas^{1*}, Manuel Thomas², Joby Paul¹ and Mahesh Mohan¹

¹School of Environmental Sciences, Mahatma Gandhi University, Kottayam - 686 560, India.

*Department of Botany, CMS College, Kottayam, Kerala - 686 001, India.

²Consortium & Training Academy for Biosciences (CTAB)IInd Floor,

Geo Printers Building, Manarcadu P.O., Kottayam - 686 019, India.

(Received: 15 April 2013; accepted: 25 May 2013)

Traditional curatives based on herbal medicinal rationales are time tested and widely accepted across various cultural and socioeconomic strata all around the world. The emergence of drug resistant germs makes the circumstances more assailable for emerging pathogens, especially fungi. Thus an attempt to assess the spectrum of antifungal activity among selected Verbenaceae members (*P. serratifolia, L. nodiflora, V. trifolia, V. negundo* var. *negundo, V. negundo* var. *pubescens, V. altissima, V. leucoxylon, V. pinnata, S. jamaicensis, C. inermis* and *C. serratum*) was conducted against a panel of 8 fungi (*Aspergillus flavus, A. fumigatus, A. niger, Penicillium* sp., *Verticillium* sp., *Curvularia* sp., *Mucor* sp. and *Fusarium* sp.). All of the tested plants were inhibitory to many of the tested fungi and both of the extracts (petroleum ether and methanolic) showed appreciable antifungal effects, though the phytochemical composition was dissimilar.

Key words: Verbenaceae, Antifungal, Fungi, Antimicrobial.

Traditional therapeutics based on herbal medicinal principles is time tested and widely accepted across various cultural and socioeconomic strata all around the world. A number of therapeutic drugs which are currently available in market have plants as origin. In short, the plant kingdom is a treasure house of potential drugs and in recent years there has been a burgeoning interest among the scientific community. Drugs from the plants are easily available, less expensive, safe, and efficient and seldom have side effects. The plants which have been selected for medicinal use over thousands of years constitute the most axiomatic choice for examining the current search for therapeutically effective new drugs such as anticancer drugs, (Haldar et al., 2010), antimicrobial drugs (Namita and Mukesh, 2012), antidiabetic drugs (Rao et al., 2001) and anti hepatotoxic compounds (Manokaran et al., 2008).

Family Verbenaceae comprises a multitude of species with medicinal and ornamental uses, which occurs in virtually all terrestrial ecosystems. Several members of the family Verbenaceae are considered as medicinal and are used from time immemorial. Rahmatullah (2011) critically reviewed the folk medicinal uses of Verbenaceae family plants in Bangladesh.

The antimicrobial activities of members of Verbenaceae are well reported like *Lippia palmeri* (Ortega-Nieblas *et al.*, 2011); *Clerodendrum viscosum* (Oly *et al.*, 2011); *Vitex negundo* (Mahmud *et al.*, 2009); *Lippia triplinervis* (Leitão *et al.*, 2011). However, there exists a lacuna of knowledge with regard to antifungal activity. However, traditional ayurvedic healers are using a wide ambit of plants in the family to treat various ailments. Thus scientific validation of existing knowledge is indispensible for developing new drug combinations and treatment modalities to combat fungal diseases, as fungal diseases are overwhelmingly emerging. The present study is mooted in this background.

^{*} To whom all correspondence should be addressed. E-mail: roji.cms@gmail.com

MATERIALS AND METHODS

Extract preparation

Leaves of 11 plants of Verbenaceae were collected from different regions in the State of Kerala. The taxonomic identification was performed by using standard taxonomic keys and expert consultations. Herbarium was also prepared and voucher numbers and the codes are assigned. The collected plants are dried under shade, crushed and subject to soxhlet extraction with petroleum ether and methanol. The extract was filtered and concentrated.

Antifungal susceptibility testing Fungal isolates

The test species used for this investigation are *Aspergillus flavus*, *A. fumigatus*, *A. niger*, *Penicillium* sp., *Verticillium* sp., *Curvularia* sp., *Mucor* sp. and *Fusarium* sp. The fungi were maintained on Sabouraud Dextrose Agar (SDA) slants at 10°C and subcultured for the study. **Agar well diffusion bioassay**

The antifungal activity of the extracts was determined by using the agar well diffusion assay as depicted by Patel and Trivedi (1962) and Holder and Boyce (1994) with some modifications. SDA plates were prepared and a well of 5mm diameter was bored at the middle of the plate aseptically. Then spore suspension of the fungi to be tested was prepared (0.2-2.5 x 105 CFU/mL) and swabbed on to the agar surface. 100 µl of extract was added to the well and incubated. The respective solvents used for extraction was (petroleum ether or methanol) used as a negative control. Four plates were prepared for each test and were incubated at room temperature. The antifungal activity was taken on the basis of diameter of zone of inhibition around the well, which was measured after 4-8 days of incubation and the mean value along with standard deviation of the readings are presented.

RESULTS AND DISCUSSION

Since antiquity, botanicals have provided a wide spectrum of compounds of healing properties, for several ailments. In recent years, antimicrobial properties of plant extracts are reporting incessantly throughout the world. The quantum of medicinal plants and herbal medicinal products used worldwide has climbed up

o. Fusarium sp.	- 26.72±0.71 26.72±0.71 01 29.91±2.09 19 17.19±0.11 04 12.82±0.13 37 27.62±0.83 11 32.00±1.39 11 15.48±0.91 - -
Mucor st	- - 36.76±1. 16.67±0. 17.79±0. 20.18±0. 23.38±0. 13.18±0. -
Penicillium sp.	13 ± 2.89 17.02 ± 0.87 27.77 ± 0.17 14.75 ± 0.11 16.80 ± 0.88 14.28 ± 0.81 19.81 ± 0.09 - 12.11 ± 0.88 12.68 ± 0.11 14.91 ± 0.18
Curvularia sp.	20.11 ± 0.11 - 18.85\pm0.88 14.11\pm0.09 12.80\pm0.18 16.00\pm0.10 15.00\pm0.02 12.95\pm0.02 12.95\pm0.02 13.71\pm0.71 24.66\pm0.49 -
Verticillium sp.	17.25±0.2 22.11±3.12 14.68±0.19 - 12.18±0.05 - 14.11±0.17 - - 17.78±0.33
A. fumigatus	- 24.00±1.81 19.71±2.01 19.29±0.99 24.49±2.77 19.98±1.31 - 17.11±1.37
A. niger	17.21±2.10 20.11±1.11 28.67±3.18 18.00±1.09 31.00±1.61 25.71±1.16 29.00±1.69 -
A. flavus	15±0.1 - - 33.01±0.89 24.26±3.21 30.67±4.16 26.92±2.13 32.11±1.18 - - - -
Plants tested	P. serratifolia L. nodiflora V. trifolia V. negundo var. negundo V. negundo var. pubescens V. altissima V. leucoxylon V. pinnata S. jamaicensis C. inermis C. serratum
S.No.	11

Table 1. Sensitivity pattern of petroleum ether extracts (mm)

dramatically in the last decades (Reich and Schibli, 2006). The Family Verbenaceae comprises several species with a myriad of characteristics and applications which occurs in virtually all terrestrial ecosystems. In folkloric medicine, various parts of the plants are used to rectify several disease conditions (Cowan, 1999; Burkill, 2000; Das *et al.*,

2003 and Edeoga et al., 2005). The antifungal effects of P. serratifolia, L. nodiflora, V. trifolia, V. negundo var. negundo, V. negundo var. pubescens, V. altissima, V. leucoxylon, V. pinnata, S. jamaicensis, C. inermis and C. serratum against fungi such as Aspergillus flavus, A. fumigatus, A. niger, Penicillium sp., Verticillium sp., Curvularia sp., Mucor sp. and Fusarium sp. were evaluated. In an overall analysis, all of the tested plants were inhibitory to many of the tested fungi. Interestingly, both of the extracts (petroleum ether and methanolic) showed appreciable antifungal effects, though the phytochemical composition was dissimilar.

The petroleum ether extract of P. serratifolia was more effective against Curvularia sp. (20.11 ± 0.11) followed by *Verticillium* sp. (17.25 ± 0.2) . On the other hand, the methanolic extract of P. serratifolia exhibited more effect against Penicillium sp. (16.11+0.13) (Table 1 and 2). The obtained results are in tune with the available results. The antimicrobial activity of P. serratifolia was well established (Rajendran and Basha, 2010; Singh, 2011). Several phytochemicals from both leaves and roots, are reported from P. serratifolia like 1HCycloprop [e]azulen -7-ol, decahydro-1,1,7-trimethyl-4methylene-, [1ar-(1aà,4aà,7á,7aá,7bà)]- (2.98%), 2-Furancarboxaldehyde, 5-(hydroxymethyl)- (2.44%), 2-Hydroxy-3-methylbenzaldehyde (6.39%), :2s,6s-2,6,8,8-Tetramethyltricyclo [5.2.2.0(1,6)] undecan -2ol (6.35 %), Benzofuran, 2,3-dihydro- (29.94 %), Glycerin (1.14%), n-Hexadecanoic acid (13.94%), 2-Propenoic acid and 3-(4-methoxyphenyl)-(13.84%) (Singh et al., 2011). The obtained antifungal activity is the result of synergestic action of all these active components.

L. nodiflora petroleum ether extract was also promising in controlling the tested fungi, *Fusarium* sp. (26.72 ± 0.71) , *Verticillium* sp. (22.11 ± 3.12) while the methanolic extract was effective against the genus *Aspergillus* only. Durairaj *et al.* (2007) critically studied the antimicrobial and lipid peroxide scavenging activity

	ucor sp. Fusarium sp.	I		± 0.18 29.06±0.11	13 ± 1.29 18.06 ±0.89	$.01\pm1.16$ 14± 0.03			.18±2.87 -	13.01 ± 0.08	19.81 ± 2.05	± 0.01 16.84±2.13
	Penicillium sp. Mı	16.11±0.13 -		25.79±0.19 35	17.77±1.61 15	22.18±2.17 18		33.72±1.75 -	19.18±0.86 13	13.09±1.04 -	- 13.89	16.91±0.15 14
xtracts (mm)	Curvularia sp.	ı		15.09 ± 2.18	15.08 ± 0.15	14.91 ± 1.93	16.72 ± 0.78		18.77 ± 1.04	18.70 ± 1.08		13.17±0.10
n of methanolic ey	Verticillium sp.	12.00 ± 0.08		17.79 ± 1.34	ı		13.73 ± 0.43		14.76 ± 0.59	ı	14 ± 0.24	1
asitivity patterr	A. fumigatus	14.11 ± 0.18	14.00 ± 0.17	25.77 ± 0.19	20.16 ± 1.11	ı	25.17 ± 3.29	23.89 ± 2.08	ı	ı	16.44 ± 0.34	
Table 2. Sei	A. niger	I	18.01 ± 2.07	31.07 ± 3.18	19.11 ± 0.14	33.01 ± 2.90	23.14 ± 0.19	18.12 ± 1.09	ı	ı	16 ± 0.07	1
	A. flavus		16.11 ± 1.87	26.71 ± 0.59	25.43 ± 1.20	25.43 ± 1.32	26± 2.27	31 ± 3.11	ı	,	19.80 ± 1.05	1
	Plants tested	P. serratifolia	L. nodiflora	V. trifolia	V. negundo var. negundo	V. negundo var. pubescens	V. altissima	V. leucoxylon	V. pinnata	S. jamaicensis	C. inermis	C. serratum
	S.No.	1.	5.	3.	4.	5.	6.	7.	8.	9.	10.	11

of *Lippia nodiflora* with significant outputs. The present results also supported the findings. Moreover, other potentials of the plant such as hepatoprotective activity were also well accomplished (Naik *et al.*, 2012).

The genus *Vitex* showed copious antifungal activity irrespective of petroleum ether and methanol extracts. The petroleum ether extract of the *Vitex* sp. *V. trifolia, V. negundo* var. *pubescens* and *V. leucoxylon* demoed notable antifungal effect against all the tested fungi. *V. trifolia* demonstrated significant antifungal effects towards *Mucor* sp. (36.76 ± 1.01) followed by *A. flavus* (33.01+0.89) and *Fusarium* sp. (29.91 ± 2.09) . *V. negundo* var. *pubescens* showed commendable activity against *A. flavus* (30.67 ± 4.16) and *A. niger* (31 ± 1.61) . *V. leucoxylon* was more active against *A. flavus* (31.11 ± 1.18) and *Fusarium* sp. (32 ± 1.39) .

A substantial antifungal activity was observed in the case of petroleum ether extract of *V. altissima* against *Fusarium* sp. (27.62 ± 0.83) followed by *A. flavus* (26.92 ± 2.13) and *A. niger* (25.71 ± 1.16) . The methanolic extract of *V. altissima* also showed activity against five of the tested fungi with high sensitivity towards *A. flavus* (26 ± 2.27) and *A. niger* (23.14 ± 0.19) .

S. jamaicensis petroleum ether extract was effective against only two of the tested fungi such as Curvularia sp. (13.71±0.71) and Penicillium sp. (12.11+0.88). On the other hand, Curvularia sp. (18.70±1.08), Penicillium sp. (13.09±1.04) and Fusarium sp. (13.01+0.08) was sensitive towards methanolic extract. The widespread use of S. jamaicensis in the treatment of a number of diseases is documented from time immemorial. The antimicrobial activity and the presence of phytochemicals such as phenolic compound, tannin, saponin, terpenoids and flavonoid was reported earlier (Putera, 2010) with notable variation in activity of root and leaf extracts. The current observation also referred the same, though antifungal reports of the plant are less.

C. inermis petroleum ether and methanolic extracts were effective against six of the tested fungi with notable variations. *C. serratum* petroleum ether extract was effective against three while methanolic extract was effective against four of the tested fungi. *C. inerme* possess antifungal potential irrespective of extracting solvents. The antifungal activity of *C. inerme* was thoroughly

		-	Table 3. Sensi	tivity pattern of	f essential oil of V	itex sp. (mm)			
S.No.	Plants tested	A. flavus	A. niger	A. fumigatus	Verticillium sp.	Curvularia sp.	Penicillium sp.	Mucor sp.	Fusarium sp.
1.	V. trifolia	30.11 ± 2.19	33.04± 0.21	25.90± 1.91	15.08 ± 0.07		16.73 ± 0.86	28.07± 1.98	26.79± 2.07
5.	V. negundo var. negundo	22.18 ± 3.29	16.07 ± 1.05	21.09±0.98	ı	13.17 ± 0.86	18	17.50 ± 0.67	
з.	V. altissima	25.27 ± 0.18	20 ± 0.15	19.16 ± 0.64	ı	12	ı	1	$12.0.16\pm0.08$
4.	V. leucoxylon	28.16 ± 1.79	24.17 ± 2.06	21.06 ± 0.13	12.90 ± 0.30	I	15.05 ± 0.17	ı	I

explored (Anitha and Kannan, 2006). In Indian systems of medicine, *C. inerme* are used for treating fever, cough, skin rashes and boils, and also used to treat umbilical cord infection and for cleaning the uterus in local medicine (Nadkarni, 1976). Tharmaraj *et al.* (2012) highlighted the phytochemical and bioefficacy potential of the plant.

In an overall analysis, the genus *Vitex* showed prominent sensitivity pattern against all of the tested fungi. Thus, an attempt was also done to evaluate the antifungal activity of essential oil of the genus. The results were summarized in Table 3. This clearly demonstrated that *V. trifolia* was active against seven of the tested fungi followed by *Vitex negundo* var. *negundo* (6 species), *V. altissima* and *V. lecucoxylon* (five species each).

The genus Vitex (V. trifolia, V. negundo var. negundo, V. negundo var. pubescens, V. altissima, V. leucoxylon, V. pinnata) is well known for a cornucopia of treatment attributes (Padmalatha *et al.*, 2009). An array of phytochemicals are also detected and purified from the plant by different methods (Meena *et al.*, 2011). The present results also inferred the widely accepted notion about the plant. Concerted efforts to characterize and evaluate antimicrobial effects were also noticed in concordance with the available results (Srinivas *et al.*, 2010; Panda *et al.*, 2011; Sathish *et al.*, 2012; Natheer *et al.*, 2012).

C. serratum also showed commendable antifungal activity in the present investigation. Ethno-medicinal grandness of the plant has been reported in various indigenous systems of medicines like Ayurveda, Siddha and Unani for the treatment of various grievous diseases such as syphilis, typhoid, cancer, jaundice and hypertension (Shrivastava and Patel, 2007). The plant was rich in D-mannitol, hispidulin, cleroflavone, apigenin, scutellarein, serratagenic acid, acteoside, verbascoside, oleanolic acid, clerodermic acid, γ -sitosterol, β -sitosterol, cholestanol, clerosterol, campesterol and 24-ethyl cholesterol which paves the way for the plants antimicrobial effects (Singh *et al.*, 2012).

CONCLUSION

Establishing the antifungal activity of these plants will impart the systematic scientific probe of indigenous medicinal plants in hunting of novel drugs. It is concluded that P. serratifolia, L. nodiflora, V. trifolia, V. negundo var. negundo, V. negundo var. pubescens, V. altissima, V. leucoxylon, V. pinnata, S. jamaicensis, C. inermis and C. serratum do possess antifungal properties and that this activity is largely dependent on phytochemicals. However, further studies are required in order to gain more lucidity as to the specificity and biochemical mechanisms creditworthy for the antifungal properties of these plants. Fungal diseases are emerging and there is an urgent need in developing newer antifungals to combat fungal diseases. The phytochemical compounds should be subjected to animal and human trials to ascertain effectiveness, toxicity and effects on normal microbiota. More systematic exploration and interpretation would be facilitated to achieve targets.

REFERENCES

- 1. Anitha, R. and Kannan, P., Antifungal activity of *Clerodendrum inerme* (L). and *Clerodendrum phlomidis* (L). *Turk. J. Biol.* 2006; **30**:139-142.
- Burkill, H.M., Useful Plants of West Tropical Africa. 2nd ed. Vol. 5. Royal Botanic Garden Kew. 2000; 272-275.
- Cowan, M.M., Plant products as antimicrobial agents. *Clinical Microbiology Reviews*. 1999; 12(4): 564-582.
- Das, S., Dash, S.K. and Padhy, S.N., Ethnomedicinal informations from Orissa state, India: A Review. J. Hum. Ecol. 2003; 14(3):165-227.
- Durairaj, A.K., Vaiyapuri, T.S., Mazumder, U.K. and Gupta, M., Antimicrobial and lipid peroxide scavenging activity of *Lippia nodiflora* (Verbenaceae). *Pharmacologyonline* 2007; 3: 177-189.
- Edeoga, H.O., Okwu, D. E. and Mbaebie, B.O., Phytochemical constituents of some Nigerian medicinal plants. *African Journal of Biotechnology*. 2005; 4(7):685-688.
- Haldar, P. K., Kar, B., Bala, A., Bhattacharya, S. and Mazumder, U. K., Antitumor activity of Sansevieria roxburghiana rhizome against Ehrlich ascites carcinoma in mice. *Pharmaceutical Biology*. 2010; 48(12):1337-1343.
- Holder, I.A. and Boyce, S.T., Agar well diffusion assay testing of bacterial susceptibility to various antimicrobials in concentrations nontoxic for human cells in culture. *Burns*. 1994; 20:426-429.
- 9. Leitão, S.G., Damasceno, J.P.L., Martini, M.G., Miranda, S.N., Neufeld, P.M., Salimena, F.R.

and Bizzo, H.R., Analysis of the chemical composition and antimicrobial activity of the essential oil from *Lippia triplinervis* Gardner (Verbenaceae). *J. Essent. Oil Res.* 2011; 23: (September/October 2011).

- Mahmud, S., Shareef, H., Farrukh, U., Kamil, A. and Rizwani, G.H., Antifungal activities of *Vitex negundo* Linn. *Pak. J. Bot.* 2009; **41**(4): 1941-1943.
- Manokaran, S., Jaswanth, A., Sengottuvelu, S., Nandhakumar, J., Duraisamy, R., Karthikeyan D. and Mallegaswari, R., Hepatoprotective activity of Aerva lanata Linn. against paracetamol induced hepatotoxicity in rats. *Research J. Pharm. and Tech.* 2008; 1(4): 398-400.
- Meena, A.K., Niranjan, U.S., Rao, M.M., Padhi, M.M. and Babu, R., A review of the important chemical constituents and medicinal uses of *Vitex* genus. *Journal of Traditional Medicines*. 2011; 6(2): 54-60.
- Nadkarni, A.K., *Indian Materia Medica*. Popular Prakashan.Bombay, 1976.
- Naik, N.D., Naadella, B.C., Reddy, R. and Suresh, B., Evaluation of hepatoprotective activity of *Phyla nodiflora L.* against CCl4 induced hepatotoxicity. *International Journal of Innovative Drug Discovery.* 2012; 2(1):16-21.
- Namita, P. and Mukesh, R., Medicinal plants used as antimicrobial agents: A review. *International Research Journal of Pharmacy*. 2012; 3(1): 31-40.
- 16. Natheer, S.E., Sekar, C., Amutharaj, P., Rahman, S.A. and Khan, K.F., Evaluation of antibacterial activity of *Morinda citrifolia*, *Vitex trifolia* and *Chromolaena odorata*. *African Journal of Pharmacy and Pharmacology*. 2012; **6**(11): 783-788.
- Oly, W.T., Islam, W., Hasan, P. and Parween, S., Antimicrobial activity of *Clerodendrum* viscosum vent. (Verbenaceae). *Int. J. Agric. Biol.* 2011; 13: 222-226.
- Ortega-Nieblas, M.M., Robles-Burgueño, M.R., Acedo-Félix, E., González-León, A., Morales-Trejo, A. and Vázquez-Moreno, L., Chemical composition and antimicrobial activity of Oregano (*Lippia palmeri* s. wats) essential oil. *Rev. Fitotec. Mex.* 2011; 34(1):11-17.
- Padmalatha, K., Jayaram, K., Raju, N.L., Prasad, M.N.V. and Arora, R., Ethnopharmacological and biotechnological significance of *Vitex*. *Bioremediation Biodiversity & Bioavailability*. 2009; 3(1):6-14.
- Panda, S.K., Padhi, L., Mohanty, G., Sahoo, G. and Dutta, S.S., Phytochemical analysis and Antimicrobial activities of different plant parts of *Vitex negundo* L. *Journal of Pharmacy Research.* 2011; 4(9): 3184-3189.

- Patel. R.P. and Trivedi. B.M., The *in vitro* antibacterial activity of some medicinal oils. *Ind. J. Med. Res.* 1962; 60:211-222.
- 22. Putera, K.A.S.I., Antimicrobial activity and cytotoxic effects of Stachytarpheta jamaicensis (L.) Vahl crude plant extracts. A dissertation submitted in partial fulfillment of the requirements for the award of the degree of Master of Science (Biotechnology). Universiti Teknologi, Malaysia, 2010.
- 23. Rahmatullah, M., Folk medicinal uses of Verbenaceae family plants in Bangladesh. *African Journal of Traditional, Complementary and Alternative medicines (AJTCAM).* 2011; **8**(5S).
- Rajendran, R. and Basha, D.N., Antimicrobial activity of crude extracts and fractions of *Premna* serratifolia Linn., root. *Medicinal Plants-Int. J. Phytomed. Rel. Indus.* 2010; 2(1): 33-38.
- 25. Rao, K.B., Kesavulu, M.M. and Apparao, C., Antihyperglycemic activity of Momordica cymbalaria in alloxan diabetic rats. *Journal of Ethnopharmacology*. 2001; **78**(1):67-71.
- 26. Reich, E., and Schibli, A., High-Performance Thin-Layer Chromatography for the Analysis of Medicinal Plants, Thieme, New York, 2006.
- Sathish, S.S., Janakiraman, N. and Johnson, M., Phytochemical Analysis of Vitex altissima L. using UV-VIS, FTIR and GC-MS. International Journal of Pharmaceutical Sciences and Drug Research. 2012; 4(1): 56-62.
- Shrivastava, N. and Patel, T., *Clerodendrum* and Heathcare: An Overview. *Medicinal and Aromatic Plant Science and Biotechnology*. 2007; 1: 142-150.
- Singh, C.R., Antimicrobial effect of callus and natural plant extracts of *Premna serratifolia* L. *Intl. J. Biomed. Pharma*. 2011; 2(1):17-20.
- Singh, C.R., Nelson, R., Krishnan, P.M. and Pargavi, B., Identification of volatile constituents from Premna serratifolia L.through GC-MS. *International Journal of PharmTech Research*. 2011; 3(2): 1050-1058.
- Singh, M.K., Khare, G., Iyer, S.K., Sharwan, G. and Tripathi, D. K., *Clerodendrum serratum*: A clinical approach. *Journal of Applied Pharmaceutical Science*. 2012; 2(2):11-15.
- 32. Srinivas, P., Reddy, R., Pallav, P., Suresh, A. and Praveen, V., Screening for antimicrobial properties of *Vitex negundo*. *L* from rural areas of Warangal dist/A.P. India. *International Journal of Pharma and Bio Sciences*. 2010; **1**(4): B26-38.
- Tharmaraj, R.J.J.M., Antonisamy, J.M., Solomon, J. and Revathi I., Phytochemical and bioefficacy studies on *Clerodendron inerme* Gaertn. Asian Pacific Journal of Tropical Biomedicine. 2012; 1-3.